Modern Photocatalytic Approaches to Carbon-centered Radical Generation for Sustainable Synthesis of Pharmaceutically Relevant Scaffolds

Vor der Fakultät für Mathematik, Informatik und Naturwissenschaften der RWTH Aachen University zur Erlangung des akademischen Grades eines Doktors der Naturwissenschaften genehmigte Dissertation

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Tag der mündlichen Prüfung: 21.05.2025

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- 5. All significant sources of support and contributions have been duly acknowledged
- 6. In cases where this thesis includes collaborative work, I have clearly specified my contributions as well as those of my co-researchers.
- 7. Parts of this work have been published previously. A full list of references is provided below.
- 8. ChatGPT were used for the language corrections of this thesis.

The doctoral work in this thesis was performed from October 2021 until February 2025 at the Institute of Organic Chemistry, RWTH Aachen University, under the supervision of Professor Dr. Frederic W. Patureau.

The doctoral work has led to the following publications:

- 1. **D. Liu**, F. Xiao, B. Ebel, I. M. Oppel and F. W. Patureau, Visible-light-mediated radical α -C(sp³)-H *gem*-difluoroallylation of amides with trifluoromethyl alkenes via halogen atom transfer and 1,5-hydrogen atom transfer. *Org. Lett.* **2025**, *27*, 2377.
- 2. **D. Liu**, and F. W. Patureau, Visible-light-induced photocatalytic deoxygenative benzylation of quinoxalin-2-(1*H*)-ones with carboxylic acid anhydrides. *Org. Lett.* **2024**, *26*, 6841.
- 3. **D. Liu**, Y. Zhao and F. W. Patureau, NaI/PPh₃-catalyzed visible-light-mediated decarboxylative radical cascade cyclization of *N*-arylacrylamides for the efficient synthesis of quaternary oxindoles. *Beilstein J. Org. Chem.* **2023**, *19*, 57.

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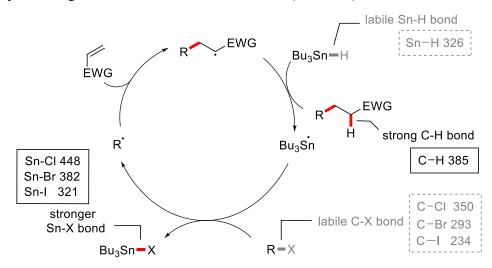
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1 Introduction

1.1 Photocatalysis

As a significant class of open-shell species, carbon-centered radicals are intriguing neutral intermediates that have gained broad utility in the fields of modern organic synthesis, material science and biological chemistry, despite the early doubt regarding their potential application. ¹⁻⁵ Particularly, alkyl and benzoyl radical generation under mild conditions, along with their controlled and selective transformations, have become research topics of increasing interest. An early example of generating alkyl radicals from organic halides is illustrated in **Scheme 1**. Organotin compounds were identified as essential for promoting radical chain reactions, as reported nearly 60 years ago. ⁶ The discovery of Bu₃SnH represented a major advancement in radical chemistry, as it enabled the production of Bu₃Sn· as the radical chain carrier and served as a hydrogen atom donor to complete the catalytic cycle. Subsequently, the alkyl or benzoyl radicals⁷⁻⁸ formed via tin-mediated processes were effectively trapped by various radical acceptors, facilitating the construction of novel functional molecules. Analysis of bond dissociation energies (BDE) of the reactants revealed that the key of this method lies in the formation of stronger Sn–X and C–H bonds, driven by the homolytic cleavage of the weaker Sn–H and C–X bonds (**Scheme 1**). ⁹⁻¹⁰



Bond energies are reported in kJ/mol from Ref. 9 and Ref. 10

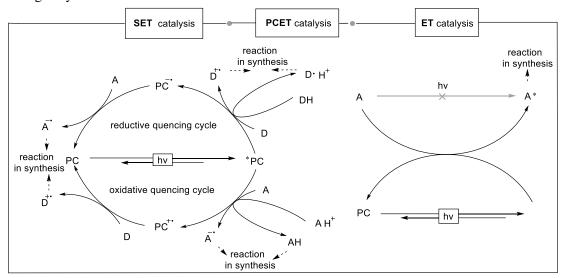
Scheme 1 Thermal radical initiation from alkyl halides using organotin compounds

Although organotin molecules performed competitively in the radical-generating processes, ¹¹⁻¹⁴ the use of these highly versatile species raised significant concerns regarding environmental and health safety, ¹⁵ due to the substantial toxicity and high biological activity of triorganotin molecules. Studies disclosed an LD₅₀ of approximately 0.7 mmol/kg in murine species and prolonged persistence of these organotins in aquatic environment, ¹⁶ these inherent issues necessitate the exploration of viable alternatives.

Efforts have been made to replace toxic tin derivatives with reagents such as (TMS)₃SiH,¹⁷ peroxides,¹⁸⁻²² xanthates,²³ organoboranes,²⁴ thiols,²⁵, P-H based reagents,²⁵ various metal oxidants²⁶ or reductants²⁷⁻²⁸ or others.²⁹ However, these traditional methods generating alkyl or benzoyl radicals typically require harsh reaction conditions such as harmful UV irradiation or high temperatures, stoichiometric amounts of hazardous reagents, or strong oxidants and reductants,

which limit their practical utility in synthetic chemistry. Herein, more sustainable and attractive synthetic strategies based on radical chemistry is desirable.

In recent years, visible-light-mediated photocatalysis has emerged as a powerful tool for the generation of a wide range of radical species, opening up new opportunities in synthesizing diverse organic scaffolds. 30-37 Upon the irradiation of light, photocatalysts (PCs) or photoactive molecules absorb the energy of photons and are promoted to excited state *PC, enabling various chemical processes such as single electron transfer (SET), proton-coupled electron transfer (PCET) or energy transfer (ET). 38-44 These processes have expanded the range of accessible radical precursors, even revolutionized the way to carry out radical chemistry (**Scheme 2**). A key factor in this transformation is the photon itself, which serves as an ideal component for chemical reactions, acting as a traceless reagent, catalyst, or promoter that leaves no toxic residues in the final mixture. 45-46 All these unique features of photocatalysis make it an ideal and elegant strategy for achieving alkylation and benzoylation reactions, which are key steps in the synthesis of versatile natural products and biologically active molecules. 47-48



Scheme 2 Mechanistic pathways of photoredox catalysis. D: donor, A: acceptor, PC: photocatalyst

1.2 Photocatalyzed radical generation from carboxylic acids and their

derivatives

The transformations of functional groups serve as a fundamental pillar of modern organic synthesis. Notably, the efficient interconversion of existing functional groups in widely accessible chemical feedstocks into other valuable functionalities has attracted significant attention in recent studies. Toward this goal, using carboxylic acids and their derivatives as starting materials is synthetically appealing, these privileged chemical entities are advantageous due to their abundant chemical sources, commercially availabilities and chemical versatility, which collectively enhance synthetic efficiency.

1.2.1 Alkyl radical generation via decarboxylation

Compared to transition-metal catalyzed reactions that typically rely on organohalides or pseudohalides as electrophiles coupled with organometallic intermediates, $^{49-52}$ which are mostly moisture-sensitive and require fresh preparation, photocatalyzed decarboxylative reactions have provided a powerful platform to construct diverse carbon-carbon and carbon-heteroatom bonds. $^{53-54}$ Among them, using readily accessible alkyl carboxylic acids and their derivatives to generate versatile alkyl radicals are appealing. Those alkyl radicals from photochemical decarboxylation are synthetically equivalent to nucleophilic organometallic species in coupling reactions, this eliminates the need for expensive and labile carbon nucleophilic reagents. Furthermore, under oxidative conditions, alkyl species with electrophilic reactivity are possibly generated via decarboxylation and serve as a substitute of alkylhalides, which are more challenging in transition-metal catalyzed coupling reaction due to the risk of β -hydride elimination of alkyl metal components formed in situ during catalytic cycle. Therefore, many efforts have been devoted to developing photocatalyzed decarboxylative reactions employing aliphatic carboxylic acids and their derivatives as $C(sp^3)$ source, with the purpose of complex and functional molecules synthesis. $^{55-57}$

1.2.1.1 Reactions involving carboxylic acids

The nature of organic fragments attached to the carboxylic acid (COOH) group can have a significant impact on radical generation, radical stability and subsequent coupling reactions. For instance, the bond dissociation energy (BDE) of the alkyl-COOH bond (\sim 380 kJ/mol) is lower than that of the vinyl-COOH (435 kJ/mol) and arene-COOH (426 kJ/mol) bonds. This difference arises from the stronger C(sp²)-COOH bond, making it easier to generate alkyl radicals from alkyl carboxylic acids via decarboxylation under relatively mild conditions. Additionally, carboxylic acids with benzylic, allylic, α -carbonyl, or α -heteroatom substituents exhibit higher reactivity compared to other less reactive carboxylic acids. Regardless of the cleavage efficiency of diverse carboxylic acid bonds, decarboxylative reactions involving carboxylic acids can be broadly categorized into three types from a photocatalytic perspective:

I redox-neutral photocatalysis

In a redox-neutral photocatalysis, decarboxylation proceeds through the action of the light and photocatalysts without requiring any external stoichiometric oxidants or reductants, this process features a photoredox catalyst-mediated electron shuttle cycle between two reactants, which is the key to decarboxylative coupling.

II dual catalysis.

Dual catalysis, also referred to as metallaphotocatalysis, combines redox-neutral photocatalysis with transition metal co-catalysis. In this pathway, a radical species, generated by the photocatalyst, reacts with an intermediate formed via the oxidative addition of a low-valent transition metal catalyst into an organic (pseudo) halide. This process is followed by reductive elimination, yielding targeted product.

III with super-stoichiometric external oxidant

In a few examples, excess oxidants are required in oxidative decarboxylative reaction.

Some pioneered works belong to type I are presented here.

In 2007, Yoshimi and Hatanaka group reported decarboxylative reduction of free aliphatic carboxylic acids via a photogenerated radical cation of phenanthrene, with thiols serving as hydrogen atom donors.⁶⁵ During this process, C(sp³)-C(sp²) coupling side reactions were observed between alkyl radicals generated from carboxylic acids and 1,4-dicyanobenzene. Soon after, this side reaction was systematically explored, and revealed that a lot of aliphatic carboxylate ions could couple with dicyanobenzenes in the presence of phenanthrene under irradiation with a 500W highpressure mercury lamp through a Pyrex filter, which restricts the light to > 300 nm wavelength.⁶⁶ Recognizing the potential application of these C(sp³)-C(sp²) coupling products, in 2014 MacMillan group disclosed a redox-neutral decarboxylative coupling of α -amino acids and electron-deficient cyanoaromatics (Scheme 3a).⁶⁷ Inspired by their earlier work involving Ir(ppy)₃-mediated single electron reduction of 1,4-dicyanobenzene,⁶⁸ the more strongly oxidizing Ir[p-F(t-Bu)ppy]₃ **3** (E_{1/2red} [Ir(IV)/*Ir(III)] = -1.67 V vs SCE) was identified as a suitable photocatalyst to promote this decarboxylative arylation. In the optimized condition, the α -amino acid 1 and cyanoarene 2 could smoothly react to afford desired product 4 utilizing iridium photocatalyst 3 and CsF as a base in DMSO under the irradiation of a compact fluorescent lamp (CFL). This method was proved efficiently for a number of α -amino and α -oxy acids, which underwent radical-radical coupling with cyanoarenes bearing electron-deficient substituents including nitrile, sulfonyl, carboxylic ester and phosphonate, offering rapid access to prevalent benzylic amine architectures. Compared to previous work of Yoshimi and Hatanaka,66 this protocol is more operative due to a simple, energy-efficient CFL without the need of specialized glass apparatus connected with mercury lamp. In the proposed reaction pathway, cyanoarene 2 is converted to radical anion 5 through the photoexcited iridium photocatalyst 3. Subsequently, single electron oxidation of protected α -amino acid 1 generates relatively stable α -aminoalkyl radical 6. Finally, the coupling of radical 5 and radical 6 formed target product 4, accompanied by the elimination of cyanide.

In 2016, Opatz reported transition-metal-free decarboxylative arylation of carboxylic acids with cyanobenzenes, a loading of 75 mol% of inexpensive organic photosensitizer phenanthrene was used under conditions similar to those reported by Yoshimi and Hatanaka. ⁶⁶ This approach exhibited a broader substrate scope, encompassing α -amino, α -oxy, α -benzyl and α -tertiary carboxylic acids. ⁶⁹ Three years later, Wang group developed an improved transition-metal-free protocol using 3-amino-9*H*-fluorene-2,4-dicarbonitrile as an organo-photocatalyst. Remarkably, only 2 mol% of the catalyst was required for effective decarboxylative cross-coupling with aromatic nitriles, significantly enhancing the reaction efficiency and practicality. ⁷⁰

In addition of $C(sp^3)$ -(hetero)aryl $C(sp^2)$ bond formation in decarboxylative reaction, in 2014, Noble and MacMillan disclosed a photocatalytic decarboxylative alkenylation. With the treatment of N-Boc- α -amino acid **8** with vinyl sulfone **9** in the presence of photocatalyst [Ir(ppy)₂(dtbbpy)PF₆] **10** and CsHCO₃ as the base under the irradiation of 24 W CFL in 1,4-dioxane, various allylic amines

a) photocatalyzed decarboxylative arylation of α -amino acids and proposed mechanism

Scheme 3 Redox-neutral photocatalyzed decarboxylative reaction for constructing C(sp³)-C(sp²) bond

11 could be obtained in good yields and with great E/Z selectivity (E/Z > 94:6) (Scheme 3b). A variety of β -(hetero)aryl vinyl sulfones bearing electron-rich and electron-deficient aromatic rings performed well with a range of α -amino acids, providing a complementary method to allyl amines in comparison to stoichiometric metal-based methods.⁷²

Moreover, Opatz and coworkers reported an efficient transition-metal free decarboxylative synthesis of β -alkylated vinyl sulfones or (acrylic) nitriles from structurally diverse carboxylic acids 12 including peptides and steroids, 1,2-bis(phenylsulfonyl)ethylenes 13a or 1-cyano-2-phenysulphonyl

ethylenes **13b** was demonstrated as viable radical acceptors in the presence of 9,10-dicyanoanthracene (DCA **14**) and redox mediator biphenyl (**Scheme 3b**).⁷³

Notably, gem-difluoroalkenes 16 were reported by Fu group to react with α -amino acids 8 with $Ir[dF(CF_3)ppy]_2(dtbbpy)PF_6$ 17, and Li_2CO_3 in DMSO irradiated by 23 W CFL under argon atmosphere, a monofluoroallylamine mixture 18 of Z-isomer and E-isomer was obtained efficiently in this decarboxylative vinylation regardless of electronic properties of aryl rings attached to gem-difluoroalkenes (Scheme 3b).

Among the earliest examples for forging $C(sp^3)$ - $C(sp^3)$ bonds via decarboxylative coupling, in 2014, the MacMillan group developed a radical-mediated decarboxylative coupling of carboxylic acids **19** with electron-deficient alkenes **20** using iridium photocatalyst **17** (1 mol%) (**Scheme 4a**).⁷⁵ The reaction has a broad substrate scope of carboxylic acids including α -amino acids, α -oxy acids, and secondary and tertiary alkyl carboxylic acids. Additionally, a diverse array of electron-deficient olefins, such as α , β -unsaturated esters, sulfones, imides, malonates and maleates, were good Michael acceptors in this decarboxylative 1,4-conjugate addition reaction. Notably, unlike earlier methods reported by the Yoshimi and Hatanaka group,⁷⁶ this procedure operates under simple conditions using a household CFL without the need for specialized glass apparatus.

