

RESEARCH

Open Access



Tracheal agenesis versus tracheal atresia: anatomical conditions, pathomechanisms and causes with a possible link to a novel *MAPK11* variant in one case

Mateja Pfeifer¹ , Helga Rehder^{1,10*} , Maria Gerykova Bujalkova¹ , Christine Bartsch^{2,3} , Barbara Fritz⁴, Cordula Knopp⁵, Björn Beckers⁶, Frank Dohle⁷, Matthias Meyer-Wittkopf⁸, Roland Axt-Fliedner⁹, Alexander V. Beribisky¹, Manuel Hofer¹, Franco Laccone¹ and Katharina Schoner¹⁰

Abstract

Background In this study we aimed to describe the morphological and pathogenetic differences between tracheal agenesis and tracheal atresia, which are not clearly distinguished from each other in the literature, and to contribute thereby to the understanding and management of these conditions. Both tracheal agenesis and tracheal atresia represent rare disorders of still unknown aetiology that cannot be detected by prenatal ultrasound. If the affected fetuses survive until birth these conditions result in respiratory failure and in futile attempts to rescue the infant's life.

Results Autopsies and genetic analyses, including singleton or trio exome sequencing, were performed on five neonates/foetuses with tracheal agenesis and three foetuses with tracheal atresia. Tracheal agenesis was characterized by absence of the sublaryngeal trachea and presence of a bronchooesophageal fistula and by pulmonary isomerism and occurred as an isolated malformation complex or as part of a VACTERL association. Special findings were an additional so-called 'pig bronchus' and a first case of tracheal agenesis with sirenomelia. Tracheal atresia presenting with partial obliteration of its lumen and persistence of a fibromuscular streak resulted in CHAOS. This condition was associated with normal lung lobulation and single, non-VACTERL type malformations. Trio ES revealed a novel variant of *MAPK11* in one tracheal agenesis case. Its involvement in tracheooesophageal malformation is herein discussed, but remains hypothetical.

Conclusion Tracheal agenesis and tracheal atresia represent different disease entities in terms of morphology, pathogenesis and accompanying anomalies due to a primary developmental and secondary disruptive possibly vascular disturbance, respectively.

Keywords Tracheal agenesis, Tracheal atresia, Congenital high airway obstruction sequence (CHAOS), VACTERL association, Sirenomelia, *MAPK11* variant, p38beta

*Correspondence:

Helga Rehder

helga.rehder@alumni-club.meduniwien.ac.at

Full list of author information is available at the end of the article



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

The terms tracheal agenesis (TAG) and tracheal atresia (TAT) are not clearly distinguished from each other in the literature and are commonly referred to as TA. Tracheal agenesis is defined as absence of the sublaryngeal trachea [1–3] whereas tracheal atresia describes the congenital obliteration of the tracheal lumen leaving a fibromuscular streak without cartilages between the nonobliterated segments. TAG and TAT are rare life-threatening conditions with a prevalence rate of less than 1:50,000 and a male preponderance of m:f = 1:0.5 [4, 5]. TAG is most commonly associated with a narrow tracheo- or bronchooesophageal fistula (TOF/BOF), not avoiding polyhydramnios and fatal respiratory distress at birth but allowing drainage of lung fluid, thus preventing in utero development of pulmonary hyperplasia, consecutive diaphragmatic eversion and foetal hydrops, a condition referred to as “congenital high airway obstruction sequence”. CHAOS is not associated with a TOF or BOF [6, 7].

Concerning the position of the TOF/BOF, Floyd et al. [8] devised a classification, distinguishing between partial TAG with upper tracheal agenesis and with the lower trachea proximally communicating with the oesophagus (type 1), TAG with fusion of both main bronchi before communicating with the oesophagus (type 2) and TAG showing separate outlets of the main bronchi from the oesophagus (type 3). The frequency distribution recorded in 137 affected individuals was 33.6%, 49.6% and 16.8%, respectively [5]. The Faro classification on 40 tracheal agenesis cases from the literature includes 6 cases corresponding to TAT [9]. His types B, C and E correspond to Floyd's type 3, 2 and 1.

In approximately 50% of cases, TAG is associated with prematurity and polyhydramnios [2]. As tracheal abnormalities cannot be visualised by ultrasound, it was recommended to perform a prenatal MRI in the presence of unexplained polyhydramnios and to consider tracheal or laryngeal atresia as possible cause of pulmonary hyperplasia [3, 10, 11]. The accompanying anomalies reported in 80–94% of the TAG cases mainly concern features of the VACTERL association [1–3, 5]. Evans et al. [1] assigned their pattern in 80 TAG cases to four subgroups according to the number of developmental fields involved. Radial limb defects were always right-sided. Lungs were always bilaterally bilobed or trilobed.

Perinatal intervention via “extrauterine intrapartum therapy” (EXIT) allows for targeted oesophageal intubation and life-saving postnatal oxygenation [5, 10]. Attempts at surgical neotrachealization of the oesophagus by vertical division or by internal or external stenting and oesophageal reconstruction have thus far been

unable to provide long-term survival without neurological impairment [12, 13].

Tracheal development

The etiology of TAG/TAT remains controversial. TAG has been induced by exposing rat embryos to adriamycin [14]. However, multiple genes have been described as being involved in tracheoesophageal development [15]. It was shown in mouse and *Xenopus* that preceding patterning of a dorsal and ventral foregut domain is a precondition for proper tracheoesophageal separation from the foregut, and that separation occurs by compartmentalization with locally increased proliferation of epithelial and mesenchymal cells along the lateral midlines [16, 17]. Patterning is established by expression of the transcription regulator Sox2 in the dorsal endoderm and of the thyroid transcription factor Nkx2-1 in the ventral endoderm [18–21]. Reduction of Sox2 leads to loss of dorsal patterning and—depending on the threshold levels—to oesophageal agenesis, and reduction of Nkx2-1 to loss of ventral patterning and thus to tracheal agenesis. In either condition the foregut remains a single tube with either oesophageal or tracheal wall structures and with connections to the stomach and lungs through a BOF or TOF. However, different signaling pathways and transcription factors establish and regulate SOX2 and NKX2-1 expression indicating involvement of numerous different genes in tracheoesophageal separation, e.g., *BMP4* and *SMAD1*, downregulating SOX2 in the ventral foregut epithelium and Wnt2/2b inducing Nkx2-1+ respiratory progenitor ligands in the surrounding splanchnic mesoderm. In addition the ligands Shh, Ihh, and Wnt, whose expression is stimulated by retinoic acid (RA), are involved in the differentiation of splanchnic mesoderm into tracheal cartilage or oesophageal smooth muscle. It was also shown, that loss of RA and Hh, WNT2/2b- or BMP4- signalling results in loss of the tracheal phenotype [15–17, 22, 23]. However, no sequencing analyses have thus far been reported in affected humans.

We examined 5 cases of TAG and 3 cases of TAT in order to distinguish the anatomical structures and associations and discuss the possible causes and pathomechanisms of these conditions.

Methods

Foetal autopsies, X-ray or MRI and thorough photographic documentation were performed on 3 neonates and 2 fetuses with TAG and 3 fetuses with TAT. Cases 1 and 3–8 had been sent for autopsy from different hospitals in Germany. Case 2 was derived from the Institute of Forensic Pathology in Zurich.