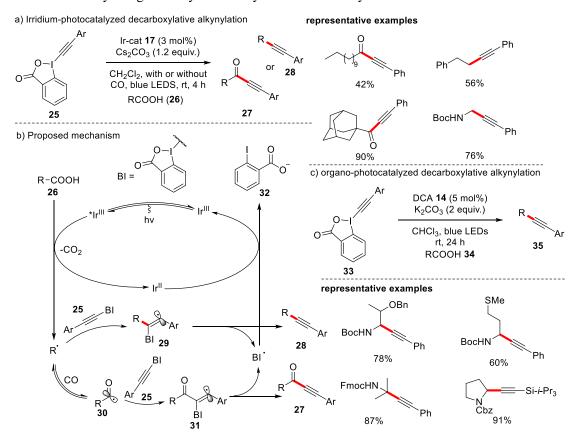
Subsequently this 1,4-conjugate radical addition strategy was extended to decarboxylative coupling of α -heteroatom substituted carboxylic acids with a distinct class of styrene-based, electron-deficient alkenes.⁷⁷ The reaction was conducted under conditions similar to the MacMillan protocol.⁷⁵

a) Ir-photocatalyzed decarboxylative 1,4-conjugate addition to alkenes

Scheme 4 Redox-neutral photocatalysis for C(sp³)-C(sp²) coupling

In 2017, the Fujimoto group elegantly extended the application of 1,4-conjugate radical addition method to the formal preparation of L-ossamine (24) prevalent in natural product ossamycin.⁷⁸ The protected D-threonine 22 was subjected to react with methyl acrylate under the conditions of Ircatalyst 17, Cs₂CO₃ as the base, and a DMF/H₂O (10:1) solvent system. The reaction was performed in a flow reactor, irradiated with blue LEDs, and maintained at a residence time (t_R) of 4 hours to afford the addition product 23, which was subsequently converted into L-ossamine 24 (Scheme 4b).

To expand the range of Michael acceptors in this photocatalyzed decarboxylative radical addition process, Aggarwal⁷⁹ in 2018 described a procedure involving the addition of alkyl radicals to vinyl boronic esters, which are relatively weakly electron-deficient alkenes compared to acrylates. Recently, Shah developed a Giese-type radical addition protocol for synthesizing racemic unnatural α -amino acids by using a variety of carboxylic acids with dehydroalanine esters.⁸⁰



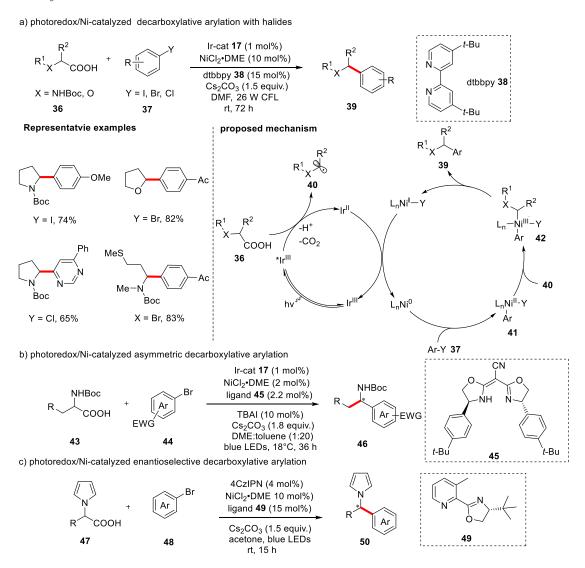
Scheme 5 Redox-neutral photocatalysis for C(sp³)-C(sp) bond formation

Regarding $C(sp^3)$ -C(sp) bond formation in decarboxylative coupling reaction, the Xiao and Lu group in 2015 reported visible light-induced decarboxylative alkynylation and carbonylative alkynylation of readily available carboxylic acids **26** with ethynylbenziodoxolones **25** as the alkynylating agent in the presence of Ir-cat **17** and Cs_2CO_3 as the base. A wide variety of alkyl carboxylic acids including α -heteroatom substituted acids performed well in this coupling reaction. Interestingly, the addition of carbon monoxide enabled the production of carbonylative products **27** with high efficiency. In contrast to Li's approach, this reaction demonstrated a broader carboxylic acid substrate scope and good functional group tolerance, attributed to the absence of additional oxidants (**Scheme 5a**).

In the first proposed pathway, alkyl radical $R\cdot$, generated through SET oxidation of carboxylic salt of 26 by photoexcited iridium catalyst, undergoes radical addition with reagent 25, leading to alkenyl radical 29. Then the elimination of benziodoxolonyl radical species (BI·) affords product alkyl-substituted alkyne 28. In the second proposed pathway involving CO, the alkyl radical $R\cdot$ combines with CO to generate acyl radical 30. This intermediate undergoes radical addition and elimination steps analogous to those in the first pathway, ultimately yielding the α -carbonylated alkyne 27. Within the photocatalytic cycle, the iridium catalyst in its Ir^{II} state reduces the benziodoxolonyl radical $BI\cdot$, producing byproduct 32 and regenerating the Ir^{III} photocatalyst to

complete the catalytic cycle (Scheme 5b).

Soon after, DCA 14 as a more cost-effective alternative to expensive iridium photocatalyst was investigated to catalyzed this photoredox decarboxylative alkynylation by Li and co-workers (Scheme 5c). Biverse α -heteroatom substituted and tri-alkylsilyl substituted alkyne 35 were efficiently synthesized under the optimized conditions of DCA 14 (5 mol%), K_2CO_3 (2.0 equiv.) in CHCl₃ irradiated with blue LEDs for 24 h.



Scheme 6 Photoredox/Ni-catalyzed decarboxylative arylation

Representative reactions demonstrating dual catalysis approach are summarized below.

The pioneering work on metallaphotocatalysis was conducted by the MacMillan and Doyle groups, ⁸⁴ who designed a dual catalyst system merging photocatalysis and nickel catalysis. This innovative approach enabled the decarboxylative coupling of a broad range of (hetero)aryl halides as sp²-hybridized carbon sources. Notably, this strategy provided a novel platform for C(sp³)-C(sp²) bond formation, which could not be achieved using either photoredox catalysis or transition metal catalysis alone. Under optimized conditions, the treatment of carboxylic acid **36** and aryl halide **37** with Ir-cat **17**, NiCl₂·DME, the ligand 4,4'-di-*tert*-butyl-2,2'-bipyridine (dtbbpy, **38**) and base Cs₂CO₃ in DMF under irradiation of 26 W CFL yielded desired product **39** in respectable yields. A variety of α-amino acids and α-oxy acids were proved to be highly efficient partners with aryl and

heteroaryl halides bearing diverse electron donating and electron-withdrawing substituents (Scheme 6a).

In the postulated mechanism, the SET oxidation of carboxylic salt of **36** by photoexcited *Ir^{III} generates α-amino/oxy alkyl radical **40** and Ir^{III} species. Then alkyl radical **40** is captured by intermediate **41** formed through oxidative addition of Ni⁰ catalyst to aryl halides **37**, leading to the formation of Ni^{III} complex **42**. Next the reductive elimination of **42** produces coupling product **39** and Ni^I species. Finally, the Ni⁰ catalyst and Ir^{III} photocatalyst are regenerated via SET process between Ir^{III} species and Ni^I species, thereby completing their respective catalytic cycle (**Scheme 6a**).

Furthermore, using cost-effective organophotocatalyst 4CzIPN (2,4,5,6-tetrakis(carbazol-9-yl)-1,3-dicyanobenzene) in place of more expensive iridium catalysts was demonstrating to be viable in these decarboxylative reactions, delivering diverse C(sp³)-C(sp²) cross-coupling arenes including bioactive aryl and heteroaryl-*C*-nucleoside.⁸⁵

Later, this multi-catalyst system was extended to decarboxylative asymmetric synthesis of $C(sp^3)$ - $C(sp^2)$ bond by the Fu and MacMillan group in 2016.⁸⁶ While employing the same combination of an Ir-photocatalyst and a Ni-catalyst as used in their previous studies,⁸⁴ the chiral cyanobisoxazoline ligand **45** was introduced in place of the achiral bipyridine ligand for Ni. This modification enabled the enantioselective coupling process, facilitating the synthesis of chiral benzyl amine **46** with good to excellent enantiomeric excess (**Scheme 6b**). Unlike the racemic methods,^{84,87} which allowed electronically unbiased aryl halides to serve as effective coupling partners, this enantioselective decarboxylative arylation was limited to electron-deficient aryl bromides as substrate.

Soon after, the Davidson and Bonifazi group expanded the substrate scope to α -amino-N-heterocyclic carboxylic acid **47** for asymmetric decarboxylative $C(sp^3)$ - $C(sp^2)$ cross-coupling reaction with electron-withdrawing aryl bromides. ⁸⁸ A synergistic merger of the organophotocatayst 4CzIPN and a Ni-catalyst combined with a chiral pyridine-oxazoline **49** (PyOX) ligands was used to achieve an enantioselective synthesis of N-benzyl-substituted heterocycles **50** in good enantioselectivity and efficiency (**Scheme 6c**).

Building upon these foundational work, aryl triflates 51⁸⁹ and DNA-tagged aryl iodide 53⁹⁰ respectively were identified as competent substrates for visible-light mediated Ir/Ni-catalyzed decarboxylative arylation under conditions similar to those established by the MacMillan and Doyle protocol⁸⁴ (Scheme 7).

R1-X
$$R^2$$
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 R^6

Scheme 7 Photoredox/Ni-catalyzed decarboxylative reactions

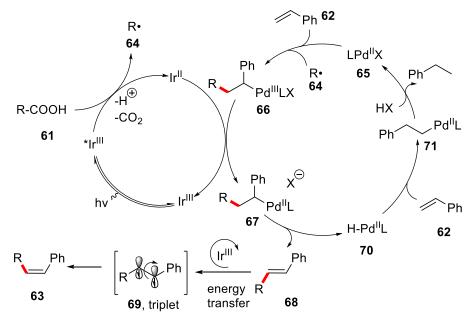
As an extension of metallaphotoredox-catalyzed decarboxylative arylation,⁸⁴ MacMillan and coworkers reported radical decarboxylative alkenylation of alkyl carboxylic acid with vinyl halide **55** in 2015.⁹¹ Diverse vinyl iodides and bromides give rise to vinylation products **56** in high efficiency and with good stereoselectivities (**Scheme 7**). Afterwards, they exploited a dual catalytic system for the decarboxylative hydroalkylation of unactivated terminal and internal alkynes **57**, delivering alkylated olefin **58** with excellent Z/E selectivity (**Scheme 7**).⁹² Moreover, in 2016 the MacMillan group developed photoredox/Ni-catalyzed decarboxylative alkylation with alkyl bromides (**Scheme 7**).⁹³ A wide range of bromoalkanes bearing epoxide, alcohol, aldehyde, olefin ester and chloride functional groups were compatible partners to couple with α -amino acids, α -oxy acids, and primary and secondary alkyl carboxylic acids. This discovery represents a significant breakthrough in the realm of C-C bond formation since both coupling partners are bench-stable and commercially available.

In an effort towards selectively synthesize thermodynamically disfavored (Z)-olefins, Na and Shang developed a decarboxylative cross-coupling reaction between carboxylic acids and styrenes by integrating photoredox catalysis with palladium catalysis.⁹⁴ Through extensive screening experiments, they identified a catalytic system comprising Ir-cat 17 and Pd(OAc)₂/TMP (3,4,7,8-tetramethyl-1,10-phenanthroline) as highly effective in producing β -alkylated styrenes 63 with excellent (Z)-selectivity (Scheme 8a).

The proposed mechanism involves the alkyl radical **64** formed via SET oxidation of carboxylic salt of **61** by excited Ir photocatalyst, which is subsequently intercepted by styrene **62** and Pd^{II} species **65** to form Pd^{III} complex **66**. Then the SET reduction of **66** generates Pd^{II} intermediate **67**, which undergoes a β -elimination to offer *E*-olefin **68** and Pd^{II}-H species **70**. Next the Pd^{II}-H species **70** inserts into styrene **62** to generate phenethyl Pd^{II} **71** followed by protonation to give ethylbenzene and regenerated Pd^{II} catalyst **65**. Simultaneously, the excited Ir photocatalyst in its triplet state facilitates the sensitization of *E*-olefin **68** to its triplet state **69** through energy transfer process. This energetically uphill step drives the conversion of the *E*-olefin **68** to its thermodynamically less stable (*Z*)-product **63**. (**Scheme 8b**).

a) photoredox/Pd-catalyzed decarboxylative alkenylation

b) proposed mechanism



Scheme 8 Photoredox/Pd-catalyzed decarboxylative alkenylation

In Minisic-type reaction research belong to type III, Glorius⁹⁵ employed excess carboxylic acid 72 to react with heterocycles 73 under irradiation of visible light in the presence of a combination of Ir-cat 17 and peroxysulfate, generating alkylated heterocycles 74 (Scheme 9a). Afterward, a facile procedure for the expedient alkylation of electron-deficient N-heterocycles 76 with alkyl carboxylic acids 75 was developed. It proceeded under mild and metal-free conditions that relied upon visible light irradiation, albeit in the absence of a photocatalyst and acid additive (scheme 9b).⁹⁶

a) photo-induced, Ir-catalyzed Minisci-type reaction

b) photocatalyst-free photo-induced Minisci-type reaction

Scheme 9 Photoinduced external oxidants-required Minisci-type reaction

1.2.1.2 Reactions involving redox-active esters

Redox-active esters (RAEs) of carboxylic acids such as N-(acyloxy) phthalimides (NHPI esters) were first introduced to photochemistry by Okada⁹⁷ as a substitute for the Barton ester.⁹⁸⁻¹⁰¹ These are readily prepared from a wide range of carboxylic acids and *N*-hydroxyphthalimide under standard coupling conditions. Visible-light-induced photocatalyzed decarboxylative coupling of redox-active esters provides a novel and versatile route to construct functionalized molecules^{56,102} In this aspect, three distinct catalytic pathways are concluded based on the role of the radical formed and necessity for the inclusion of an additional reductant in the catalytic cycle:

i. visible-light induced photocatalysis with additional reductants

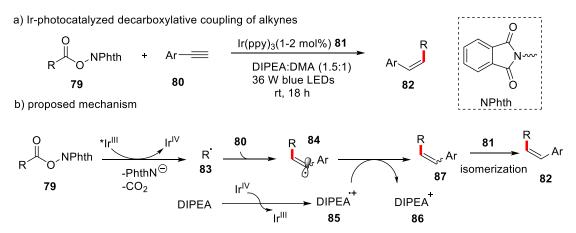
In this case, visible-light-activated photocatalyst facilitates a chemical transformation, with the help of an external reductant to drive the catalytic cycle forward. The reductant plays a crucial role in regenerating the photocatalyst and/or providing electrons for the reaction, enabling the overall process to proceed efficiently.

ii. visible-light-induced dual catalysis

By the merger of photocatalysis and another catalysis, this approach leverages the complementary reactivity of both catalytic systems, often resulting in chemical transformations that would be challenging or inefficient with a single catalyst.

iii. visible-light-induced photocatalysis with radical intermediates as the reductant

The key alkyl radical or derived radical, generated during the photocatalytic cycle, serves as the reductant to transfer electrons to the photocatalyst species or other intermediates This results in their oxidation to carboncationic species, which are subsequently trapped by nucleophiles to form the final product. It is also a photo-mediated redox-neutral process and eliminate the need for external reductants by relying on in situ generated radicals, thereby making the process more self-sustaining and efficient.



Scheme 10 Ir-photoredox-catalyzed decarboxylative coupling with alkynes and its proposed mechanism

A representative example of **pathway i** is reported by Luo and co-workers.¹⁰³ They developed a visible-light-mediated Ir(ppy)₃-catalyzed decarboxylative coupling of NHPI ester **79** with terminal aryl alkyne **80**, enabling the stereoselective synthesis of thermodynamically disfavored (*Z*)-alkene **82** (**Scheme 10a**). This approach features a broad substrate scope with no apparent sensitivity to the electronic properties and substitutional pattern of substrates. In the postulated mechanism, the SET reduction of NHPI ester **79** by photoexcited *Ir^{III} generates alkyl radical **83** and oxidized Ir^{IV} species. Subsequently radical addition of **83** to alkyne **80** at the terminal position forms vinyl radical **84**. Concurrently, DIPEA reduces Ir^{IV} species to regenerate Ir^{III} photocatalyst while it is oxidized DIPEA⁺⁺ **85**. Then the hydrogen atom transfer from **85** to **84** affords alkylated alkenes **87**, which further isomerizes selectively to less stable (*Z*)-isomer **82** in the presence of photocatalyst Ir(ppy)₃ **81**. (**Scheme 10b**)

Some other decarboxylative reactions via **pathway i** are summarized in **Scheme 11**, different radical acceptors such as alkynyl phenyl sulfone **88**,¹⁰⁴ electron-withdraw alkene **90**,¹⁰⁵ 3-cyanocoumarin **92**,¹⁰⁶ *N*-sulfinaldimines **94**,¹⁰⁷ were demonstrated as competent substrates for coupling with NHPI esters under the combination action of the photocatalyst and the reductant (**Scheme 11**). These reactions yield C–C coupling products with high efficiency, highlighting the versatility and utility of this methodology.

Scheme 11 Photoredox-catalyzed decarboxylative reactions with NHPI esters via pathway i

A representative example via **pathway ii** was reported in 2017 by Fu and co-workers, 108 they developed a metallaphotoredox-mediated protocol for the coupling of NHPI ester of N,N-dialkyl- α -amino acids 96 with terminal alkyne 97 bearing aliphatic or aromatic group, leading to synthetically useful propargylamines 98 (Scheme 12a).

One year later, the Fu and Shang group developed an irradiation-induced palladium-catalyzed decarboxylative Heck reaction between vinyl arenes and NHPI esters. ¹⁰⁹ In this method, Pd catalyst Pd(PPh₃)Cl₂ in combination with 4,5-bis(diphenylphosphino)-9,9-dimethylxanthene (Xantphos) under irradiation plays the role of traditional photocatalyst, inducing single-electron transfer to activate N-(acyloxy)phthalimides **99**. Accompanied Pd-catalyzed insertion and β -H elimination process, diverse secondary, tertiary, and quaternary carboxylates, including α -amino acid derived esters were efficiently transformed to substituted styrene **101** with a high ratio of Z/E (**Scheme 12b**).

Scheme 12 Photoredox-catalyzed decarboxylative reactions with NHPI esters via pathway ii

A classical reaction associated with **pathway iii** is decarboxylative Minisci-type reaction. In 2018, Sherwood and co-workers described a one-pot process for decarboxylative Minisci-type reaction using generated NHPI esters in situ with quinolines **103**. ¹¹⁰ The organic photocatalyst 4CzIPN was identified as highly suitable for this one-pot reaction and only near-stoichiometric levels of radical precursor was required, thereby enhancing the method's cost efficiency and practicality (**Scheme 13**) However, partly reactions proceeding at 48 °C resulted in varying amount of bis-alkylated heteroarenes including pyridine, quinoline and phthalazine.

Later, Shang and Fu discovered that a complex of PPh₃ and NaI under blue LEDs irradiation could efficiently catalyze the decarboxylative Minisci-type addition of NHPI esters to heteroarenes. ¹¹¹ This phosphine-iodide-based photoredox system offers significant advantages for industrial applications and large-scale synthesis due to its low cost compared to traditional dye-based or metal-based photocatalysis. In their study, the NHPI esters **102** of various alkyl carboxylic acids were treated with heteroarene **105** in the presence of catalytic PPh₃ and NaI, along with TFA as an acid additive, in acetone under blue LED irradiation, yielding the alkylated *N*-heteroarene **107** with high efficiency (up to 97% yield). Notably, replacing TFA with chiral biphenyl-based phosphoric acid **106** enabled the enantioselective synthesis of heteroaryl-substituted amines, marking a significant advancement in asymmetric Minisci-type alkylation (**Scheme 13**). These findings highlight the potential of this EDA complex system in the development of cost-effective and sustainable methodologies for organic synthesis.

Through **pathway iii** certain alkenes were proved to be efficiently partners to couple with NHPI esters, providing a mild method to construct alkylated alkenes via radical addition or heterocycles via radical cascade cyclization. (**Scheme 13**)¹¹²⁻¹¹⁵ Additionally, *N*-hydroxybenzimidoyl chloride (NHBC) esters **117** as an alternative of NHPI esters were applied to photoredox decarboxylative Minisci-type coupling with heteroarenes. (**Scheme 13**)¹¹⁶ Interestingly, this method obviates the need for any acidic additives, distinguishing it from the majority of Minisci-type reactions that

typically proceed under redox-neutral conditions and necessitate the protonation of *N*-heteroarenes to facilitate radical addition. This result expanded the scope of Minisci-type reactions, offering new opportunities for the development of acid-free methodologies in radical coupling chemistry.

Scheme 13 Photoredox-catalyzed decarboxylative reactions with NHPI esters via pathway iii

1.2.2 Aroyl radical generation via PIII-assisted deoxygenation

The Mitsunobu reaction 117-119 is a widely recognized method in modern organic synthesis for the substitution of hydroxyl groups in alcohols or carboxylic acids. This reaction utilizes the PPh₃/diethyl azodicarboxylate (DEAD) system to overcome the inherent limitation of the hydroxyl group as a poor leaving group. Activation of the hydroxyl group is achieved through the formation of acyloxyphosphonium intermediates via phosphorus(III) reagents. (**Scheme 14a**) Despite its utility, the Mitsunobu reaction has significant drawbacks. It requires the use of stoichiometric amounts or slight excesses of DEAD, a toxic and potentially explosive oxidant, making it less suitable for large-scale synthesis. Furthermore, the process inevitably generates hydrazine derivatives as by-products, contributing to waste. Therefore, the development of more atom-economic and practical methods for the efficient conversion of hydroxyl groups remains a highly desirable goal in organic synthesis. In recent years, photoredox and PIII-mediated deoxygenative reactions provide a powerful platform to achieve hydroxyl-containing substrates conversion. The general mechanism involves the SET oxidation of PIII reagent I to phosphorous radical cation II. Subsequently this intermediate II combines with carboxylate salt to generate phosphoranyl radical III. Then two different processes

could occur. In the first pathway, the intermediate III experience β -selective C-O bond cleavage to product acyl or alkyl radical, followed by trapped by reaction partners to afford coupling products. In the second pathway, species III is oxidized to further phosphonium cations VI, then undergoes nucleophilic substitution to form target compounds VII. (Scheme 14b) Representative examples of these transformations are summarized here, highlighting the versatility and utility of this approach.

a) The classic Mitsunobu reaction

b) General mechanism of photocatalyzed P^{III}-reagent-mediated deoxygenative reactions

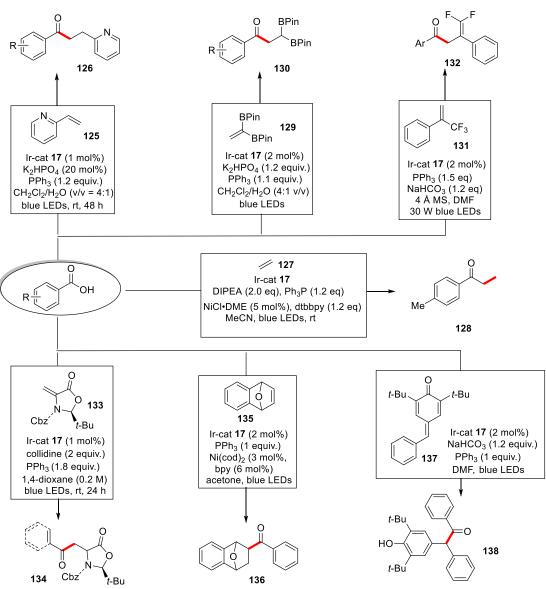
Scheme 14 P^{III} reagent-mediated deoxygenative reactions

In order to synthesize a wide range of $C(sp^2)$ - $C(sp^3)$ coupling products, in 2018,¹²² Zhu and Xie group made a great advancement for the streamlined synthesis of aromatic ketones from feedstock chemicals (**Scheme 15a**). In the optimized conditions, Ir-cat **17** as photocatalyst, K_2HPO_4 as base, and PPh₃ as additive in CH_2Cl_2/H_2O (v/v=4:1) irradiated by blue LEDs were chosen. Commercially cheap aromatic carboxylic acids reacted smoothly with conjugate alkenes such as 2-vinyl pyridines **125**, styenes and electron-deficient alkenes, affording structurally diverse ketones **126** in moderate to good yields. Notably, this synthetic robustness was supported by the late-stage modification of several pharmaceutical compounds and complex molecules. In the propose mechanism visible-light photoredox catalysis facilitates the direct deoxygenation of acids as acyl sources with PPh₃, representing a distinct perspective on radical activation mode.

Afterwards, photoredox/nickel-catalyzed hydroacylation of ethylene 127 with aromatic acids was also reported by Xie and coworkers, 123 proving a general, practical and scalable method to prepare high-value-added aromatic ketone 128. Beyond ethylene 127, other alkenes like vinyl boronic esters 129, 124 α -trifluoromethyl alkene 131, 125 Karady—Beckwith alkene 133, 126 oxabenzonorbornadiene 135, 127 and p-quinone methides 137 128 featuring a merger of alkenyl and carbonyl moieties, were found to be suitable radical acceptors in deoxygenative coupling. These transformations offered wide acyl products with good yields under mild conditions (Scheme 15a).

Moreover, several carboxylic acid derivatives including benzoyl fluorine 139¹²⁹ and carboxylic acid anhydrides 142¹³⁰ effectively engaged in PPh₃-mediated deoxygenative reactions. These reactions facilitated the regioselective diacylation of alkenes, further broadening the scope of accessible acylated products. Additionally, the Tobisu¹³¹ group demonstrated a synergistic photoredox and nickel-catalyzed method for converting allylic esters 146 into the corresponding ketones 148 or 149 through the formal deletion of an oxygen atom using PPh₃. This innovative approach highlighted new opportunities for synthesizing acyl products, though challenges such as site-selectivities need further investigation (Scheme 15b).

a) photoredox PPh₃-mediated deoxygenative coupling of aryl carboxylic acids for constructing C(sp²)-C(sp³) bond



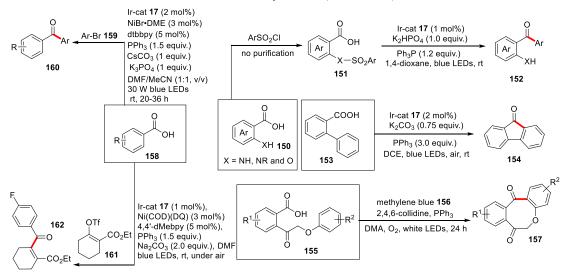
b) photoredox PPh₃-mediated deoxygenative coupling of aryl carboxylic acid derivatives for constructing C(sp²)-C(sp³) bond

Scheme 15 Photoredox-catalyzed deoxygenative reactions of carboxylic acids and derivatives for the construction of $C(sp^2)$ - $C(sp^3)$ bond

For the constructing of C(sp²)-C(sp³) coupling products, photocatalyzed P^{III}-mediated

intramolecular deoxygenative reaction provide a viable approach. For examples, *ortho*-substituted benzoic acids **150**,¹³² **153**,¹³³ and **155**¹³⁴ were discovered to prepare asymmetric diaryl ketones **152**, fluorenones **154**, eight-membered dibenzocycloketones **157** respectively. (**Scheme 16**) Although these diaryl ketone products were able to be gained efficiently, limited substrate scope restricts their broader applicability in synthetic chemistry.

With the continued advancement of metallaphotoredox system in modern organic synthesis, the Xie group in 2020 reported an upgrading protocol for ketone synthesis using commercially available carboxylic acids **158** and organohalides **159** (**Scheme 16**). The combined action of Ir-cat **17** [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆, nickel catalyst NiBr·DME and phosphoranyl radical allows for concise synthesis of highly functionalize diaryl ketones or arylketones. Notably, this approach eliminates the need for pre-activation of carboxylic acids or the use of organometallic reagents, which are typically required in conventional protocols such as the Weinreb ketone synthesis, thereby representing this strategy practicality. Building on this work, in 2022, they introduced vinyl triflates **161** to the PPh₃-mediated deoxygenative coupling of carboxylic acids under similar Ir-photoredox/Ni-catalyzed conditions. This approach enabled the highly selective synthesis of all-carbon tetrasubstituted olefins under mild conditions, even in the presence of significant steric hindrance and uncontrolled *Z/E* stereoselectivity issues. (**Scheme 16**)



Scheme 16 Photoredox-catalyzed deoxygenative reactions of carboxylic acids for the construction of $C(sp^2)$ - $C(sp^2)$ bond

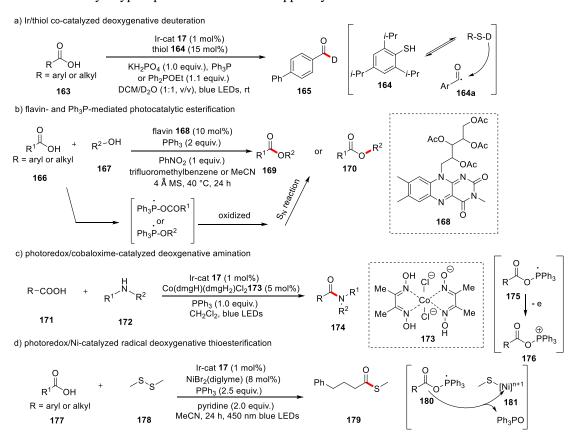
In 2018, Xie group developed a synergistic catalysis strategy for deoxygenative deuteration of carboxylic acids with $D_2O.^{137}$ The optimized reaction conditions [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ 17 as the photocatalyst, 2,4,6-triisopropylbenzenethiol 164 as the HAT catalyst, Ph₃P or Ph₂POEt as an oxygen-atom transfer reagent, and CH₂Cl₂/D₂O (1:1, v/v) as the solvent. Under these conditions, the desired deuterated aldehydes 165 could be prepared in good yields with high levels of D incorporation (Scheme 17a). This precise and efficient deoxygenation strategy shows great promise for the late-stage deoxygenative deuteration of natural product derivatives and pharmaceutical compounds. The method's ability to achieve high deuterium incorporation under mild conditions makes it particularly valuable for applications in drug development and isotopic labeling studies.

Aa a complement to classic Mitsunobu reaction, in 2018 Cibulka and co-workers reported an organophotocatalyzed azodicarboxylate-free esterification of carboxylic acids. ¹³⁸ This protocol

relied on flavin 168 (3-methyl riboflavin tetraacetate) as photocatalyst, PPh₃ as deoxygenative reagent and PhNO₂ as sacrificial oxidant under blue LEDs irradiation in the presence of 4Å molecular sieves (MS). The proposed mechanism involved alcohol nucleophilic substitution to crucial reactive phosphonium species. (Scheme 17b)

The utilization of secondary amines as nucleophiles in PPh₃-mediated photoredox-catalyzed deoxygenation coupling was developed by Zhao group in 2022 (**Scheme 17c**). The combination of Ir-photocatalyst **17** and cobaloxime catalyst (Co(dmgH)(dmgH₂)Cl₂) **173** enables the catalytic generation of acyloxyphosphonium ions **176** for subsequent nucleophilic additions. This method provides a practical route to amines due to the avoidance of stoichiometric oxidants, making it possible for late-stage amidation of drug molecules.

Recently, Glorius, and Qi, and Temps group developed a radical thioesterification via nickel-catalyzed sensitized electron transfer.¹⁴⁰ Under the conditions of Ir-cat **17** as photocatalyst, NiBr₂(diglyme) as metal catalyst, PPh₃ as additives and pyridine as base in MeCN under irradiation of blue LEDs, diverse carboxylic acids could undergo deoxygenative coupling with dimethyl disulfide **178**, constructing C-S products with good efficiency (**Scheme 17d**). The mechanism involves the acyloxyphosphorous radical **180** trapped by Ni adduct **181**.