Cytogenetic analyses were performed according to standard procedures. Molecular analyses were performed

on DNA derived from umbilical cord and foetal muscle (cases 1, 7–8), chorionic villi (cases 3 and 5), cultured amnion cells (case 4) and from parental blood (cases 1 and 5). The DNA isolated from different formalin fixed paraffin embedded (FFPE) probes of cases 2 and 6 was highly fragmented and not eligible for molecular testing. Array-CGH was performed using 4×44 K Human Genome CGH microarrays (Oxford Gene Technology, Begbroke, Oxfordshire, UK) and SNP arrays (GeneChip®, Affymetrix). Data were analysed using Agilent CytoGenomics software (Agilent Technologies, Inc., Santa Clara, CA). The probe sequence annotation was based on NCBI Build GRCh37 (hg19) of the human genome.

In cases 3, 4, 7 and 8 a singleton exome-sequencing (ES) was performed and in cases 1 and 5 a trio ES was performed. DNA samples were prepared following the workflow of the TruSeq Exome Library Kit (Illumina) or Twist Comprehensive Exome Panel (Twist Bioscience) for enrichment of exonic regions. The final library was paired-end sequenced on an Illumina NextSeq500 or NextSeq 2000 sequencer. Variants were evaluated using the program VarSeq Golden Helix® (Bozen, Montana) and were classified according to the American College of Medical Genetics and Genomics (ACMG) guidelines [24]. A copy number variation (CNV) analysis of ES data by a module from VarSeq software was performed. The potential effects of the Met109Leu variant in MAPK11 were analysed using a structure of this protein (PDB ID: 3GP0) [25] already determined by X-ray crystallography in PyMol [26].

Results

Clinical data of TAG cases 1–5 (s. Table 1)

In none of the cases tracheal agenesis had been recognized prenatally. The pregnancies had been uneventful in the neonates of cases 1–3 except for late polyhydramnios and premature rupture of membranes and for prenatally suspected ‘oesophageal atresia’ in case 3. Thus, the obstetricians were completely unaware of the condition. Any attempts to intubate the trachea or to find the trachea for tracheotomy failed. The neonates died 1–4 h postpartum due to insufficient oxygenation by high pressure ventilation through an endo-oesophageal tube. In the foetuses of cases 4 and 5, ultrasound diagnosis of the associated malformations led to termination of pregnancy.

Post-mortem examination of TAG cases 1–5 The foetuses/neonates showed an identical type of TAG with a sub-laryngeal blind pouch without cartilages, absence of the entire trachea and a tracheo-oesophageal fistula type 2 [8] (Fig. 1a, b, d–f). On histology the oesophagus showed a normal oesophageal wall without tracheal cartilages and with the mucous surface lined by a squamous epithelium (Fig. 1a inset). In case 3, TAG was associated with

an additional proximal BOF derived from the right upper lobe bronchus (Fig. 1b). A single tracheo-oesophageal tube was recognized by MRI in case 2 (Fig. 1c).

The associated anomalies diagnosed prenatally or at autopsy in cases 3 to 5 comprised vertebral, anal, cardiovascular, tracheo-oesophageal, renal and right-sided radial limb defects, (Fig. 3a–c), allowing diagnosis of a VACTERL association in cases 3–4 and, with respect to the sympodia in case 5, of a VACTERL variant (Fig. 2a–c). The TAG cases with VACTERL association as well as the isolated TAG in cases 1 and 2 were accompanied by either bilaterally two-lobed (cases 1 and 3–5) or three-lobed lungs with a right descending aorta in case 2. The sex ratio in cases 1–5 was m:f = 1:1.5.

Clinical data of TAT cases 6–8 (s. Table 1)

CHAOS was diagnosed in cases 7 and 8 at 20 and 23 week's gestation, while the hyperplastic lungs in case 6 had been interpreted as adenomatoid pulmonic hyperplasia at 19 weeks in 1994. Prenatal ultrasound revealed a transverse defect of the left hand in the foetus of case 8.

Post-mortem examination of TAT cases 6–8 displayed a long distance tracheal atresia leaving a narrow fibromuscular streak without recognizable lumen or glandular or cartilaginous components. The atretic segment was located in the cervical trachea in case 6 and in the thoracic trachea in cases 7–8 (Fig. 3a–c). All three foetuses showed airway distension distal to the obstruction with lung hyperplasia, inverted diaphragm and hydrops thus fulfilling the criteria of ‘CHAOS’ (Fig. 4a–d). Associated anomalies comprised double left ureter and an amniotic strangulation in case 6, agenesis of the corpus callosum in case 7, and a VSD and transverse defect of the left hand with the absence of carpals and metacarpals and with rudimentary phalanges within five finger buds in case 8 (Fig. 2d). Sex ratio in cases 6–8 was m:f = 1:2.

In all 8 cases, parental consanguinity and a positive family history were excluded.

Genetic analyses in cases 1–8 (Table 1)

Karyotyping and CNV analyses showed normal results. Singleton ES in cases 3, 4, 7 and 8 and trio ES in case 5 revealed no disease-associated variants. Trio exome sequencing in case 1 revealed a heterozygous missense variant in the *MAPK11* gene of de novo origin (NM_002751.7:c.325A>T, NP_002742.3:p.Met109Leu; hg19). The identified variant site lies in a region which is referred to as a crossover connection in related structures [27], and next to the ATP-binding pocket (Fig. 5a) of the protein. The apparently conservative Met109Leu substitution [28] results in the loss of two ion electron pairs of methionine's sulphur atom S_8 (Fig. 5b).

Table 1 Clinical data, morphological, cytogenetic and molecular findings

Features	Case 1	Case 2	Case 3	Case 4	Case 5	Case 6	Case 7	Case 8
Gravida/para	II/0	IV/I	II/I	II/0	I/0	ND	III/II	I/0
Gest. weeks/ outcome	3rd trimester (PROM)	liveborn (S)	3rd trimester (CS)	3rd trimester (TP)	2nd trimester (IUFD)	2nd trimester (TP)	2nd trimester (TP)	2nd trimester (TP)
Weight	2136 g	3000 g	1560 g	1020 g	21.8 g	350 g	362 g	630 g
Length (CHL)	49.3 cm	50 cm	41 cm	39 cm	11.3 cm	22.5 cm	24.3 cm	29.8 cm
Prenatal US	normal	normal	Possible oesophageal atresia, no TOF	VACTERL association	omphalocele sympodia	CCAM	CHAOS	CHAOS and defect of left hand
Polyhydram- nios	+	–	+	–	–	Oligohydr.	–	–
Hydrops	–	–	–	–	–	+	+	+
Tracheal mal- formation	TAG + BOF	TAG + BOF	TAG + BOFs	TAG + BOF	TAG + BOF	TAT cranial seg- ment	TAT caudal seg- ment	TAT caudal segment
Pulmonary hyperplasia, lung weight in grams (times the norm for GA)	– 32.5 (0.96)	– n.d	– 27.0 (0.89)	– 21.1 (0.83)	– 0.72 (0.2)	– 17.9 (2.5)	– 41.6 (5.8)	– 58.0 (5.5)
Lung lobes (right/left)	2/2	3/3	2/2	2/2	2/2	3/2	3/2	3/2
Disease entity/ associated malformations	isolated TAG complex	TAG complex with right descend. aorta	VACTERL including AA, cardiac VSD, TAG, OF and right RLRD	VACTERL incl. SVD, ARA, card.-vasc. PLSVC, ARSA, TAG, OF, R(A) D, and right RLRD	VACTERL variant incl. SVD, ARA, cardiac AVSD, TAG, OF, RA, right RLRD, AG, omphalocele and sympodia	CHAOS and double left ureter, strangulating amniotic band	CHAOS and agenesis of corpus cal- losum	CHAOS and VSD, transverse defect of the left hand
BOF/TOF with Floyd classif. ^a	2	2	2+ “bronchus suis”	2	2	NA	NA	NA
BOF/TOF with Faro classif. ^b	C	C	C+ “bronchus suis”	C	C	G	G	G
VACTERL Evans subgroup ^c	NA	NA	3	4	4	NA	NA	NA
Family history/ Consanguinity	–/–	–/–	–/–	–/–	–/–	–/–	–/–	–/–
Karyotype	46,XN	46,XN	46,XN	46,XN	46,XN	46,XN	46,XN	46,XN
CNV analysis (aCGH and/ or ES)	– (aCGH and ES)	ND	– (aCGH and ES)	– (ES only)	– (aCGH and ES)	ND	– (aCGH and ES)	– (aCGH and ES)
Singleton- or Trio-ES/ Result	Trio VUS in <i>MAPK11</i> (de novo)	ND	Singleton –	Singleton –	Trio –	ND	Singleton –	Singleton –