Scheme 17 Photoredox-catalyzed deoxygenative reactions of carboxylic acids for the construction of C-D and C-heteroatom bond

1.3 Photocatalyzed radical generation via 1,5-HAT

The selective and efficient functionalization of specific C(sp³)-H bonds has long been a significant objective in synthetic chemistry, owing to their substantial synthetic utility. 141-143 However, achieving this remains challenging due to the intrinsically low reactivity of these bonds. 144 While transition-metal (TM)-catalyzed C-H activation is well-established, 145-146 radical-mediated hydrogen atom transfer (HAT) strategies have emerged as a compelling alternative. 147-149 These methods enable selective functionalization of remote C(sp³)-H bonds in complex molecules under relatively mild conditions. Early examples of 1,5-hydrogen atom transfer (1,5-HAT) include the Hofmann-Löffler-Freytag (HLF) and Barton reactions, both of which involve the generation of nitrogen- or oxygen-centered radicals. 150-152 These radicals undergo intramolecular 1,5-HAT to form carbon-centered radicals, which are subsequently functionalized, providing a regioselective approach to modify remote C(sp³)-H bonds. Despite being discovered in the late 19th century, the utility of 1,5-HAT with heteroatom-centered radicals for such transformations remained limited for decades due to the requirement for harsh reaction conditions. However, recent advancements in photochemistry have revitalized this field. By integrating visible-light photoredox catalysis with 1,5-HAT, diverse methods for the remote functionalization of C(sp³)—H bonds have been developed, greatly expanding the scope and applicability of this approach. 153-154 In this section, representative 1,5-HAT reactions triggered by carbon-centered radical are summarized

1.3.1 Aryl radical-mediated 1,5-HAT

In 2016, Xu and co-workers reported an intramolecular radical cascade cyclization from 2-iodoacetanilides **182** under photocatalytic conditions.¹⁵⁵ The key step involved a 1,5-HAT process occurring at the transient aryl radical intermediate **182a**, which was generated through SET reduction by photoexcited *Ir^{III}. This step led to the formation of an alkyl radical **182b**, which subsequently underwent cyclization to afford the desired oxindole products **183** (**Scheme 18a**). By utilizing this Ir-photocatalyzed system mediated by 1,5-HAT, the intermolecular coupling reactions involving 2-iodoarenes were also demonstrated to be feasible. These reactions enabled the construction of diverse coupling products, including C(sp³)–C(sp²),¹⁵⁶⁻¹⁵⁷ C(sp³)–C(sp³),¹⁵⁸ C(sp²)–C(sp²),¹⁵⁹ and C(sp³)–P¹⁶⁰bonds, thus showcasing the broad applicability of this methodology. In 2020, Gevorgyan group introduced a visible-light-mediated Pd-catalyzed intramolecular C-H arylation of acetanilides **184** to synthesize oxindoles **185**. ¹⁶¹ The reaction features a key 1,5-HAT of hybrid aryl palladium intermediates **184a** to **184b**. Interestingly, aryl triflates **184** were utilized as precursors for the formation of the hybrid aryl palladium radical intermediates, although the precise mechanism governing this transformation has yet to be fully clarified (**Scheme 18b**).

Based on this pioneered work, ¹⁶¹ Chen, Yang et al devoted many efforts to this dual

photoredox/palladium-catalyzed remote functionalization. In their research, this aryl-to-alkyl radical relay strategy was effectively applied to a variety of transformations, including desaturation of amides, ¹⁶² radical (cascade domino) Heck coupling, ¹⁶³⁻¹⁶⁴ desaturation/sulfonation cascade, ¹⁶⁵ radical relay formal [5+2] reactions respectively ¹⁶⁶. These contributions have provided valuable insights into photoinduced palladium catalysis, significantly expanding the scope of this field.

In 2022, Gevorgyan reported that cleavage of C-I bond of amines **186** could be induced by blue LEDs irradiation of EDA complex formed between amines **186** and B₂cat₂ (2,2'-bis-1,3,2-benzodioxaborole). The following 1,5-HAT and coupling with another B₂cat₂ lead to highly regio-and diastereo-selective synthesis of valuable α-amino-boronates **187** (**Scheme 18c**). Remarkably,

this protocol provides a general, mild, transition-metal and strong-base-free method to achieve radical α -C(sp³)-H borylation of aliphatic amines, enabling it employed in late-stage functionalization of structurally complex amines. ¹⁶⁷

In 2023, Chen and co-workers reported an EDA complex strategy for the photoactivation of unprotected *o*-anilide aryl chlorides/iodides **188**. ¹⁶⁸ In this method, toluene anion or *t*-BuOK forms an EDA complex with chloride, then blue light trigger of the EDA complex enabled the synthesis of heterocycles **189** (Scheme **18d**).

Recently, Guo and Zhang group presented photoinduced EDA-enabled $C(sp^3)$ -H alkenylation of amines using *t*-BuOLi as electron donor under ambient air. ¹⁶⁹ This strategy offered a general, operationally simple, and transition-metal free route to prepare prevalent and synthetically valuable allylic amines 192 in excellent E/Z and diastereoselectivities (Scheme 18e).

a) visible-light-mediated Ir-photocatalyzed 1,5-HAT reactions of aryl iodides

b) visible-light-mediated Pd-catalyzed 1,5-HAT reaction of aryl triflates

c) photoactivated metal-free radical α -C-H borylation of aliphatic amines

d) visible-light-mediated 1,5-HAT of aryl chlorides and aryl iodides

e) photoinduced EDA-complex-enabled α -C(sp³)-H alkynylation of amines

Scheme 18 1,5-HAT of aryl radicals

1.3.2 Vinyl radical-mediated 1,5-HAT

In 2019,¹⁷⁰ Zhu group reported a method for regioselectively incorporating alkenyl groups into aliphatic sites under Ir-photocatalyzed conditions. The pathway for this remote $C(sp^3)$ -H bond vinylation involves the CF_3 · generation by the SET reduction of Togni's reagent 195. The following CF_3 · addition to propargyl alcohols 194 product vinyl radical 197, Then 1,5-HAT of 197 form alkyl

radical species **198**, which undergoes intramolecular radical addition to C-C double bond to access five-membered ring intermediate **199**. Accompanied with ring-opening rearrangement and subsequent SET oxidation via photocatalyst, fluoroalkylated alkenes **196** could be obtained. The reaction demonstrates high product diversity and synthetic efficiency, enabling the synthesis of a wide range of synthetically valuable *E*-alkenes featuring tri-, di-, or mono-fluoromethyl as well as perfluoroalkyl groups. (**Scheme 19a**)

Subsequently, the same group demonstrated site-selective remote $C(sp^3)$ -H bond vinylation using sulfonyl chloride as radical precursors. A broad scope of α,β -unsaturated sulfones were accessed under mild conditions.¹⁷¹

In 2019, Zhu and co-workers developed an efficient way to synthesize a variety of fluoroalkylated cyclic ketones **203** from alkynyl aldehydes **201** and bromofluoroalkanes **202**. Two selective HAT processes under visible-light irradiation are the key of this transformation. On the one hand, the 1,5-HAT of alkenyl radical **204** provides formyl radical **205**, which then occurs a 5-exo-trig cyclization to afford alkyl radical **206**. On the other hand, a polarity-matched intermolecular HAT from nucleophilic 2-CH of THF to the electrophilic α -perfluoroalkyl carbon radical **206** provides targeted compounds **203**. (Scheme **19b**)

Later, Sun and co-workers reported similar HAT process presented at the coupling of 2-alkynylarylethers 207 and sodium sulfinate 208 under photocatalytic conditions. This domino approach enabled the synthesis of diverse sulfonyl substituted dihydrobenzofurans 209. (Scheme 19c)¹⁷³

In 2020, Chu group reported a three-component decarboxylative coupling via Ir/Ni dual catalysis.¹⁷⁴ Treatment of cyclic oxalates **210**, aryl halide **211** and aliphatic alkyne **212** in the presence of photocatalyst Ir-cat **17**, nickel catalyst NiCl₂(Py)₄, ligand dtbbpy and additive bis(4-methoxyphenyl)methanone in DMSO irradiated by blue LEDs, providing a wide range of site-selective 1,3-difunctionalized cycloalkanes **213** in one single operation. The decarboxylative vinylation/C-H arylation relies on the formation of crucial alkyl radical **215** via 1,5-HAT of addition product **214**. And the subsequent Ni^I complex **216** from **215** and Ni catalyst facilitates aryl coupling. (Scheme **19d**)

In addition, Gevorgyan group focused on photocatalytic active palladium species and reported a novel and mild hydrogen atom translocation/atom-transfer radical cyclization cascade of vinyl iodide 217 via Pd(OAc)₂ and DPEphos (bis[(2-diphenylphosphino)phenyl] ether)system. This protocol involves a 1,5-HAT process mediated hybrid vinyl palladium radical intermediates, thus resulting in iodomethyl carbo- and heterocyclic structures 218. (Scheme 19e) 175

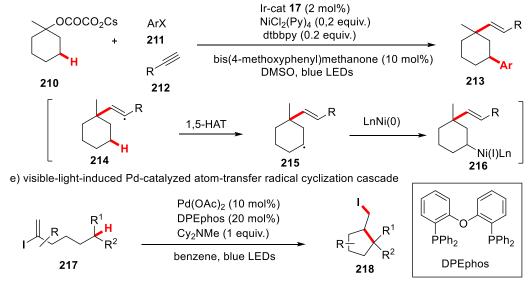
a) remote C(sp³)-H vinylation of propargyl alcohols

OH R³
$$R^3$$
 R^3 R^2 R^3 R^3 R^2 R^3 R^3 R^2 R^3 $R^$

b) alkenyl radical-mediated 1,5-HAT of alkynyl aldehydes

c) synthesis of sulfonylated dihydrobenzofurans

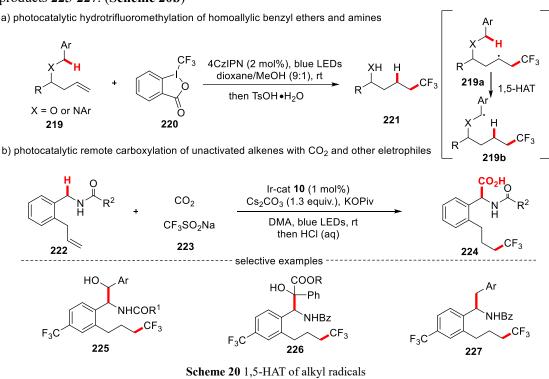
d) photoredox/Ni-catalyzed decarboxylative vinylation/C-H arylation of cyclic oxalates



Scheme 19 1,5-HAT of alkenyl radicals

1.3.3 Alkyl radical-mediated 1,5-HAT

group 2019, Xu described competent metal-free photoredox-catalyzed a hydrotrifluoromethylation of benzyl-protected homoallylic alcohols or amines 219. Valuable δ fluoromethylated free alcohols and amines 221 were delivered with in situ deprotection of benzyl protecting group under mild conditions. The difference of BDE between the benzylic proton of benzyl ethers or benzyl amines (~85 kcal/mol) and the secondary aliphatic C(sp³)-H (~98 kcal/mol) rendered the 1,5-HAT of intermediate 219a to 219b thermodynamically favorable (Scheme 20a)¹⁷⁶. Afterwards Yu¹⁷⁷et al reported remote difunctionalizing carboxylation of unactivated alkenes 222 with CO_2 via photoinduced 1,5-HAT. Other electrophiles, such as aldehydes, α -ketoesters and benzyl bromides, can be used to couple with unactivated alkenes 222 to afford corresponding products 225-227. (Scheme 20b)



1.4 Conclusion

- (1) Carboxylic acids and their derivatives are abundant and cost-effective chemical feedstocks, which have been widely applied in versatile organic synthesis. Among these transformations, decarboxylative reactions triggered by the light and photocatalyst provide a powerful platform to construct diverse molecules under mild conditions. As discussed in the introduction part, redoxneutral photoinduced methods and dual catalysis methods have broaden extensively scope of substrates. Despite the successful development of various coupling products, most decarboxylative reactions relied on metal reagents and additives. The use of metals, particularly rare or expensive ones, along with additives, increases undoubtly the reaction cost and poses challenges for large-scale production. Moreover, residual metal reagents in final products have raised environmental and safety concerns. Therefore, developing green decarboxylative methods remains a longstanding objective in sustainable chemistry. In this context, exploring transition-metal-free and additives-free catalytic systems in decarboxylative reactions is much more significant and requires further research. Especially, applying these practical methods to synthesize pharmaceutically relevant molecule scaffolds is in great demand, as they have great potential for drug design and development.
- (2) Carboxylic acids and their derivatives are privileged chemical entities due to their wide availability, low cost, and structural diversity. Recently, they have been extensively applied in P^{III}-assisted deoxygenation under light irradiation. Satisfactorily, these well-developed methods provide a mild and efficient way to construct carbonyl-containing compounds. However, functional group transformations involving carboxylic acids remain relatively underexplored. Herein, the development of novel reactivity patterns and reaction modes in photocatalyzed deoxygenation is essential, providing more possibilities for using readily available carboxylic acids and their derivatives to the functionalization of molecules especially bioactive molecules.
- (3) Direct and site-selective C(sp³)–H activation has been a longstanding challenging subject due to its intrinsic inert nature. The merger of photoredox catalysis and 1,5-hydrogen atom transfer (1,5-HAT) strategies have emerged as a powerful tool to achieve selectively remote C(sp³)-H functionalization under mild conditions. Although photoinduced HLF-type and Barton-type transformations have been extensively developed, most of these strategies focused on the generation of heteroatom-centered radicals. In contrast, reactions triggered by carbon-centered radicals remain underdeveloped and continued efforts are required in this regard to access more valuable drug-like frameworks.

2 Results and discussion

2.1 NaI/PPh₃-catalyzed visible-light-mediated decarboxylative radical cascade cyclization of *N*-arylacrylamides for the efficient synthesis of quaternary oxindoles

Author contributions: D. L. performed the corresponding experiments and drafted the manuscript. Y. Z. finished the preparation of some starting materials. Prof. Dr. Patureau supervised the project and revised the draft.

This work has been published: Beilstein J. Org. Chem. 2023, 19, 57.

2.1.1 Introduction

Radical-triggered cascade reactions have emerged as a powerful tool in organic synthesis, enabling the formation of multiple carbon-carbon and carbon-heteroatom bonds in one pot. This efficiency makes them particularly valuable for developing new synthetic methodologies, facilitating the construction of complex natural products and pharmaceuticals.¹⁷⁸ Recently, radical-initiated cascade cyclizations involving acrylamides have garnered increasing attention due to their potential for assembling functional oxindole scaffolds. These structures are widely present in functional molecules and bioactive compounds, which play crucial roles in drug discovery (**Scheme 1-1**)¹⁷⁹⁻¹⁸¹. Despite the progress in this area, ¹⁸²⁻¹⁸⁵ existing methods often require stoichiometric reagents ¹⁸⁶⁻¹⁹³ or high reaction temperatures, ¹⁹⁴⁻²⁰³ limiting their practicality and broader application.

In recent years, photocatalytic reactions have been demonstrated as one of the most powerful methods in developing radical-initiated addition/cyclization cascades from acrylamides.²⁰⁴⁻²⁰⁶ Significant advancements have been made in generating diverse radicals from alkyl halides,²⁰⁷⁻²⁰⁹ carboxylic acids,²¹⁰⁻²¹² simple alkanes,²¹³ alkylboronic acids,²¹⁴ and isocyanides,²¹⁵ or other.²¹⁶⁻²¹⁸ In this context, the Fu group reported a Ru(bpy)₃Cl₂-catalyzed synthesis of *N*-Boc proline oxindole derivatives under visible-light irradiation, utilizing *N*-hydroxyphthalimide (NHPI) esters as efficient alkyl radical precursors.²¹² Similarly, in 2015, Cheng and colleagues developed a visible-light-mediated tandem radical cyclization of *N*-arylacrylamides with *N*-(acyloxy)phthalimides, producing 3,3-dialkylated oxindoles in the presence of [Ru(bpy)₃Cl₂]·6H₂O.²¹¹ However, the reliance on noblemetal photocatalysts and the limited substrate scope have restricted the broader application of these approaches.(**Scheme 1-2**)

Scheme 1-1 Representative examples containing oxindole moiety

Scheme 1-2 Photocatalytic decarboxylative radical cascade cyclization via transition-metal catalysis

As the field of green chemistry continues to advance, there is a growing demand for cost-effective and transition-metal-free photocatalysis. In 2019,¹¹¹ Fu and Shang introduced a pioneering photocatalytic system based on a NaI/PPh₃ electron donor-acceptor (EDA) complex, enabling decarboxylative alkylation of silyl enol ethers and *N*-heteroarenes. Compared to conventional radical-based methods, this catalytic system offers several advantages: it eliminates the need for external redox additives and noble metals, relies on inexpensive and readily available and cost-effective reagents NaI/PPh₃, and operates under mild conditions. Building on this foundation, NaI/PPh₃ catalysis has since been applied to various transformations, including alkene functionalization, decarboxylative C(sp³)–X bond formation, and cyclization of 1,7-enynes.²¹⁹ Despite its demonstrated potential, iodide/phosphine catalysis remains somewhat underutilized in organic synthesis, particularly in the preparation of biologically significant oxindole derivatives. Given the broad utility of these molecules in pharmaceuticals and bioactive compounds, ¹⁷⁹⁻¹⁸¹ further exploration of iodide/phosphine-based radical methodologies could unlock new opportunities for their efficient and sustainable synthesis.

2.1.2 Condition optimizations

Initially, *N*-arylacrylamide **1-1a** and redox-active ester **1-2a** were chosen as model substrates to react for 36 h under 456 nm blue LEDs irradiation and N₂ atmosphere. Key results of screening iodides are summarized in **Table 1-1**. Surprisingly, NaI delivered desired oxindole derivative **1-3aa** with 72 % isolated yield (entry 1). When changing NaI to other iodides, LiI, KI, RbI, CsI and CaI₂ were all effective catalysts at providing **1-3aa**, albeit in slightly lower yields (entries 2-6). In addition, the oxindole product **1-3aa** could be afforded with moderate yield in the presence of *n*-Bu₄NI (entry 7). It should be noted that all tested iodides were found to be soluble under those conditions.

Table 1-1 Screening of iodide sources^a

| entry | iodide | 1-3aa , yield (%) ^b |
|-------|--------|---------------------------------------|
| 1 | NaI | 76 (72)° |
| 2 | LiI | 70 |
| 3 | KI | 62 |
| 4 | RbI | 64 |
| 5 | CsI | 39 |

| 6 | CaI ₂ | 56 |
|---|------------------------------------|----|
| 7 | $n	ext{-}\mathrm{Bu}_4\mathrm{NI}$ | 57 |

^aUnless otherwise noted, the reaction conditions were as follows: **1-1a** (0.3 mmol), **1-2a** (0.2 mmol), iodide (0.04 mmol), and PPh₃ (0.04 mmol) in MeCN (2 mL) irradiated by 456 nm blue LEDs for 36 h at room temperature. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

Then different phosphines were screened (**Table 1-2**). Aromatic phosphines including PPh₃, tris(4-fluorophenyl)phosphine $P(4-F-C_6H_4)_3$, tris(4-methoxyphenyl)phosphine $P(4-OMe-C_6H_4)_3$ performed well, the cheapest PPh₃ remaining however optimal (entries 1-3). In contrast, tricyclohexylphosphine PCy₃ was tested and performed poorly (entry 4), and bulky tri-o-tolylphosphine P(2-Me-C₆H₄)₃ almost shut down the reaction (entry 5). These results suggest that the accessibility of the phosphorus center plays a crucial role.

Table 1-2 Screening of phosphine source^a

| entry | phosphine | 1-3aa , yield (%) ^b |
|-------|---------------------|---------------------------------------|
| 1 | PPh ₃ | 76 (72)° |
| 2 | $P(4-F-C_6H_4)_3$ | 73 |
| 3 | $P(4-OMe-C_6H_4)_3$ | 60 |
| 4 | PCy_3 | 23 |
| 5 | $P(2-Me-C_6H_4)_3$ | trace |

^aUnless otherwise noted, the reaction conditions were as follows: **1-1a** (0.3 mmol), **1-2a** (0.2 mmol), NaI (0.04 mmol), and phosphine (0.04 mmol) in MeCN (2 mL) irradiated by 456 nm blue LEDs for 36 h at room temperature. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

In further condition optimation (**Table 1-3**), replacing acetonitrile (MeCN) with dimethyl sulfoxide (DMSO), or dimethylacetamide (DMA) or acetone, or ethyl acetate (EA), led to inferior yields (entries 1-5), and no product was detected using 1,4-dioxane and CH₂Cl₂ as reaction solvents (entries 6-7). Control experiments indicate both PPh₃ and irradiation are essential for this decarboxylative tandom cyclization (entries 9-10). Although the reaction proceeded without NaI, it resulted in a low yield of **1-3aa** (entry 8).

Table 1-3 Screening of solvents and control experiments^a

| entry | variation from standard conditions | 1-3aa , yield (%) ^b |
|-------|------------------------------------|---------------------------------------|
| 1 | MeCN | 76 (72)° |
| 2 | DMSO instead of MeCN | 60 |
| 3 | DMA instead of MeCN | 44 |
| 4 | acetone instead of MeCN | 52 |

| 5 | EA instead of MeCN | 57 |
|----|---|------|
| 6 | CH ₂ Cl ₂ instead of MeCN | N.D. |
| 7 | 1,4-dioxane instead of MeCN | N.D. |
| 8 | without NaI | 14 |
| 9 | without PPh ₃ | N.D. |
| 10 | without blue LEDs | N.D. |

^aUnless otherwise noted, the standard reaction conditions were as follows: **1-1a** (0.3 mmol), **1-2a** (0.2 mmol), NaI (0.04 mmol), and PPh₃ (0.04 mmol) in MeCN (2 mL) irradiated by 456 nm blue LEDs for 36 h at room temperature. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

2.1.3 Substrate scope studies

With the optimized conditions established, a diverse range of acrylamides featuring various substituents was synthesized and evaluated. Under standard conditions, numerous acrylamides demonstrated excellent compatibility, yielding the desired oxindoles in moderate to good efficiencies (**Scheme 1-3**). Electron-donating groups, such as methyl and methoxy, positioned at the *para*-location of phenyl rings slightly reduced reaction activity, yet the corresponding products were still obtained in 68% and 66% yield, respectively (**1-3ba** and **1-3ca**). When these substituents were replaced with common halogens or electron-withdrawing groups, they all reacted smoothly with redox-active ester **1-2a** to afford the target oxindoles (**1-3da-1-3ga**) in good yields. Notably, the presence of a trifluoromethyl group enhanced performance, delivering **1-3fa** in very high 85% yield. Furthermore, *ortho*-substitution on the *N*-aryl moiety was well tolerated, though it led to a slight reduction in yield (**1-3ha-1-3ka**, 50%-63%).

Interestingly, a cyclic *N*-arylacrylamide derivative also proved to be a viable substrate, affording the corresponding polycyclic structure **1-3la** in a 67% yield. Variations in the *N*-substituent, including ethyl, benzyl, and phenyl, were well accommodated, providing the expected products (**1-3ma-1-30a**) in good efficiency. A noteworthy observation was the significant impact of substituent modifications at the 2-position of the *N*-arylacrylamide core. Replacing a methyl group with a phenyl ring led to a substantial drop in yield from 72% to 34% (**1-3pa**), highlighting the steric and electronic influence of the substituent. Encouragingly, substrate **1-1q** successfully underwent decarboxylative cascade cyclization, furnishing **1-3qa** in 70% yield. This compound serves as a key intermediate in the synthesis of (±)-physovenine and (±)-physostigmine alkyl analogues, known for their inhibitory activity against acetylcholinesterase and butyrylcholinesterase. ^{195,220,221} To further broaden the scope of this methodology, a benzamide-derived acrylamide **1-1r** was subjected to the reaction, resulting in the formation of the anticipated six-membered annulated product **1-3ra** with a commendable yield of 66%.

^aReaction conditions: **1-1** (0.3 mmol), **1-2** (0.2 mmol), NaI (0.04 mmol), and PPh₃ (0.04 mmol) in MeCN (2 mL) irradiated by 456 nm blue LEDs for 36 h at room temperature. Isolated yields are reported.

Scheme 1-3 Scopes of acrylamides^a

To further explore catalytic efficiency through the decarboxylative radical cascade cyclization, then the scope of redox-active esters was evaluated (**Scheme 1-4**). It was demonstrated that redox-active esters that derived from primary, secondary, and tertiary aliphatic carboxylic acids were all well-suited to this approach. Cyclic substrates containing cyclobutyl, cyclopentyl, and indenyl groups successfully yielded the corresponding products in good efficiency (**1-3ab-1-3ad**, 63–74%), whereas the incorporation of an adamantyl-derived substituent posed greater difficulty, affording **1-3ae** in a moderate 40% yield. Additionally, cyclic systems featuring oxygen- or nitrogen-containing rings proved effective, delivering the target oxindoles (**1-3af-1-3ah**) in yields ranging from 65% to 76%. Furthermore, a symmetrically α -substituted redox-active ester led to the formation of quaternary oxindole **1-3ai** in 69% yield, while an asymmetrically α -branched precursor underwent the reaction with comparable efficiency, generating oxindole **1-3aj** as a 1:1.1 mixture of

diastereomers. Notably, this strategy facilitated the synthesis of the sterically demanding oxindole **1-3ak** in good yield using a *tert*-butyl *N*-hydroxyphthalimide ester as the *tert*-butyl radical source. Significantly, a redox-active ester derived from methionine was efficiently transformed into the α-aminoalkylation product **1-3al** in overall 70% yield, demonstrating the method's potential for modifying and derivatizing both natural and synthetic amino acids. Moreover, functional groups such as a terminal alkene **1-3am**, a terminal alkyne **1-3an**, and an alkyl chloride **1-3ao** were well-tolerated, yielding the corresponding products in encouraging efficiencies. To further highlight the versatility of this protocol, we applied it to a complex scaffold derived from lithocholic acid, which underwent smooth decarboxylative cyclization to furnish oxindole **1-3ap** in 63% yield. Finally, it should be noted that benzoyl ester substrate **1-2q** did not deliver the corresponding cyclized product **1-3aq**.

^aReaction conditions: **1-1** (0.3 mmol), **1-2** (0.2 mmol), NaI (0.04 mmol), and PPh₃ (0.04 mmol) in MeCN (2 mL) irradiated by 456 nm blue LEDs for 36 h at room temperature. Isolated yields are reported.

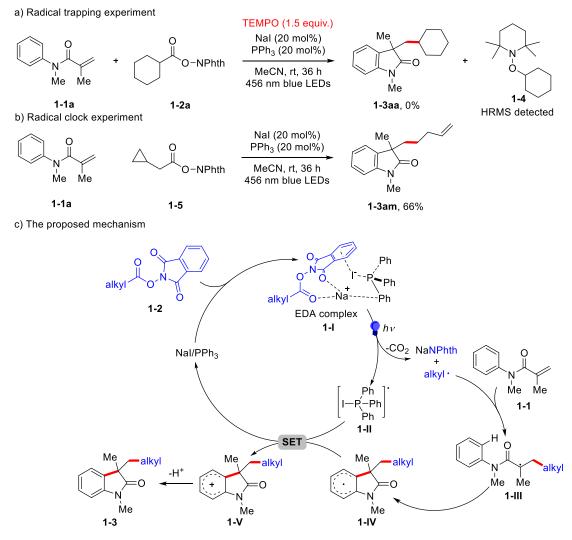
Scheme 1-4 Scope of alkyl radical precursors^a

2.1.4 Mechanistic investigation

To better understand the reaction mechanism, several control experiments were conducted. When a radical scavenger, such as 2,2,6,6-tetramethyl-1-piperidinyloxyl (TEMPO), was introduced into the

catalytic system under standard conditions, the reaction was completely suppressed, and a TEMPO-trapped adduct 1-4 was detected by HRMS (Scheme 1-5a). Additionally, a radical clock experiment using redox-active ester 1-5 in a reaction with acrylamide 1-1a yielded the radical-mediated ring-opening product 1-3am in 66% yield (Scheme 1-5b). These findings strongly suggest that a radical species plays a key role in the NaI/PPh₃-catalyzed decarboxylative cascade cyclization leading to oxindoles.

Based on these experimental results and previous reports, ²¹⁹ a plausible catalytic cycle involving a radical pathway was proposed (**Scheme 1-5c**). The reaction is initiated by photoactivation of a transient electron donor-acceptor (EDA) complex (**1-I**), formed in situ from NaI, PPh₃, and redoxactive ester **1-2a** in MeCN. This activation leads to the generation of an alkyl radical and a PPh₃–I radical species **1-II**, accompanied by the release of carbon dioxide and NaNPhth. The alkyl radical then undergoes addition to acrylamide **1-1**, forming radical intermediate **1-III**. This intermediate undergoes an intramolecular radical C–H functionalization, yielding cyclic intermediate **1-IV**. Subsequent oxidation of the delocalized radical species **1-IV** by **1-II** generates the cationic intermediate **1-V** while simultaneously regenerating the NaI/PPh₃ photocatalyst for the next cycle. Finally, deprotonation of **1-V** produces the desired oxindole product **1-3**.



Scheme 1-5 Control experiments and proposed catalytic cycle of NaI/PPh₃ photoredox catalysis

2.1.5 Conclusion

In this study, we developed an efficient photocatalytic decarboxylative radical cascade cyclization of *N*-arylacrylamides using a range of redox-active esters derived from readily available carboxylic acids. Conducted under mild conditions, this approach offers a practical alternative to conventional transition-metal and organophotocatalysis²²². Notably, the inexpensive and easily accessible NaI/PPh₃ system serves as an effective photoredox catalyst, providing a cost-efficient route to construct oxindole scaffolds featuring a quaternary carbon center.

This methodology is distinguished by its broad substrate scope, high functional group tolerance, and operational simplicity. Mechanistic studies suggested that the reaction proceeds via a radical cascade pathway, further supporting the role of NaI/PPh₃ in facilitating the transformation. We expect that these findings will inspire further exploration of NaI/PPh₃-catalyzed reactions and related radical-based synthetic strategies for functional molecules.

2.2 Visible-light-induced photocatalytic deoxygenative benzylation of

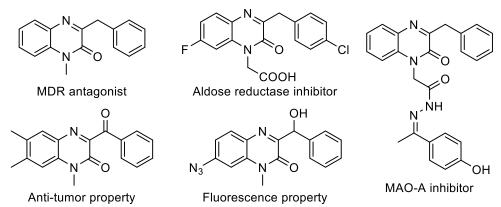
quinoxalin-2-(1H)-ones with carboxylic acid anhydrides

Author contributions: D. L. performed the corresponding experiments and drafted the manuscript. Prof. Dr. Patureau supervised the project and revised the draft.

This work has been published: Org. Lett. 2024, 26, 6841.

2.2.1 Introduction

Quinoxalin-2(1*H*)-ones are an important class of nitrogen-containing heterocycles with broad applications in pharmaceutical and synthetic chemistry. ²²³⁻²²⁵ In particular, 3-benzylquinoxalin-2(1*H*)-ones are commonly found in functional molecules and therapeutic agents, exhibiting diverse activities such as multidrug resistance (MDR) antagonism, aldose reductase inhibition, *N*-type calcium channel blockade, fluorescence property, and monoamine oxidase A (MAO-A) inhibition (**Scheme 2-1**). ²²⁶⁻²²⁹ Given their significance, the efficient synthesis of these compounds has attracted considerable attention in recent years, with substantial efforts focused on direct C3 benzylation via step- and atom-economical C–H bond activation/ functionalization strategies. ²³⁰⁻²³³ However, early methods often required excess peroxides as radical initiators at high temperatures, ²³¹⁻²³³ resulting in harsh reaction conditions and poor tolerance for sensitive functional groups.