aCGH array comparative genomic hybridisation, A(R)A ano(rectal) atresia, AG absent external genitalia, ARSA aberrant right subclavian artery, AVSD atrioventricular septal defect (persistent AV canal), (B)OF (broncho)oesophageal fistula, CCAM congenital cystic adenomatoid malformation of lungs, CHL crown-heel length; CS caesarean section; ES exome sequencing; GA gestational age; IUFD intrauterine fetal demise; NA not applicable; ND not determined; PLSVC persistent left superior vena cava, PROM premature rupture of membranes, RA renal agenesis, R(A)D renal(a)dysplasia, RLRD radial (longitudinal) limb reduction defect, S spontaneous birth, SVD sacral vertebral defects, TAG tracheal agenesis, TAT tracheal atresia, TLRD transverse limb reduction defect, TP termination of pregnancy, US ultrasound, VSD ventricular septal defect, VUS variant of unknown significance

^a Floyd et al. [8]

^b Faro et al. [9]

^c Evans et al. [1]

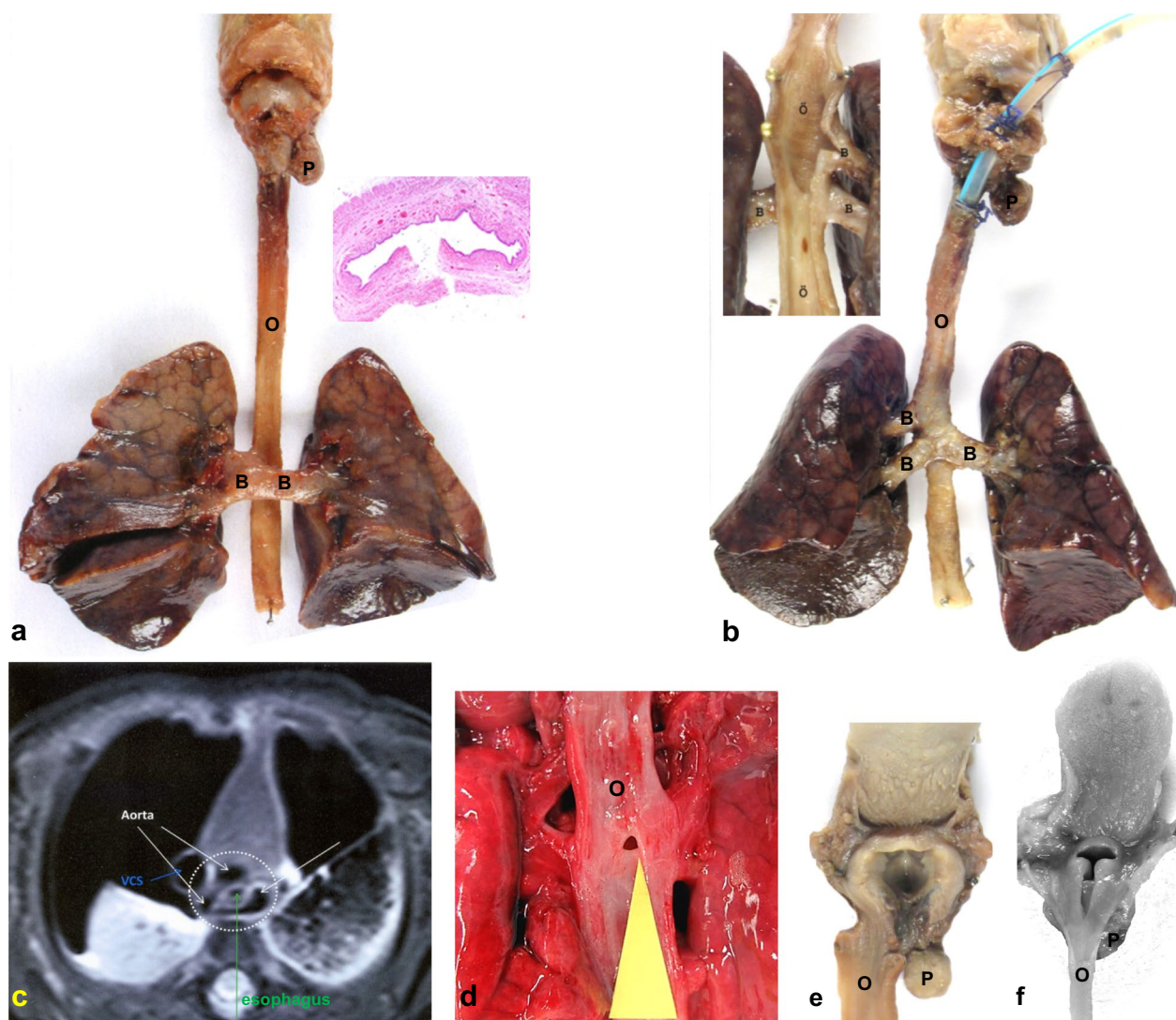


Fig. 1 Tracheal agenesis—Case 1: Ventral view of the respiratory tract showing defect of the entire trachea except for a cartilage-free sublaryngeal pouch (P), with fused main bronchi (B), connected to the oesophagus (O) by a broncho-oesophageal fistula (BOF—TAG Floyd type 2) and with bilaterally bilobed lungs. Transversal section plane (H&E, 2,5) of the oesophagus with normally structured wall and without any tracheal components on histology (inlay) (a). Case 3: Ventral view of the respiratory tract with an absent trachea, sublaryngeal pouch (P), bilaterally bilobed lungs and a BOF connecting the fused right main bronchus (B) and left lower lobe bronchus with the oesophagus (O) and a second separate BOF connecting the left upper lobe bronchus with the oesophagus. Dorsal view of the opened oesophagus (O) and BOFs (inlay) (b). Case 2: Post-mortem MRI, axial T2 weighted image (T2WI) showing aorta (white arrows), vena cava superior (VCS) (blue arrow), oesophagus (green arrow) and lack of a trachea (c). Dorsal view of the internal surface of the oesophagus showing the opening to the BEF of only 1.5 mm in diameter (yellow arrow) (d). Case 4: View of the opened larynx from dorsal with the oesophagus (O) laid aside showing hypoplasia of the epiglottis and the tracheal pouch (P) dorsally opened (e). Case 5: Dorsal view of the larynx with a hypoplastic epiglottis, an intact position of the oesophagus (O) and a tracheal pouch (P) behind (f)

Discussion

Tracheal agenesis

In cases 1–5 TAG was characterized by a blind sublaryngeal pouch, most likely representing the former laryngotracheal diverticulum, by absence of a trachea, by a BOF of Floyd's type 2 or Faro's type C and by pulmonary isomerism. However, in case 3 there was an

additional proximal BOF, connecting the right upper lung lobe bronchus with the oesophagus. A separate upper 'tracheal bronchus, a so called 'pig bronchus (bronchus suis)', occurs with an incidence of 0.2% in humans [29]. Based on the ~200 TAG-cases described thus far, it was observed twice, including our case [30, 31].