Scheme 2-1 Representative compounds bearing the 3-benzylquinoxalin-2-(1H)-one scaffold

Scheme 2-2 Reported benzylating reagents for C3 benzylation of quinoxaline-2(1H)-ones

In recent years, visible-light-induced photocatalysis has emerged as a powerful, mild, and practical approach for constructing C–C and C–heteroatom bonds through radical processes. 30-44 This has led to some progress in the direct C3 benzylation of quinoxalin-2(1*H*)-ones under visible-light irradiation (**Scheme 2-2**). 234-238 In 2020, the Xuan group developed an acetoxybenziodoxole-accelerated benzylation strategy using 4-benzyl-1,4-dihydropyridines as benzyl radical precursors. 234 Later, in 2022, Yang and Adiyala independently demonstrated the use of benzylsulfonium hydrazides and Katritzky salts for the photochemical benzylation of *N*-heteroarenes. 235-236 More recently, the Yang and Yu groups reported direct benzylation using benzyl halides under visible-light photocatalysis. 237-238 Despite these advancements, many of these methods suffer from limitations such as the need for complex benzylation reagents or the use of toxic components. Consequently, developing safer, more efficient strategies for synthesizing 3-benzylquinoxalin-2(1*H*)-ones remains an important goal.

Scheme 2-3 Direct synthesis of C3-functional quinoxaline-2(1H)-ones via SET oxidation

Carboxylic acids and their derivatives are highly valuable synthetic precursors due to their abundance, wide availability, and structural diversity. Recently, they have been extensively explored for the C3 functionalization of quinoxalin-2(1*H*)-ones under visible-light irradiation.²³⁹⁻²⁴⁸ However, most reported methods focus on generating 3-acylquinoxalin-2(1*H*)-one derivatives via single electron transfer (SET) oxidation of acyl radical intermediates (**Scheme 2-3**). Inspired by recent phosphoranyl radical-mediated deoxygenative functionalizations of carboxylic acids, ¹²¹⁻¹⁴⁰ we

envisioned a complementary approach: a photochemical SET reduction of intermediate A, followed by deoxygenative reduction, to efficiently access valuable 3-benzylquinoxalin-2(1*H*)-ones.

2.2.2 Condition optimizations

To evaluate the feasibility of our photochemical reaction design, we selected *N*-methylquinoxalin-2(1*H*)-one **2-1a** and benzoic anhydride **2-2a** as model substrates. Under blue LEDs irradiation in an anhydrous MeCN solvent and N₂ atmosphere, the desired 3-benzylquinoxalin-2(1*H*)-one **2-3aa** was successfully obtained in 77% isolated yield using Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ as the PC, with Ph₃P and DABCO as additives (entry 1). In contrast, alternative photocatalysts such as Ir(ppy)₃, Ru(bpy)₃Cl₂·6H₂O, and 4CzIPN proved ineffective, failing to facilitate the reaction (**Table 2-1** entries 2–4).

Table 2-1 PC screening^a

^aUnless otherwise noted, the reaction conditions were as follows: **2-1a** (0.15 mmol), **2-2a** (0.3 mmol), PC (3%), Ph₃P (250%), DABCO (50%) in MeCN (1.5 mL), irradiation by 40W blue LEDs under N₂ at room temperature for 12 h. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

Then, diverse organic base and inorganic base were investigated (**Table 2-2**). Among them, DABCO proved to be the most effective (entry 1). Substituting DABCO with other bases led to significantly lower yields, while no product formation was observed when TMEDA or DIPEA was used (entries 2–7).

Table 2-2 Base screening

Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (3%)

^aUnless otherwise noted, the reaction conditions were as follows: **2-1a** (0.15 mmol), **2-2a** (0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (3%), Ph₃P (250%), base (50%) in MeCN (1.5 mL), irradiation by 40W blue LEDs under N₂ at room temperature for 12 h. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

Next, various solvents to optimize the reaction conditions was evaluated (**Table 2-3**). Acetone provided a yield comparable to MeCN, whereas reactions conducted in DCE and DMF resulted in

significantly lower yields of 17% and 53%, respectively (entries 2-4). Control experiments confirmed that the photocatalyst, light, Ph₃P, and DABCO were all essential for successful C3 benzylation with benzoic anhydride (entry 5). Additionally, altering the light source yielded suboptimal results. For instance, using a 370 nm wavelength led to diminished efficiency (entry 6), and simply heating the reaction to 60 °C failed to substitute for light irradiation (entry 7). Finally, replacing benzoic anhydride 2-2a with benzoic acid severely reduced the yield, emphasizing the crucial role of the anhydride structure in substrate activation (entry 8).

Table 2-3 Solvent screening and control experiments^a

| entry | variation from standard conditions | 2-3aa, yield ^b |
|-------|---|---------------------------|
| 1 | MeCN as the solvent | 81 (77)° |
| 2 | DCE as the solvent | 17 |
| 3 | DMF as the solvent | 53 |
| 4 | acetone as the solvent | 72 |
| 5 | no light or no PC, or no PPh3 or no DABCO | 0 |
| 6 | $\lambda = 370 \text{ nm}$ | 62 |
| 7 | no light @ 60 °C | 0 |
| 8 | benzoic acid (0.3 mmol) instead of 2-2a | 19 |

^aUnless otherwise noted, the standard reaction conditions were as follows: **2-1a** (0.15 mmol), **2-2a** (0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (3%), Ph₃P (250%), DABCO (50%) in solvent (1.5 mL), irradiation by 40W blue LEDs under N₂ at room temperature for 12 h. ^bThe yield was determined by ¹H NMR analysis of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. ^cisolated yield.

2.2.3 Substrate scope studies

With the optimized conditions in hand, the substrate scope of quinoxalin-2(1*H*)-ones in this photocatalytic reductive benzylation reaction was explored (**Scheme 2-4**). A broad range of quinoxalin-2(1*H*)-ones with varying substituents on the aromatic ring proved to be compatible under standard conditions. Halogens such as fluoro, chloro, and bromo at the C6 or C7 positions were well tolerated, affording the corresponding 3-benzylquinoxalin-2(1*H*)-ones in good yields (**2-3ba, 2-3ca, 2-3da, 2-3ha, 2-3ia,** 72–77%). Likewise, substituents with electron-donating or electron-withdrawing groups yielded the desired products efficiently (**2-3ea, 2-3fa, 2-3ga, 2-3ja**; 62–81%). The benzylation reaction also proceeded smoothly with 8-chloro- and 5-chloroquinoxalin-2(1*H*)-ones, albeit with slightly reduced yields (**2-3ka,** 35% and **2-3la,** 49%). Moreover, 5-methylquinoxalin-2(1*H*)-one demonstrated excellent reactivity, affording **2-3ma** in 76% yield. Additionally, difluoro-, dichloro-, and dimethyl-substituted derivatives underwent the reaction effectively, delivering the target products with high efficiency (**2-3na, 2-3oa, 2-3qa**). However, a dibromo-substituted quinoxalin-2(1*H*)-one yielded only 18% (**2-3pa**), likely due to its limited solubility.

Quinoxalin-2(1*H*)-ones with extended aromatic or heteroaromatic ring produced **2-3ra** and **2-3sa** in 40% and 82% yield, respectively. Furthermore, various *N*-protecting groups, including ethyl, n-butyl, allyl, propargyl, ester, benzoyl, and benzyl, were well accommodated, yielding products **2-3ta-2-3za** in good to excellent yields (69-88%). Notably, this method tolerated an *o*-vanillin moiety, enabling the synthesis of product **2-3aaa**, which holds potential for pharmaceutical applications.

Finally, we successfully applied this strategy to the benzylation of the 6-azauracil-derived heterocycle 2-1ab, achieving an excellent yield of 87% for product 2-3aba.

^aReaction conditions: **2-1** (0.15 mmol, 1 equiv), **2-2** (0.3 mmol, 2 equiv), $Ir[dF(CF_3)ppy]_2(dtbbpy)PF_6$ (0.0045 mmol, 3 mol %), Ph_3P (0.375 mmol, 2.5 equiv), and DABCO (0.075 mmol, 50 mol %) in anhydrous MeCN (1.5 mL) with irradiation by 40 W blue LEDs (456 nm) under N2 at room temperature for 12 h. Isolated yields are reported. ^bReaction scaled to 1 mmol: anhydrous MeCN (10 mL), irradiation for 24 h.

2-3aba, 87%

Scheme 2-4 Scope of quinoxaline-2-(1H)-ones^a

To further assess the versatility of this benzylation reaction, the scope of benzoic anhydride derivatives was examined, as shown in Scheme 2-5. Fluorinated benzoic anhydrides, regardless of whether the fluoro substituent was positioned at the para, meta, or ortho site on the phenyl ring, exhibited good reactivity, yielding the corresponding products efficiently (2-3ab-2-3ad). Interestingly, the *meta*-methyl-substituted anhydride delivered a higher yield (76%) compared to its para- and ortho-methyl counterparts (2-3ae-2-3ag). Additionally, the reaction proceeded well with a trifluoromethyl-substituted anhydride (2-3ah), demonstrating compatibility with strongly electron-withdrawing groups. In contrast, electron-donating groups led to a significant decline in yields (2-3ai, 2-3aj). Moreover, cyclohexanecarboxylic anhydride failed to produce the expected coupling product (2-3ak). Encouragingly, this protocol was successfully scaled up to a 1 mmol reaction, yielding 182 mg of product 2-3aa (73%) after 24 hours of blue-light irradiation (Scheme 2-4; see Part 4.2.2 for details).

^aReaction conditions: same as Scheme 2-4. Isolated yields are reported.

Scheme 2-5 Scope of carboxylic anhydrides^a

2.2.4 Synthetic applications

To further highlight the utility and practicality of this reductive benzylation strategy, we applied it to the synthesis of bioactive molecules derived from simple 3-benzylquinoxalin-2(1*H*)-ones. For example, compound **2-3xa** was treated with hydrazine hydrate in methanol, yielding hydrazide derivative **2-4xa**. Subsequent condensation of **2-4xa** with 4-hydroxyacetophenone led to the formation of MAO-A inhibitor **2-5xa**²²⁹, achieving an overall 75% isolated yield (**Scheme 2-6a**). In another application, quinoxalin-2(1*H*)-one **2-1x** underwent benzylation with anhydride **2-21** under standard conditions, affording product **2-3xl** in 58% yield. Further hydrolysis of **2-3xl** produced derivative **2-6xl** with 79% yield (**Scheme 2-6b**). Product **2-6xl** has previously been investigated for its potential as an aldose reductase inhibitor.²⁴⁹

Scheme 2-6 Synthetic applications

2.2.5 Mechanistic studies

To gain deeper insight into the reaction mechanism, a series of control experiments were conducted. First, when the reaction was performed in deuterated acetonitrile (MeCN- d_3) under standard conditions, no deuterium incorporation was detected in product **2-3aa-d** (**Scheme 2-7a**). However, upon adding 2 equivalents of D₂O to anhydrous MeCN, 62% deuterium labeling was observed at the benzylic position. This finding suggests that trace amounts of water, potentially originating from the reagents or reactants, may serve as a hydrogen source in the reaction. Additionally, the introduction of the radical scavenger 2,2,6,6-tetramethylpiperidin-1-oxyl (TEMPO) significantly

suppressed the reaction, yielding only trace amounts of **2-3aa**, and the benzoyl-TEMPO adduct **2-7a** was successfully isolated and characterized, strongly indicating the involvement of an acyl radical pathway (**Scheme 2-7b**). Interestingly, no evidence of a TEMPO-trapped benzyl radical intermediate was found in the crude reaction mixture. Inspired by previous reports featuring C3 functionalization of quinoxalin-2(1*H*)-ones under visible-light irradiation, ²³⁹⁻²⁴⁸ we synthesized ketone **2-8** and benzyl alcohol **2-9** and subjected them to the standard reaction conditions (**Scheme 2-7c**). Notably, only benzyl alcohol **2-9** was efficiently converted to product **2-3aa**, achieving 86% isolated yield, whereas ketone **2-8** remained unreacted. These observations suggest that benzyl alcohol **2-9** is a plausible intermediate in the transformation. Next, Stern–Volmer fluorescence quenching experiments provided additional mechanistic insights. The iridium photocatalyst exhibited fluorescence quenching in the presence of Ph₃P, quinoxalin-2(1*H*)-one **2-1a**, and DABCO, but not with anhydride **2-2a** (see **Part 4.2.2** for details).

a) Deterium labeling experiments

Scheme 2-7 Mechanistic investigations

Building on the experimental results and previous studies, ²⁵⁰⁻²⁵² we propose the reaction pathway illustrated in **Scheme 2-8**. Initially, the photoexcited *Ir^{III} catalyst abstracts an electron from Ph₃P, generating a phosphine radical cation. Due to its strong affinity for oxygen, this phosphine radical cation facilitates the activation of anhydride intermediate **2-II**. The resulting intermediate undergoes hydrolysis to form intermediate **2-III**, followed by the elimination of phosphine oxide, which generates the corresponding benzoyl radical **2-IV**. This benzoyl radical is then captured by quinoxalin-2(1*H*)-one **2-1a**, leading to the formation of stabilized radical intermediates **2-V** and **2-VI**. Subsequent one-electron reduction of these intermediates yields anionic intermediate **2-VII** and protonated intermediate **2-8**, which we have previously identified as a viable intermediate (**Scheme 2-7c**). This species then reacts with another equivalent of the phosphine radical cation (intermediate **2-VIII**), triggering a second phosphine oxide elimination step to form intermediate **2-IX**. Another

round of one-electron reduction produces intermediate **2-X**, completing this portion of the catalytic cycle. The significant presence of triphenylphosphine oxide was confirmed through ³¹P NMR analysis of the crude reaction mixture, supporting the proposed mechanistic pathway. As for the role of DABCO, it may act as a proton shuttle, consistent with the observation that other bases also facilitate the reaction (**Table 2-2**, entries 2-4). However, its potential function as a redox relay cannot be entirely ruled out.

Scheme 2-8 Proposed mechanism

2.2.6 Conclusion

In summary, we have developed a visible-light-driven photocatalytic deoxygenative benzylation method for the efficient synthesis of 3-benzylquinoxalin-2(1H)-ones. This reaction is characterized by mild, safe conditions and demonstrates excellent functional group compatibility, as well as the broad availability of starting materials. A variety of 3-benzylquinoxalin-2(1H)-ones were synthesized in a single step with generally high yields. Additionally, this approach was successfully applied to the rapid synthesis of two compounds with notable pharmacological potential.

2.3 Visible-light-mediated radical α -C(sp³)-H gem-difluoroallylation

of amides with trifluoromethyl alkenes via halogen atom transfer and

1,5-hydrogen atom transfer

Author contributions: D. L. performed the corresponding experiments and drafted the manuscript. F. X. finished some starting material preparation. Prof. Dr. Oppel and B. E. measured and analyzed crystal structure. Prof. Dr. Patureau supervised the project and revised the draft.

This work has been published: Org. Lett. 2025, 27, 2377.

2.3.1 Introduction

gem-Difluoroalkenes have found widespread applications in synthetic chemistry, pharmaceutical development, and materials science. These versatile fluorinated motifs serve as precursors for various organofluorine derivatives, including monofluoromethylenes, difluoromethylenes, and gem-difluorocyclopropanes. Notably, their incorporation into biomolecules as carbonyl bioisosteres has been shown to enhance pharmaceutical efficacy, metabolic stability, and target specificity, offering new avenues for drug discovery (Scheme 3-1). Conventional methods for synthesizing gem-difluoroalkenes often involve harsh reaction conditions, highly reactive intermediates, or well-defined organometallic reagents. These approaches frequently suffer from limited substrate scope and poor functional group tolerance due to their strong basic or nucleophilic nature.