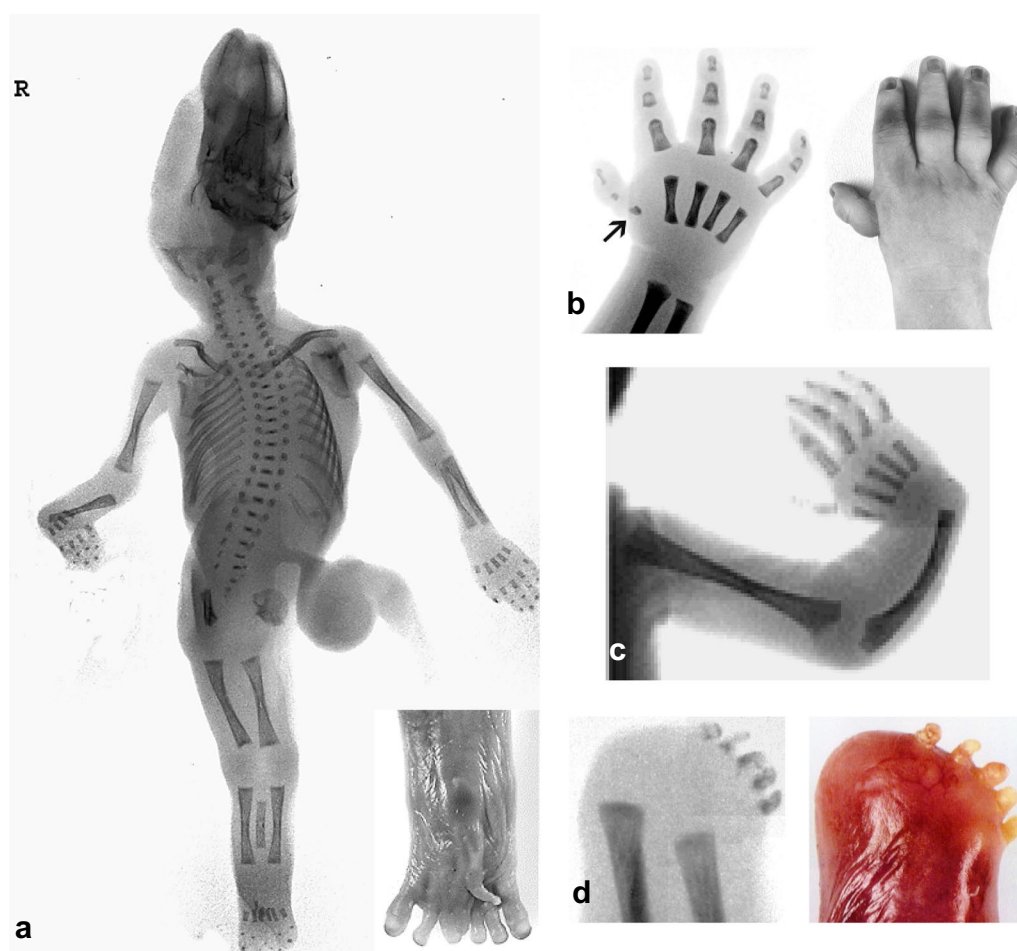


Fig. 2 Limb reduction defects Case 5: AP-X-ray of the entire foetus showing sirenomelia with two femora and tibiae, a single fused fibula and foot, and eight metatarsals and toes (toes 4 and 5 fused) as well as radial deviation of the right hand due to radial aplasia combined with a defect of the 1st metacarpal and the thumb, furthermore scoliosis of the spine and only 11 ribs (a). Case 3: Right hand displaying a rudimentary 1st metacarpal and hypoplastic thumb appendage (b). Case 4: Radial adduction of the right hand due to absence of the radius and bowing of the ulna (c). Case 8: End of the slightly shortened left forearm displaying a distal transverse reduction defect including absence of the hand and hand bones with rudimentary five phalanges within five finger buds (d)

Frequently associated malformations in TAG concern anomalies seen in the VACTERL association. According to Evans' subclassification, considering the number of developmental fields involved [1], our three TAG cases with VACTERL could be assigned to subgroups 3 (case 3) and 4 (cases 4 and 5), the latter representing a first observation of TAG associated with sirenomelia in terms of an extended developmental midline defect. We consider the 'associated' TAG to be part of the spectrum of possible VACTERL malformations, which is thus based on the same causes.

Tracheal atresia

Contrary to TAG, TAT in CHAOS cases 6–8 presented with a trachea that is distally properly divided into two

main bronchi, with accompanying non-VACTERL type malformations, with normal lung lobulation and with a sequence rather than an association. However, the trachea showed a long-distance obliteration of its lumen and lack of cartilages and mucous glands, leaving a residual fibromuscular cord between the non-obliterated segments. The obliteration affected either the cervical or the thoracic trachea. There was no TOF and thus no possibility for drainage of lung fluid, resulting in pulmonary hyperplasia and ultimately in CHAOS. Associated malformations included double ureters, absent corpus callosum, VSD and a terminal transverse LRD. These dissimilarities may indicate a differing etiology and pathogenesis of TAT compared with TAG. In this context the left-sided terminal transverse LRD in TAT as compared

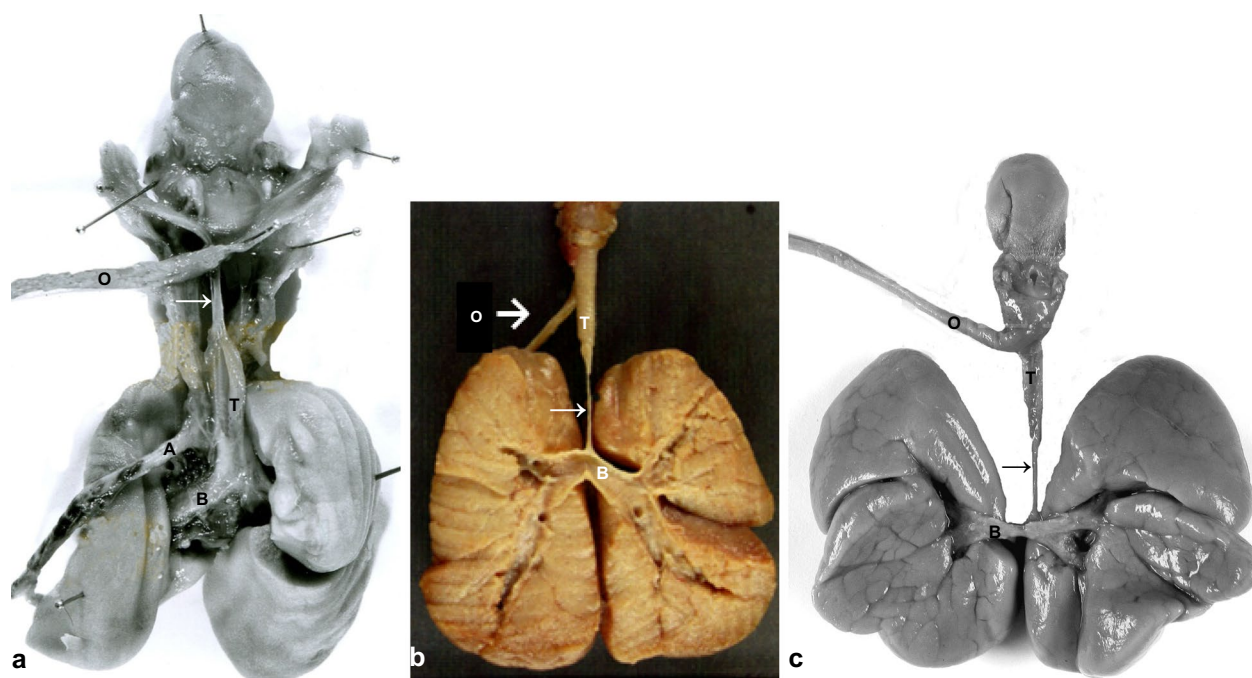


Fig. 3 Tracheal atresia Case 6: Dorsal view of the respiratory tract with oesophagus (O) and aorta (A) laid aside, showing atresia with a streak-like remnant (arrow) of the upper trachea (T), dilated main bronchi (B) and hyperplasia of the lungs (**a**). Case 7: Ventral view of the respiratory tract with the oesophagus (O) laid aside displaying atresia with a streak-like remnant of the lower trachea (arrow), dilatation of the bronchi and hyperplastic lungs shown on the sagittal cut surface (**b**). Case 8: Dorsal view of the respiratory tract with the oesophagus (O) laid aside, atresia of the lower trachea (arrow), dilatation of the bronchi and hyperplasia of the lungs (**c**)

to the VACTERL type right-sided longitudinal LRDs in TAG is of interest. Preaxial longitudinal LRDs are often syndromic or part of an association, whereas terminal transverse LRDs are not, with a few exceptions such as Adams-Oliver syndrome, which shows additional vascularization and coagulation disorders resulting, e.g., in characteristic scalp defects. Transverse LRDs are generally sporadic and are thought to arise from teratogenic or vascular disruptions [32–34]. A vascular disruption could also be a conceivable cause of TAT, insofar as the tracheal blood supply involves different vessels. While the cervical trachea is supplied by the inferior thyroid arteries and its distal end by the bronchial arteries, the blood supply to the thoracic segment is variable and can involve the intercostal, subclavian, internal thoracic or innominate arteries [35]. The pattern of blood supply matches the pattern of TAT. However, the abovementioned vessels only supply the lateral and anterior tracheal walls. The dorsal wall receives its blood solely through oesophageal vessels [31]. This could explain the persistence of a fibromuscular cord in the atretic portion of the trachea.

MAPK11 variant

In two cases each neither TAG nor TAT singleton ES revealed any pathogenic variants in OMIM-associated

or relevant candidate genes. While a trio ES in the sirenomelic VACTERL foetus excluded de novo variants compared to the parental exomes, case 1 with the isolated TAG presented with a novel heterozygous missense variant of unknown significance (VUS) c.325A > T (p.Met109Leu) in *MAPK11*. This variant is neither listed in the current international databases, including gnomAD, nor has *MAPK11* been linked to any disease in OMIM. However, the residue in question, Met109, concerns a highly conserved region in MAPK11 protein homologues [27, 28]. Although it is only involved in a few intramolecular contacts (namely with Ala155), its location in a region that corresponds to the crossover region in related structures, and next to the ATP-binding pocket of the protein makes this residue crucial for the function of MAPK11. The seemingly conservative Met109Leu substitution [28], results in the loss of two ion electron pairs of methionine's sulphur atom S_δ (Fig. 5b) which may affect proper ATP recognition by MAPK11 or distort its binding pocket, leading to compromised ATP hydrolysis and subsequently phosphorylation of downstream targets of MAPK11.

MAPK11 encodes the mitogen-activated protein kinase p38β. There are four p38 isoforms p38α–δ encoded by *MAPK14*, *MAPK11*, *MAPK12* and *MAPK13*. They are

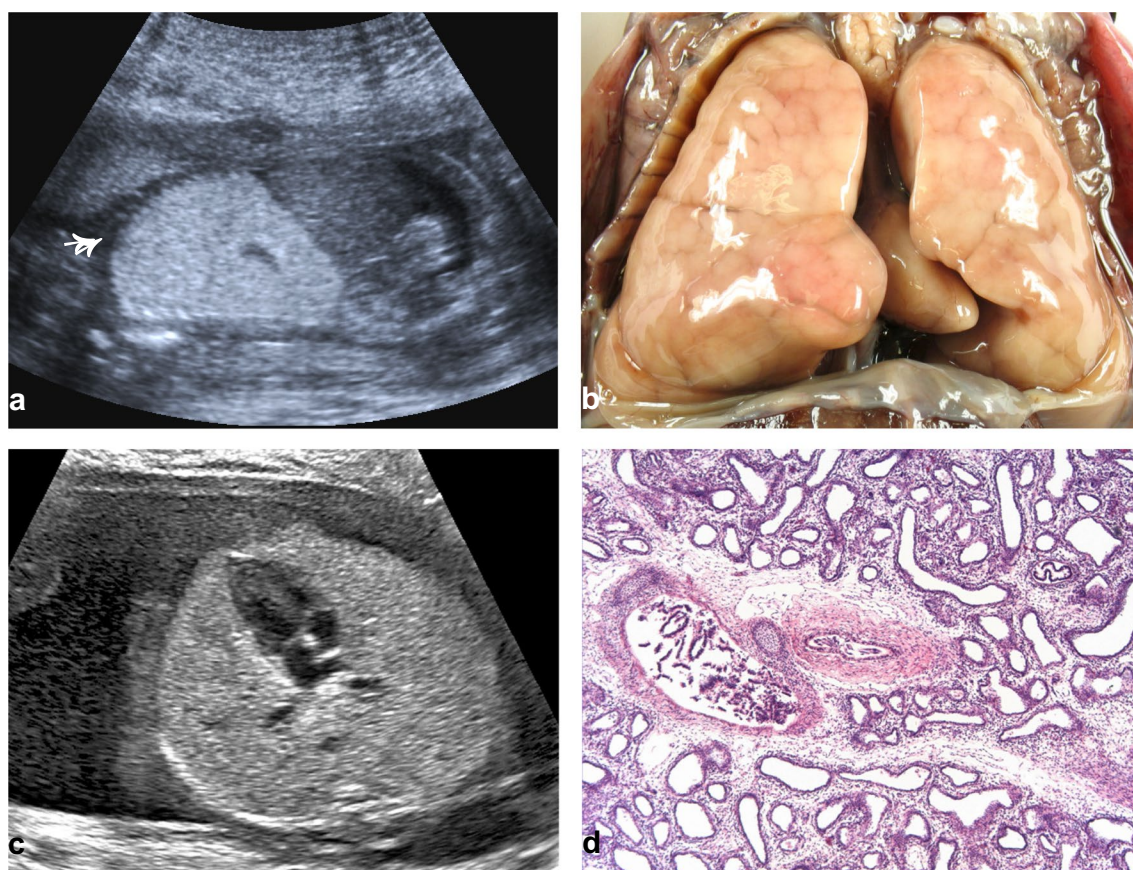


Fig. 4 CHAOS due to tracheal atresia Case 7: Hyperplastic lungs on longitudinal section of the fetal thorax with inverted diaphragm in prenatal ultrasound (a), in situ (b), and on histology (H&E, $\times 5$), showing normal lobulation and distended bronchi and alveoli (d). Case 8: Hyperplastic lungs on transverse section of the thorax in prenatal ultrasound with compression of the heart (c)

expressed ubiquitously in the mouse embryo with only p38 α displaying additional extraembryonic expression in the placenta. p38 is involved in multiple signaling pathways determining the regulation of proliferation, differentiation and transcription and is activated through dual phosphorylation in response to post-inflammatory cytokines and environmental stress-induced signals [36, 37]. p38 α and p38 β have synergistic roles during mouse development and ~70% amino acid sequence identity, but p38 α is expressed at higher levels in many cell types. This may explain the early embryonic lethality associated with intra- and extraembryonic *MAPK14* deletions and the postnatal nonviability owing to respiratory dysfunction associated with embryo-specific *MAPK14* deletions, as opposed to biallelic knockout deletions of *MAPK11* not resulting in aberrant phenotypes, presumably due to compensation of *loss of function* through p38 isoforms [37–39]