Scheme 3-1 Representative biologically active gem-difluoroalkenes

In recent years, mild radical defluorinative strategies for constructing *gem*-difluoroalkenes have gained increasing interest, leveraging transition-metal catalysis, $^{265-270}$ photocatalysis, $^{271-279}$ and electrocatalysis. These methodologies typically involve radical generation through the cleavage of C—halogen, C—B, C—Si, C—C, C—O, or C—S bonds in various substrates (**Scheme 3-2**, **path a**). Compared to these radical precursors-utilized approaches, radical C(sp³)-H functionalization $^{153,283-286}$ offers a practical and efficient route to access structurally diverse *gem*-difluoroalkenes (**Scheme 3-2**, **path b**). In this field, several pioneering studies have achieved site-selective defluorinative *gem*-difluoroallylation of C(sp³)-H bonds using photocatalysis or synergistic light/metal catalysis. $^{287-293}$ For instance, in 2020, the Zhou group employed a photoredox/organocatalytic system to achieve β -C-H *gem*-difluoroallylation of aliphatic aldehydes and cyclic ketones. 287 In the following year, the Martin group successfully introduced *gem*-difluoroalkenes at saturated hydrocarbon sites via selective defluorinative C(sp³)-H alkylation of amides. 289 More recently, the Guo group reported a metal-free, visible-light-induced *gem*-difluoroallylation of glycine derivatives. 292 However, these methods primarily target α -heteroatom

 $C(sp^3)$ -H sites²⁸⁹⁻²⁹³ or remote $C(sp^3)$ -H positions.²⁸⁷⁻²⁸⁹

Despite these advances, the direct incorporation of *gem*-difluoroalkene units at α -carbonyl C(sp³)-H bonds remains an unresolved challenge. Herein, developing novel approaches that enable direct access to carbon-centered radicals at α -carbonyl positions remains a crucial goal for expanding the scope of functionalized α -carbonyl derivatives.

path a well-developed
$$R^1$$
— R^1 — R^1 — R^1 — R^2 — R^1 — R^2 — R^1 — R^1 — R^2 — R^2 — R^1 — R^2 —

Scheme 3-2 Radical defluorinative routes to *gem*-difluoroalkenes

2.3.2 Condition optimizations

Inspired by recent progress in 1.5-HAT, 153-177 we selected iodoamide 3-1a and trifluoromethyl alkene 3-2a as model substrates. Encouragingly, when the reaction was conducted under N2 using a catalytic amount of Ir(ppy)₃ and 1,3,5-trimethyl-1,3,5-triazinane as an additive in MeCN, irradiation with 456 nm blue LEDs for 12 hours yielded the desired gem-difluoroalkene 3-3aa in 58% isolated yield (entry 1). Substituting Ir(ppy)₃ with alternative photocatalysts, such as Ru(bpy)₃Cl₂·6H₂O, Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆, or 4CzIPN, led to a significant reduction in efficiency (entries 2-4). Various amines, including TMEDA, DIPEA, and Et₃N, facilitated the reaction but proved less effective than triazinane (entries 5-7). Interestingly, DABCO was found inferior, (entry 8) presumably because it is less prone to α -amino radical formation which is crucial in XAT. Notably, replacing triazinane with either K₂HPO₄ or pyridine completely suppressed product formation (entries 9-10), suggesting that the amines function as electron transfer mediators in the photocatalytic gem-difluoroallylation of amides. Additionally, the reaction proceeded in other polar solvents such as acetone, DCE, and DMSO, though with reduced yields (entries 11-13). Control experiments showed that the presence of light, the photocatalyst, and triazinane were all indispensable for the successful α -C(sp³)-H gem-difluoroallylation of 3-1a with trifluoromethyl alkene 3-2a (entry 14). Finally, adjusting the wavelength to 370 nm provided 3-3aa in lower yield (entry 15). The reaction was not improved by replacing the iridium photocatalyst with 10-phenylphenothiazine (PHT, entry 16).

Table 3-1 Condition optimizations^a

| entry | variations from standard conditions | 3-3aa , yield (%) ^b |
|-------|---|---------------------------------------|
| 1 | none | 65 (58) ^c |
| 2 | Ru(bpy) ₃ Cl ₂ ·6H ₂ O instead of Ir(ppy) ₃ | 8 |

| 3 | Ir[dF(CF ₃)ppy] ₂ (dtbbpy)PF ₆ instead of Ir(ppy) ₃ | 18 |
|-------------------|--|---|
| 4 | 4CzIPN instead of Ir(ppy) ₃ | 21 |
| 5 | TMEDA instead of triazinane | 34 |
| 6 | DIPEA instead of triazinane | 51 |
| 7 | Et ₃ N instead of triazinane | 52 |
| 8 | DABCO instead of triazinane | 17 |
| 9 | K ₂ HPO ₄ instead of triazinane | 0 |
| 10 | Pyridine instead of triazinane | 0 |
| 11 | Acetone instead of MeCN | 48 |
| 12 | DCE instead of MeCN | 53 |
| 13 | DMSO instead of MeCN | 42 |
| 14 | no light, or photocatalyst, or no triazinane | 0 |
| 15 | $\lambda = 370 \text{ nm } (40 \text{ W}) \text{ instead of } 456 \text{ nm}$ | 52 |
| 16 | $\lambda = 370 \text{ nm } (40 \text{ W}) \text{ with PHT as PC}$ | trace |
| STT 1 41 ' 4 1 41 | 4 1 1 4' 1'4' 0.15 | 1 |

^aUnless otherwise noted, the standard reaction conditions were as follows: **3-1a** (0.15 mmol, 1 equiv.), **3-2a** (0.3 mmol, 2 equiv.), photocatalyst (0.003 mmol, 2 mol %), and base (0.3 mmol, 2 equiv.) in solvent (1.5 mL) with irradiation by 40W LEDs (λ = 456 nm) under N₂ at room temperature for 12 h. ^bThe yields were determined by ¹⁹F NMR analysis of the crude reaction mixture using benzotrifluoride as an internal standard. ^cisolated yield.

2.3.3 Substrate scope studies

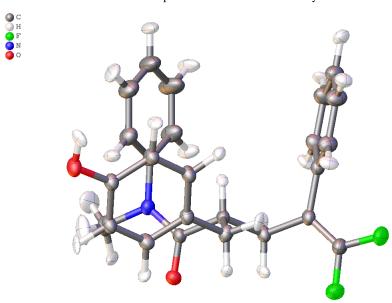
With the optimized conditions established, the substrate scope of this α -C(sp³)-H gemdifluoroallylation reaction was explored (Scheme 3-3). Notably, altering the N-protecting group from methyl to benzyl had little impact on efficiency, yielding 3-3ba in 57%. Several commonly used amides (3-1c-3-1f) were successfully transformed into their corresponding gemdifluoroallylated products (3-3ca-3-3fa) in 54%-67% yields. Delightedly, amides featuring diverse functional groups at the carbonyl fragment-including nitrile, alkene, phenol, and alkyne-were well tolerated under the optimized conditions, affording products 3-3ga-3-3ja in moderate to good yields (41%–68%). The structure of **3-3ia** was definitively confirmed by X-ray single-crystal analysis (Scheme 3-4). Furthermore, iodoarene substrates bearing electron-donating groups, such as methyl or methoxy (3-1k-3-11), provided the target compounds in 61% and 40% yield, respectively. Likewise, amides containing para-substituted fluoro, chloro, or trifluoromethyl groups delivered the desired products in yields of 32%-40% (3-3ma-3-3oa). Importantly, a primary α -carbon iodoamide substrate (3-1p) could also be converted to the expected gem-difluoroalkene 3-3pa, however in degraded yield (22%). Unfortunately, a benzylic α -carbon iodoamide substrate 3-1q led only to traces amount of the corresponding coupling product 3-3qa. These results illustrate some of the limits of the method. We also investigated whether the method would tolerate some heterocycles. Fortunately, incorporate both a pyridine unit and an indole core was achieved with encouraging yields (3-3ra-3-3sa, 39-41%).

Next the scope of trifluoromethyl alkenes in the α -C(sp³)-H gem-difluoroallylation of amide **3-1f** was examined. *meta*-Substituted trifluoromethyl styrenes with chloro, methyl, methoxy, or methylthio functionalities yielded the corresponding products in 52%-64% (**3-3fb-3-3fe**). Similarly, Trifluoromethyl styrenes bearing halogens (fluoro, chloro, bromo) or electron-rich groups (methyl, methoxy, *tert*-butyl) exhibited good reactivity, producing the target compounds in satisfactory yields (**3-3fg-3-3fl**, 49%-71%). However, *ortho*-fluoro substitution led to a significant drop in efficiency, affording only 30% yield (**3-3ff**).

Ir(ppy)₃

^aReaction conditions: **3-1** (0.15 mmol, 1 equiv.), **3-2** (0.3 mmol, 2 equiv.), $Ir(ppy)_3$ (0.003 mmol, 2 mol%), and triazinane (0.3 mmol, 2 equiv.) in anhydrous MeCN (1.5 mL) with irradiation by 40W LEDs (λ = 456 nm) under N₂ at room temperature for 12 h. ^b0.45 mmol **3-2** (3 equiv.) and 2.0 mL MeCN was used. ^c1 mmol scale: anhydrous MeCN (10 mL), react for 24 h. ^csee scheme **3-4** for detail.

Scheme 3-3 Scope of amides and trifluoromethyl alkenes^a



Scheme 3-4 X-ray image of **3-3ia**. Suitable single crystal for X-ray diffraction were grown by liquid-liquid diffusion from a concentrated solution of compound **3-3ia** in CH₂Cl₂ and *n*-hexane. And B.E. and Prof. Oppel measured and analyzed crystal structure. CCDC: 2411490

Moreover, the *gem*-difluoroallylation strategy was successfully applied to the late-stage modification of complex bioactive molecules (**Scheme 3-5**). Amides derived from gemfibrozil, dihydroquinolinone, and lithocholic acid were smoothly converted into their respective *gem*-difluoroalkene products in 60%, 45%, and 51% yield (**3-3ta**, **3-3ua**, **3-3va**). Remarkably, amide **3-1v**, synthesized from oleanolic acid, underwent selective α-carbonyl functionalization, affording **3-3va** in good yield while preserving its free hydroxyl group. Additionally, an estrone amide derivative **3-1w** was efficiently transformed under the mild conditions, providing **3-3wa** in 44% yield. Importantly, no *gem*-difluoroallylation occurred at the ketone moiety of **3-1w**, the lactam core of **3-1u**, or the ester functionality of **3-1x**, highlighting the high 1,5-regiospecificity of this transformation.

^aReaction conditions: **3-1** (0.15 mmol, 1 equiv.), **3-2** (0.3 mmol, 2 equiv.), $Ir(ppy)_3$ (0.003 mmol, 2 mol%), and triazinane (0.3 mmol, 2 equiv.) in anhydrous MeCN (1.5 mL) with irradiation by 40W LEDs (λ = 456 nm) under N_2 at room temperature for 12 h. ^bdr ratios were determined by ¹⁹F NMR analysis of purified product.

Scheme 3-5 Late-stage gem-difluoroallylation of complex molecules^a

2.3.4 Mechanistic studies

Next, a tertiary α -carbon iodoamide substrate **3-1y** was considered (**Scheme 3-6a**). However, only the known intramolecular spiro cyclization product **3-4y** could be found, and in good yield (83%).²⁹⁴ The *gem*-difluoroallylation coupling product **3-3ya** could not be detected. In order to gain more insight into the reaction mechanism, we first performed a radical trapping experiment. The addition of 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) as a radical scavenger significantly suppressed the reaction, and the TEMPO-adduct **3-5a** was isolated in 38% yield (**Scheme 3-6b**). This finding suggests that the α -C(sp³)-H *gem*-difluoroallylation of amides proceeds via a radical pathway. Next, fluorescence quenching experiments were conducted to assess interactions between the photocatalyst Ir(ppy)₃ and key reaction components, including the triazinane, iodoarene **3-1f**, and trifluoromethyl styrene **3-2a** (see Part **4.2.3** for details). Stern-Volmer analysis revealed pronounced quenching by the triazinane and minor quenching by iodoarene **3-1f**, supporting the hypothesis that the reaction is initiated by a photoredox cycle in which the triazinane participates in a reductive quenching process.

Based on these mechanistic studies, including the Stern-Volmer analysis and TEMPO inhibition experiment, and drawing from previous reports identifying the triazinane as a halogen atom transfer (XAT) agent, ²⁹⁵⁻²⁹⁶ a plausible reaction mechanism involving XAT and 1,5-hydrogen atom transfer (1,5-HAT) was proposed (**Scheme 3-6c**). Upon visible-light irradiation, the photocatalyst Ir(ppy)₃

undergoes photoexcitation and engages in a single-electron transfer (SET) event, oxidizing the triazinane to its radical cation form. Deprotonation then generates a diamino-substituted radical 3-I, which facilitates halogen atom transfer (XAT) to activate substrate 3-1, yielding the aryl radical intermediate 3-III. Subsequently, an intramolecular 1,5-HAT process converts intermediate 3-III into α -carbonyl radical 3-IV, which is then intercepted by trifluoromethyl styrene 3-2, forming intermediate 3-V. Finally, single-electron reduction of intermediate 3-V promotes C-F bond cleavage, leading to the formation of the target *gem*-difluoroalkene 3-3 while simultaneously regenerating the photocatalyst to complete the catalytic cycle.

a) Tertiary α -carbon substrate

Scheme 3-6 Mechanistic study and proposed catalytic cycle

2.3.5 Conclusion

In summary, a visible-light-driven aryl-to-alkyl radical relay strategy for the *gem*-difluoroallylation of amides using trifluoromethyl alkenes was developed. This approach leverages a mild and selective iodoarene-mediated photoredox process, integrating halogen atom transfer (XAT) and hydrogen atom transfer (HAT) to enable α -C(sp³)-H functionalization. The method exhibits broad functional group tolerance and efficiently delivers a variety of carbonyl-containing *gem*-difluoroalkenes. Furthermore, its applicability to the late-stage modification of bioactive molecules underscores its potential significance in drug discovery and development.

3 Summary and outlook

Oxindoles, quinoxalinones, and *gem*-difluoroalkenes are important motifs ubiquitous in nature products, drugs and bioactive molecules. Here we employed visible-light-mediated photochemical protocol to access these valuable compounds under mild conditions and with good efficiency.

First, a practical NaI/PPh₃-catalyzed decarboxylative radical cascade cyclization of N-arylacrylamides with redox-active esters was developed, utilizing visible light irradiation as a key driving force. This transition-metal-free strategy offers a mild, cost-effective, and highly efficient approach for constructing diverse oxindoles with a C3 quaternary stereogenic center. A broad scope of substrates, including α -amino acid-derived compounds and various carboxylic acid derivatives, were demonstrated under this optimal, simple and cheap catalytic system. Mechanistic investigations indicate that the reaction proceeds via a radical pathway (**Scheme 4-1**).

✓ transition-metal-free catalysis ✓ optimal, simple and cheap catalytic system ✓ broad scope

Scheme 4-1 Photocatalytic decarboxylative cyclization via NaI/PPh₃ catalysis

Next, a visible-light-driven photocatalytic strategy for the deoxygenative benzylation of quinoxalin-2(1*H*)-ones was reported. This innovative method offers a straightforward, mild, and practical approach to synthesizing 3-benzylquinoxalin-2(1*H*)-ones using readily available and non-hazardous carboxylic acid anhydrides. A diverse array of substrates bearing various functional groups exhibited excellent compatibility, leading to the efficient formation of functionalized 3-benzylquinoxalin-2(1*H*)-ones with significant potential for pharmaceutical applications. Mechanistic studies indicate that H₂O serves as the proton donor, while hydroxylated quinoxalin-2(1*H*)-ones may act as crucial intermediates in the deoxygenative photocatalytic transformation (Scheme 4-2).