The regulatory function of p38 also applies to Sox2. Thus it was shown in melanoma cell lines that p38-dependent phosphorylation and thereby activation

increases SOX2 stability and transcriptional activity [40]. Since Sox2 is expressed throughout the foregut epithelium and needs repression in the ventral foregut to establish Nkx2.1 expression [16], a causal relationship between a *gain of function* *MAPK11* variant promoting phosphorylation and thereby loss of tracheal patterning due to non-suppression of Sox2 seems theoretically conceivable. Unfortunately, we could not sequence the FFPE-DNA of case 2 in order to verify or exclude a *MAPK11* variant in our second child with *isolated* TAG. The lack of evidence of a pathogenic gene variant in our TAG cases 3 to 5 with accompanying VACTERL features may indicate that TAG is part of the VACTERL specific characteristics, a clinical picture affecting different developmental fields at different developmental periods for which no potential teratogenic or genetic alteration common to the majority of cases has yet been identified [41–43].

In summary, TAG and TAT represent different disease entities with respect to clinical features, aetiology and pathogenesis. TAG presents with absence of a trachea due to early lack of ventral tracheal patterning of and

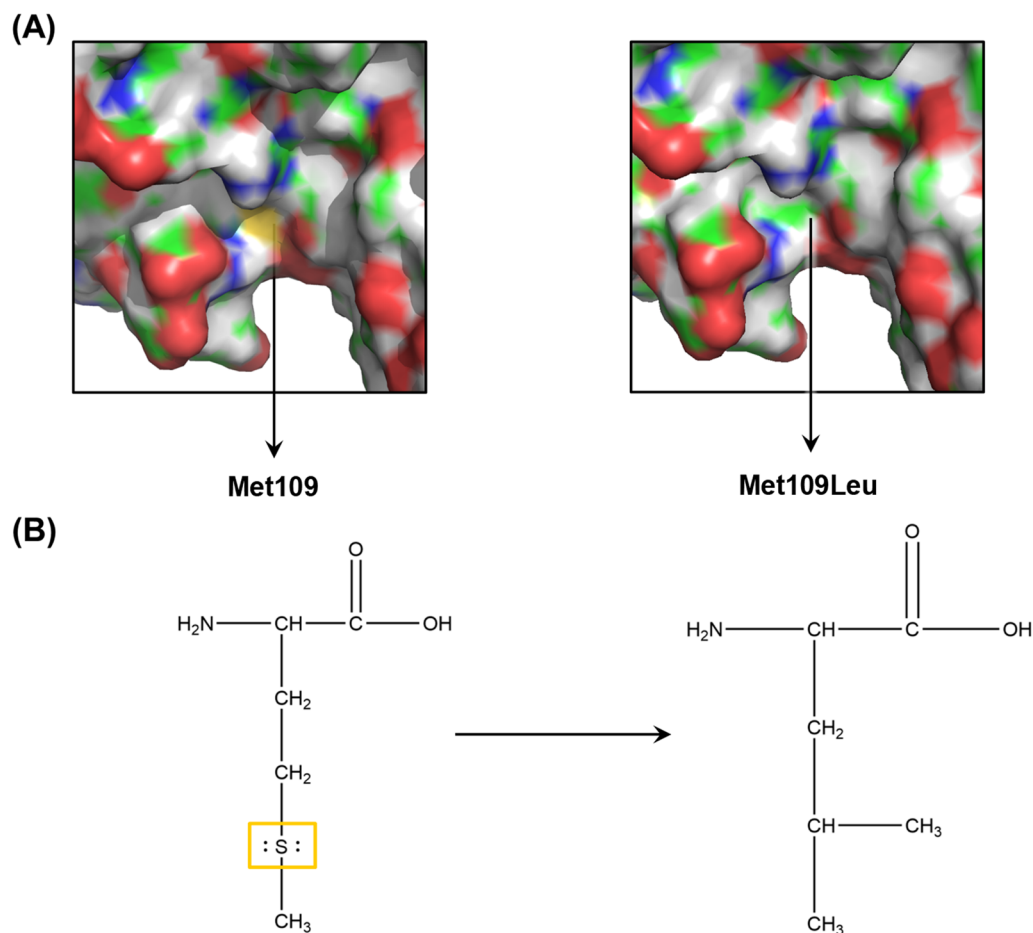


Fig. 5 Structural analysis of the implications of Met109Leu substitution in MAPK11. The ATP binding site of MAPK11 with sulphur atom S_{δ} of Met109 marked in dark yellow (left) which is absent after a substitution Met109Leu (right). The crystalline structure of MAPK11 (PDB ID: 3GP0) was analyzed using the PyMOL software (A). Schematic representation of the structural differences between the methionine and leucine side chains. The sulphur atom S_{δ} of methionine along with its two electron ion pairs is highlighted in a dark yellow rectangle (B)

separation from the foregut, with a BOF/TOF preventing CHAOS, and with pulmonary isomerism; moreover TAG may present as an *isolated* malformation or as part of a VACTERL *association*. TAT is characterized by early degeneration of an existing trachea, by lack of a TOF causing CHAOS, by regular asymmetric lung lobulation, and by single non-VACTERL type accompanying malformations, and is possibly due to vascular disruption. An adverse effect of the identified *novel MAPK11* variant on tracheal development remains speculative. This could be explained by an increase in p38 activation and subsequent Sox2 stability in the ventral foregut. However, it should be kept in mind that the *MAPK11* variant if resulting in *loss of function* may not be pathogenic due to compensation through p38 isoforms, and that p38 β is not the predominant isoform, being expressed at lower levels in most tissues compared to p38 α .

Abbreviations

Array-CGH	Array-based comparative genomic hybridization
ATP	Adenosine triphosphate
Bmp4	Bone morphogenetic protein 4
BOF	Bronchoesophageal fistula
CHAOS	Congenital high airway obstruction sequence
CNV	Copy number variation
ES	(Whole) exome sequencing
FFPE probes	Formalin fixed paraffin embedded probes
gnomAD	Population database (genome aggregation database)
Leu	Leucine
LRD	Limb reduction defect
MRI	Magnetic resonance imaging
<i>MAPK11</i>	Mitogen-Activated Protein Kinase 11 gene
Met	Methionine
Nkx2-1	NKX2 homeobox 1 protein (=thyroid transcription factor1 TTF1)
p38beta	Mitogen activated protein kinase p38beta
PROM	Premature rupture of membranes
Shh	Sonic Hedgehog protein
SMAD1	Suppressor of mothers against decapentaplegic
SNP array	Single nucleotide polymorphism
Sox2	SRY-box 2 protein

TAG	Tracheal agenesis (agenesis= failure of development of an organ)
TAT	Tracheal atresia (atresia= closed or absent lumen of a hollow organ)
TOF	Tracheo-oesophageal fistula
VACTERL	Association of vertebral anomalies, anal atresia, cardiac septal defect, tracheo-oesophageal fistula, renal and radial limb defects
Wnt	Wingless/integrated protein

Acknowledgements

'I can't breathe'—Dedicated to George Perry Floyd (namesake of the creator of Floyd's classification of tracheal agenesis), who died on May 25, 2020 in Minnesota due to tracheolaryngeal compression by a police officer kneeling on his neck and back for nine and a half minutes. We are grateful to Dr. Kurt Köhler, Marienkrankenhaus Papenburg (case 4), to Dr. Reiner Arenz, Kreiskrankenhaus Herzberg (case 6) and to Dr. Gerhard Bender, Frauenklinik Oldenburg (case 5) for providing the clinical data. We greatly appreciate the transmission of parental blood samples in case 5 by Dr. Tanja Vieth (Gynecological Practice in Rhaderfehn). Our special thanks go to Brigitte Hofer (Institute of Medical Genetics Vienna) for her great help with the literature.

Author contributions

MP: Acquisition and interpretation of molecular data in cases 1 to 5, and contribution to the manuscript writing, editing. HR: Study concept, design and coordination, manuscript writing, contribution to editing, and acquisition of autopsy data of case 6. MG: Acquisition and interpretation of molecular data in cases 7 and 8. CB: Acquisition of clinical, autopsy and MRI data and images in case 2. BF: Performance of cytogenetic analyses. CK: Acquisition of genetic counseling and SNP array data in case 4. BB: Acquisition of primary care patient records in case 1. FD: Acquisition of primary care patient records in case 3. MM: Provision of clinical data in case 7. RA: Acquisition of clinical data and ultrasound images in case 8. AB: Acquisition of protein structure analysis data and image. MH: Performance of the exome sequencings. FL: Support and supervision of the study and supplementary processing. KS: Acquisition of autopsy data, X-ray findings and images in cases 1, 3–5 and 7–8 and contribution to editing. All authors approved the final manuscript.

Funding

The study received no financial support.

Availability of data and materials

Data that support the findings of this study are included in this article. Further enquiries can be directed to the corresponding author.

Declarations

Ethics approval and consent to participate

The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of the Medical University of Vienna (Approval No: 1443/2020).

Consent for publication

Written informed consent for all investigations was obtained.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Institute of Medical Genetics, Medical University of Vienna, Waehringer Strasse 10, 1090 Vienna, Austria. ²Institute of Forensic Medicine, University of Zürich, Zurich, Switzerland. ³Berlin School of Economics and Law (HWR), Berlin, Germany. ⁴Institute of Human Genetics, Philipps-University of Marburg, Marburg, Germany. ⁵Institute of Human Genetics, RWTH, Aachen, Germany. ⁶Children's Hospital St. Louise, Paderborn, Germany. ⁷Department of Pediatrics, Children's Center Bethel, University Bielefeld, Bielefeld, Germany. ⁸Department of Gynecology and Obstetrics, University Clinic Oldenburg, Oldenburg, Germany. ⁹Division of Prenatal Medicine and Fetal Therapy, University Hospital Giessen, Giessen, Germany. ¹⁰Institute of Pathology, Fetal Pathology, Philipps-University of Marburg, Marburg, Germany.

Received: 6 November 2023 Accepted: 23 February 2024

Published online: 12 March 2024

References

- Evans JA, Greenberg CR, Erdile L. Tracheal agenesis revisited: analysis of associated anomalies. *Am J Med Genet.* 1999;82:415–22.
- Van Veenendaal MB, Liem KD, Marres HA. Congenital absence of the trachea. *Eur J Pediatr.* 2000;159:8–13. <https://doi.org/10.1007/s004310050002>.
- de Groot-van der Moeren MD, Haak MC, Lakeman P, et al. Tracheal agenesis: approach towards this severe diagnosis. Case report and review of the literature. *Eur J Pediatr.* 2012;171:425–43. <https://doi.org/10.1007/s00431-011-1563-x>.
- Manschot HJ, van den Anker JN, Tibboel D. Tracheal agenesis. *Anaesthesia.* 1994;49:788–90. <https://doi.org/10.1111/j.1365-2044.1994.tb04453.x>.
- Smith MM, Huang A, Labbé M, et al. Clinical presentation and airway management of tracheal atresia: a systematic review. *Int J Pediatr Otorhinolaryngol.* 2017;101:57–64. <https://doi.org/10.1016/j.ijporl.2017.07.028>.
- Vaikunth SS, Morris LM, Polzin W, et al. Congenital high airway obstruction syndrome due to complete tracheal atresia: an accident of nature with clues for tracheal development and lessons in management. *Fetal Diagn Ther.* 2009;26:93–7. <https://doi.org/10.1159/000242454>.
- Sanford E, Saadai P, Lee H, et al. Congenital high airway obstruction sequence (CHAOS): a new case and a review of the phenotypic features. *Am J Med Genet A.* 2012;2012(158A):3126–36. <https://doi.org/10.1002/ajmg.a.35643>.
- Floyd J, Campbell DC, Dominy DE. Agenesis of the trachea. *Am Rev Respir Dis.* 1962;86:557–60. <https://doi.org/10.1164/arrd.1962.86.4.557>.
- Faro RS, Goodwin CD, Organ CH Jr, et al. Tracheal agenesis. *Ann Thorac Surg.* 1979;28(3):295–9. [https://doi.org/10.1016/s0003-4975\(10\)63123-2](https://doi.org/10.1016/s0003-4975(10)63123-2).
- Coleman AM, Merrow AC, Elluru RG, et al. Tracheal agenesis with tracheo-oesophageal fistulae: fetal MRI diagnosis with confirmation by ultrasound during an ex utero intrapartum therapy (EXIT) delivery and postdelivery MRI. *Pediatr Radiol.* 2013;43:1385–90. <https://doi.org/10.1007/s00247-013-2679-0>.
- Bertholdt C, Perdirolle-Galet E, Bach-Segura P, et al. Tracheal agenesis: a challenging prenatal diagnosis—contribution of fetal MRI. *Case Rep Obstet Gynecol.* 2015. <https://doi.org/10.1155/2015/456028>.
- Desmore JC, Oldham KT, Dominguez KM, et al. Neonatal esophageal trachealization and esophagocarinoplasty in the treatment of flow-limited Floyd II Tracheal agenesis. *J Thorac Cardiovasc Surg.* 2017;153(6):e121–5. <https://doi.org/10.1016/j.jtcvs.2017.01.029>.
- Straughan AJ, Mulcahy CF, Sandler AD, et al. Tracheal agenesis: Vertical division of the native esophagus—a novel surgical approach and review of the literature. *Ann Otol Rhinol Laryngol.* 2021;130(6):547–62. <https://doi.org/10.1177/0003489420962124>.
- Merei JM, Farmer P, Hasthorpe S, et al. Timing and embryology of esophageal fistula and trachea-esophageal fistula. *Anat Res.* 1997;249:240–8. [https://doi.org/10.1002/\(SICI\)1097-0185\(199710\)249:2%3C240::AID-AR11%3E3.0.CO;2-O](https://doi.org/10.1002/(SICI)1097-0185(199710)249:2%3C240::AID-AR11%3E3.0.CO;2-O).
- Li Y, Gordon J, Manley NR, et al. Bmp4 is required for tracheal formation: a novel mouse model for tracheal agenesis. *Dev Biol.* 2008;322:145–55. <https://doi.org/10.1016/j.ydbio.2008.07.021>.
- Billmyre KK, Hutson M, Klingensmith J. One shall become two: separation of the esophagus and trachea from the common foregut tube. *Dev Dyn.* 2015;244:277–88. <https://doi.org/10.1002/dvdy.24219>.
- Edwards NA, Shacham-Silverberg V, Weitz L, et al. Developmental basis of trachea-esophageal birth defects. *Dev Biol.* 2021;477:85–97.
- Minoo P, Su G, Drum H, et al. Defects in tracheo-oesophageal and lung morphogenesis in Nkx2.1 (–/–) mouse embryos. *Dev Biol.* 1999;209:60–71. <https://doi.org/10.1016/j.ydbio.2021.05.015>.
- Williams AK, Quan QB, Beasley SW. Three-dimensional imaging clarifies the process of tracheo-oesophageal separation in the rat. *J Pediatr Surg.* 2003;38(2):173–7. <https://doi.org/10.1053/jpsu.2003.50037>.
- Cardoso WV, Lü J. Regulation of early lung morphogenesis: questions, facts and controversies. *Development.* 2006;133:1611–24. <https://doi.org/10.1242/dev.02310>.

21. Que J, Okubo T, Goldenring JR, et al. Multiple dose-dependent roles for Sox2 in the patterning and differentiation of anterior foregut endoderm. *Development*. 2007;134:2521–31. <https://doi.org/10.1242/dev.003855>.
22. Goss AM, Tian Y, Tsukiyama T, et al. Wnt2/2b and beta-catenin signalling are necessary and sufficient to specify lung progenitors in the foregut. *Dev Cell*. 2009;17:290–8. <https://doi.org/10.1016/j.devcel.2009.06.005>.
23. Rankin SA, Han L, McCracken KW, et al. A retinoic acid-hedgehog cascade coordinates mesoderm-inducing signals and endoderm competence during lung specification. *Cell Rep*. 2016;16(1):66–78. <https://doi.org/10.1016/j.celrep.2016.05.060>.
24. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015;17:405–24.
25. Filippakopoulos P, Barr A, Fedorov O, et al. Structural Genomics Consortium (SGC). <https://doi.org/10.2210/pdb3gp0/pdb>.
26. Schrödinger L, DeLano W. PyMOL 2020. <http://www.pymol.org/pymol>.
27. Wang Z, Harkins PC, Ulevitch RJ, et al. The structure of mitogen-activated protein kinase p38 at 2.1-Å resolution. *Proc Natl Acad Sci USA*. 1997;94(6):2327–32. <https://doi.org/10.1073/pnas.94.6.2327>.
28. Patel SB, Cameron PM, O'Keefe SJ, et al. The three-dimensional structure of MAP kinase p38beta: different features of the ATP-binding site in p38beta compared with p38alpha. *Acta Crystallogr D Biol Crystallogr*. 2009;65(Pt 8):777–85. <https://doi.org/10.1107/S090744490901600X>.
29. Ghaye B, Szapiro D, Fanchamps JM, et al. Congenital bronchial abnormalities revisited. *Radiographics*. 2001;21(1):105–19. <https://doi.org/10.1038/gim.2015.30>.
30. Das BB, Nagaraj A, Rao AH, et al. Tracheal agenesis: report of three cases and review of the literature. *Am J Perinatol*. 2002;19(7):395–400. <https://doi.org/10.1055/s-2002-35610>.
31. Mohammed H, West K, Bewick J, et al. Tracheal agenesis, a frightening scenario. *J Laryngol Otol*. 2016;130(3):314–7. <https://doi.org/10.1017/S0022215115003515>.
32. Bergman JEH, Löhner K, van der Sluis CK, et al. Etiological diagnosis in limb reduction defects and the number of affected limbs: A population-based study in the Northern Netherlands. *Am J Med Genet A*. 2020;182(12):2909–18. <https://doi.org/10.1002/ajmg.a.61875>.
33. Reece AS, Hulse GK. Epidemiological association of cannabinoid- and drug- exposures and sociodemographic factors with limb reduction defects across USA 1989–2016: A geotemporal study. *Spat Spatiotemporal Epidemiol*. 2022;41:100480. <https://doi.org/10.1016/j.sste.2022.100480>.
34. Hoyme HE, Jones KL, Van Allen MI, et al. Vascular pathogenesis of transverse limb reduction defects. *J Pediatr*. 1982;101(5):839–43. [https://doi.org/10.1016/s0022-3476\(82\)80343-0](https://doi.org/10.1016/s0022-3476(82)80343-0).
35. Salassa JR, Pearson BW, Payne WS. Gross and microscopical blood supply of the trachea. *Ann Thorac Surg*. 1977;24(2):100–7. [https://doi.org/10.1016/s0003-4975\(10\)63716-2](https://doi.org/10.1016/s0003-4975(10)63716-2).
36. Stein B, Yang MX, Young DB, et al. p 38–2, a novel mitogen-activated protein kinase with distinct properties. *J Biol Chem*. 1997;272(31):19509–17. <https://doi.org/10.1074/jbc.272.31.19509>.
37. Beardmore VA, Hinton HJ, Eftychi C, et al. Generation and characterization of p38β (MAPK11) gene-targeted mice. *Mol Cell Biol*. 2005;25(23):10454–64. <https://doi.org/10.1128/MCB.25.23.10454-10464.2005>.
38. Hui L, Bakiri L, Mairhofer A, et al. p38alpha suppresses normal and cancer cell proliferation by antagonizing the JNK-c-Jun pathway. *Nat Genet*. 2007;39(6):741–9. <https://doi.org/10.1038/ng2033>.
39. Del Barco BI, Coya JM, Maina F, et al. Genetic analysis of specific and redundant roles for p38alpha and p38beta MAPKs during mouse development. *Proc Natl Acad Sci USA*. 2011;108(31):12764–9. <https://doi.org/10.1073/pnas.1015013108>.
40. Pietrobono S, De Paolo R, Mangiameli D, et al. p38 MAPK-dependent phosphorylation of transcription factor SOX2 promotes an adaptive response to BRAF inhibitors in melanoma cells. *J Biol Chem*. 2022;298(9):102353. <https://doi.org/10.1016/j.jbc.2022.102353>.
41. Stevenson RE, Hunter AGW. Considering the embryopathogenesis of VACTERL association. *Mol Syndromol*. 2013;4(1–2):7–15. <https://doi.org/10.1159/000346192>.
42. Zhang R, Marsch F, Kause F, et al. Array-based molecular karyotyping in 115 VATER/VACTERL and VATER/VACTERL-like patients identifies disease-causing copy number variations. *Birth Defects Res*. 2017;109(13):1063–9. <https://doi.org/10.1002/bdr2.1042>.
43. Thiem CE, Stegmann JD, Hilger AC, et al. Re-sequencing of candidate genes FOXF1, HSPA6, HAAO, and KYNU in 522 individuals with VATER/VACTERL, VACTER/VACTERL-like association, and isolated anorectal malformation. *Birth Defects Res*. 2022;114(10):478–86. <https://doi.org/10.1002/bdr2.2008>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.