Scheme 4-2 Photocatalyzed deoxygenative benzylation of quinoxalinones via SET reduction

Finally, we introduce a photocatalytic radical strategy for α -C(sp³)-H gem-difluoroallylation of amides with trifluoromethyl alkenes, enabling the synthesis of target compounds in good yields with broad functional group compatibility. The mild and efficient reaction conditions facilitate the concise incorporation of gem-difluoroalkene motifs as carbonyl bioisosteres into structurally complex molecules, including derivatives of gemfibrozil and estrone, highlighting their potential for late-stage modifications of pharmaceuticals, natural products, and biologically relevant

intermediates. Mechanistic insights reveal a radical-driven process involving XAT and 1,5-HAT (Scheme 4-3).

Scheme 4-3 Visible-light driven aryl-to-alkyl radical relay gem-difluoroallylation of amides

In the future, developing green and sustainable photochemistry for the synthesis of pharmaceutically relevant molecule scaffolds is the key in organic synthesis. Especially exploring non-metal photocatalyst like EDA complex and improving their reactivities are challenging and significant, and will open up new opportunities in radical generating. And researching these radical as versatile synthetic intermediates will play an important role in efficiently constructing structurally diverse functional molecules.

4 Experiment and data

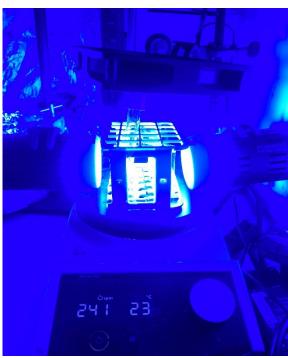
4.1 General information

Reagents

Unless otherwise noted, all commercially available compounds were used as provided without further purification and chemicals used for synthesis were purchased from Sigma Aldrich, Abcr, TCI, Fisher, Chempur and BLDPharm. Anhydrous solvents were obtained by using an Innovative Technology PS-MD-5 solvent purification system.

Photoreactor

All reactions with LEDs irradiation were carried out using a photo-reactor equipped with LED PhotoReaction Lighting (Model: PR160L, λ = 456 nm or 370 nm, max 40W), which is bought from Kessil company. The glass vials were placed at approximatively 4 cm away from the light source. In order to avoid overheating of the reaction system, the vials were cooled with a fan on top of the vials.





NMR spectroscopy

NMR spectra were recorded on an Agilent VNMRS 400 or a Bruker Av 600 using CDCl₃ or DMSO- d_6 as solvents. Chemical shifts δ are given in ppm relative to TMS as reference. The following abbreviations were used for 1 H, 13 C and 19 F NMR spectra to indicate the signal multiplicity: s (singlet), d (doublet), t (triplet), q (quartet) and m (multiplet) as well as combinations of them.

IR spectra

IR spectra were measured on a PerkinElmer 100 FT-IR spectrometer with an UATR Diamond KRS-5 unit.

Mass spectra

High-resolution mass spectra (HRMS) were obtained on a Thermo Scientific LTQ Orbitrap XL spectrometer.

X-ray measurement

Crystallographic data were collected on a Bruker Kappa APEX II CCD-diffractometer with monochromatic Mo– $K\alpha$ radiation (λ =0.71073 Å) and a CCD detector.

Stern-Volmer experiments

The Agilent Cary Eclipse Fluorescence Spectrometer is used to conduct Stern-Volmer fluorescence quenching analysis. We thank Steffen Schauerte and Prof. Dr. Markus Albrecht (Institute of Organic Chemistry, RWTH Aachen University) for access to the equipment.

Others

Aluminium blocks equipped with slots that accommodate the glass vial reactors were utilized for all experiments requiring heating. Thin-layer chromatography (TLC) was performed on VWR silica gel aluminium plates with F-254 indicator, visualized by UV light irradiation.

4.2 Experimental methods and data

4.2.1 NaI/PPh₃-catalyzed visible-light-mediated decarboxylative radical cascade cyclization of

N-arylacrylamides for the efficient synthesis of quaternary oxindoles

General procedure for the synthesis of quaternary oxindoles

To an oven-dried vial equipped with a stirring bar, acrylamide 1-1 (0.3 mmol), redox-active ester 1-2 (0.2 mmol), NaI (6 mg, 0.04 mmol) and Ph_3P (10.5 mg, 0.04 mmol) were added, then the tube was evacuated and filled with N_2 (three times) before anhydrous acetonitrile (MeCN, 2.0 mL) was added. The reaction was performed under blue LEDs irradiation (456 nm, 40 W) at room temperature. After 36 h, the solvent was removed in vacuo, and the residue was purified by column chromatography to give the corresponding quaternary oxindole products.

Product characterization

3-(Cyclohexylmethyl)-1,3-dimethylindolin-2-one (1-3aa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 72% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 (t, J = 7.8 Hz, 1H), 7.08 (d, J = 7.2 Hz, 1H), 6.98 (t, J = 7.2 Hz, 1H), 6.76 (d, J = 7.8 Hz, 1H), 3.14 (s, 3H), 1.85 (dd, J = 14.4, 6.6 Hz, 1H), 1.65 (dd, J = 14.4, 5.4 Hz, 1H), 1.47 – 1.34 (m, 3H), 1.27 (d, J = 11.4 Hz, 1H), 1.24 (s, 3H), 1.14 (d, J = 12.0 Hz, 1H), 0.98 – 0.82 (m, 4H), 0.78 – 0.64 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.1, 143.1, 134.4, 127.5, 122.7, 122.3, 107.9, 47.8, 45.4, 34.7, 34.5, 33.5, 26.2, 26.15, 26.10, 26.0.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-1,3,5-trimethylindolin-2-one (1-3ba)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 68% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.98 (d, J = 7.8 Hz, 1H), 6.89 (s, 1H), 6.65 (d, J = 7.8 Hz, 1H), 3.12 (s, 3H), 2.28 (s, 3H), 1.84 (dd, J = 14.4, 7.2 Hz, 1H), 1.63 (dd, J = 14.4, 5.4 Hz, 1H), 1.48 – 1.36 (m, 3H), 1.30 – 1.12 (m, 5H), 0.94 – 0.86 (m, 4H), 0.78 – 0.64 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.1, 140.8, 134.5, 131.8, 127.7, 123.6, 107.6, 47.9, 45.4, 34.7, 34.5, 33.5, 26.3, 26.2, 26.1, 26.0, 21.2.

Data is consistent with the literature. 197

1-(Cyclohexylmethyl)-6-methoxy-1,3-dimethyl-1,3-dihydro-2*H*-inden-2-one (1-3ca)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 38 mg product was obtained by 66% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.73 – 6.69 (m, 2H), 6.68 – 6.65 (m, 1H), 3.74 (s, 3H), 3.12 (s, 3H), 1.85 (dd, J = 14.4, 7.2 Hz, 1H), 1.62 (dd, J = 14.4, 5.4 Hz, 1H), 1.47 – 1.36 (m, 3H), 1.38 – 1.12 (m, 5H), 0.94 – 0.82 (m, 4H), 0.82 – 0.63 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.8, 155.9, 136.7, 135.9, 111.4, 110.5, 108.1, 55.8, 48.3, 45.4, 34.7, 34.4, 33.5, 26.6, 26.3, 26.1, 26.0.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-5-fluoro-1,3-dimethylindolin-2-one (1-3da)

The crude mixture was purified by silica gel column chromatography with pentane/EA (20:1). 41 mg product was obtained by 74% isolated yield as colorless liquid.

 1 H NMR (400 MHz, Chloroform-*d*) δ 7.01 – 6.85 (m, 2H), 6.74 (dd, J = 8.4, 4.0 Hz, 1H), 3.19 (s, 3H), 1.92 (dd, J = 14.2, 6.8 Hz, 1H), 1.68 (dd, J = 14.2, 5.2 Hz, 1H), 1.55 – 1.42 (m, 3H), 1.34 – 1.18 (m, 5H), 1.02 – 0.88 (m, 4H), 0.85 – 0.71 (m, 2H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 180.7, 159.3 (d, J = 242.4 Hz), 139.0 (d, J = 1.6 Hz), 136.2 (d, J = 8.1 Hz), 113.6 (d, J = 23.2 Hz), 111.8 (d, J = 25.2 Hz), 108.3 (d, J = 8.1 Hz), 48.3 (d, J = 1.6 Hz), 45.3, 34.7, 34.4, 33.4, 26.3, 26.1, 26.04, 26.01, 25.98.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ -121.02 (td, J = 8.7, 3.9 Hz).

Data is consistent with the literature.²⁰⁹

5-Chloro-3-(cyclohexylmethyl)-1,3-dimethylindolin-2-one (1-3ea)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 38 mg product was obtained by 65% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-*d*) δ 7.16 (dd, J = 8.4, 1.2 Hz, 1H), 7.06 (d, J = 2.4 Hz, 1H), 6.69 (d, J = 8.4 Hz, 1H), 3.13 (s, 3H), 1.86 (dd, J = 14.4, 7.2 Hz, 1H), 1.63 (dd, J = 14.4, 5.4 Hz, 1H), 1.49 – 1.34 (m, 3H), 1.26 – 1.12 (m, 5H), 0.98 – 0.83 (m, 4H), 0.79 – 0.63 (m, 2H).

¹³C NMR (151 MHz, Chloroform-d) δ 180.6, 141.7, 136.2, 127.8, 127.5, 123.2, 108.9, 48.1, 45.3,

34.7, 34.4, 33.4, 26.3, 26.2, 26.1, 26.03, 25.98.

Data is consistent with the literature.²⁰⁹

3-(Cyclohexylmethyl)-1,3-dimethyl-5-(trifluoromethyl)indolin-2-one (1-3fa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 55 mg product was obtained by 85% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.48 (d, J = 7.8 Hz, 1H), 7.31 (s, 1H), 6.84 (d, J = 7.8 Hz, 1H), 3.18 (s, 3H), 1.88 (dd, J = 14.4, 7.2 Hz, 1H), 1.68 (dd, J = 14.4, 5.4 Hz, 1H), 1.48 – 1.36 (m, 3H), 1.28 – 1.09 (m, 5H), 0.96 – 0.82 (m, 4H), 0.80 – 0.64 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.0, 146.1, 135.0, 125.4 (q, J = 4.7 Hz), 124.6 (q, J = 33.5 Hz), 119.7 (q, J = 4.4 Hz, 1H), 107.7, 47.9, 45.3, 34.7, 34.4, 33.5, 26.4, 26.0, 25.98, 25.94.

 19 F NMR (282 MHz, Chloroform-*d*) δ -61.32.

Data is consistent with the literature. 197

Ethyl 3-(cyclohexylmethyl)-1,3-dimethyl-2-oxoindoline-5-carboxylate (1-3ga)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 48 mg product was obtained by 73% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.96 (d, J = 8.4 Hz, 1H), 7.76 (s, 1H), 6.80 (d, J = 8.4 Hz, 1H), 4.31 (q, J = 7.2 Hz, 2H), 3.18 (s, 3H), 1.87 (dd, J = 14.4, 7.2 Hz, 1H), 1.75 – 1.68 (dd, J = 14.4, 7.2 Hz, 1H), 1.48 – 1.37 (m, 3H), 1.34 (t, J = 7.2 Hz, 3H), 1.29 – 1.11 (m, 5H), 0.97 – 0.79 (m, 4H), 0.79 – 0.63 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.4, 166.6, 147.2, 134.3, 130.3, 124.7, 123.9, 107.4, 60.9, 47.7, 45.3, 34.7, 34.4, 33.4, 26.4, 26.1, 26.0, 25.98, 14.4.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-1,3,7-trimethylindolin-2-one (1-3ha)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 31 mg product was obtained by 57% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.99 – 6.64 (m, 3H), 3.42 (s, 3H), 2.52 (s, 3H), 1.84 (dd, J = 14.4, 7.2 Hz, 1H), 1.61 (dd, J = 14.4, 5.4 Hz, 1H), 1.48 – 1.37 (m, 3H), 1.29 (d, J = 13.2 Hz, 1H), 1.21 (s, 3H), 1.15 (d, J = 13.2 Hz, 1H), 0.96 – 0.83 (m, 4H), 0.79 – 0.62 (m, 2H).

¹³C NMR (151 MHz, Chloroform-d) δ 181.9, 140.9, 135.1, 131.2, 122.2, 120.7, 119.5, 47.2, 45.7,

34.6, 34.5, 33.5, 29.5, 26.6, 26.1, 26.1, 26.0, 19.1.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-1,3-dimethyl-7-phenylindolin-2-one (1-3ia)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 41 mg product was obtained by 62% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.41 – 7.20 (m, 5H), 7.07 (d, J = 7.2 Hz, 1H), 7.04 – 6.91 (m, 2H), 2.65 (s, 3H), 1.88 (dd, J = 13.8, 7.2 Hz, 1H), 1.67 (dd, J = 13.8, 5.4 Hz, 1H), 1.56 – 1.37 (m, 3H), 1.33 (d, J = 12.0 Hz, 1H), 1.29 (s, 3H), 1.17 (d, J = 14.2 Hz, 1H), 0.98 – 0.89 (m, 4H), 0.80 – 0.67 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 182.2, 140.1, 139.2, 135.5, 130.5, 130.0, 129.8, 127.9, 127.8, 127.6, 125.4, 121.7, 121.6, 47.2, 46.0, 34.8, 34.5, 33.6, 30.2, 26.3, 26.2, 26.14, 26.1.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-7-fluoro-1,3-dimethylindolin-2-one (1-3ja)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 28 mg product was obtained by 50% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.94 – 6.82 (m, 3H), 3.36 (s, 3H), 1.86 (dd, J = 14.4, 7.2 Hz, 1H), 1.64 (dd, J = 14.4, 5.4 Hz, 1H), 1.49 – 1.36 (m, 3H), 1.28 (d, J = 12.0 Hz, 1H), 1.24 (s, 3H), 1.14 (d, J = 12.0 Hz, 1H), 0.95 – 0.83 (m, 4H), 0.81 – 0.65 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 147.8 (d, J = 243.4 Hz), 137.5 (d, J = 3.0 Hz), 129.7 (d, J = 8.0 Hz), 122.9 (d, J = 6.3 Hz), 118.6 (d, J = 3.2 Hz) 115.6 (d, J = 19.3 Hz), 48.2 (d, J = 2.0 Hz), 45.6, 34.7, 34.4, 33.5, 28.7, 28.6, 26.4, 26.08, 26.06, 26.0.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -136.73.

Data is consistent with the literature. 197

7-Chloro-3-(cyclohexylmethyl)-1,3-dimethylindolin-2-one (1-3ka)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 63% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.10 (d, J = 8.4 Hz, 1H), 6.96 (d, J = 7.2 Hz, 1H), 6.89 (t, J = 7.8 Hz, 1H), 3.51 (s, 3H), 1.86 (dd, J = 14.4, 7.2 Hz, 1H), 1.62 (dd, J = 14.4, 5.4 Hz, 1H), 1.49 – 1.37 (m, 3H), 1.27 (d, J = 13.8 Hz, 1H), 1.23 (s, 3H), 1.14 (d, J = 12.6 Hz, 1H), 0.95 – 0.82 (m, 4H),

0.78 - 0.65 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.3, 139.0, 137.3, 129.9, 123.1, 121.2, 115.4, 47.7, 45.6, 34.6, 34.5, 33.5, 29.5, 26.6, 26.1, 26.0.

Data is consistent with the literature.²⁰⁹

1-(Cyclohexylmethyl)-1-methyl-5,6-dihydro-4*H*-pyrrolo[3,2,1-ij]quinolin-2(1*H*)-one (1-3la)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 38 mg product was obtained by 67% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.98 – 6.83 (m, 3H), 3.64 (t, J = 4.8 Hz, 2H), 2.76 – 2.68 (m, 2H), 1.96 – 1.86 (m, 2H), 1.83 (dd, J = 14.4, 7.2 Hz, 1H), 1.64 (dd, J = 14.4, 4.2 Hz, 1H), 1.47 – 1.37 (m, 3H), 1.32 (d, J = 13.2 Hz, 1H), 1.18 (d, J = 14.4 Hz, 1H), 1.25 (s, 3H), 0.96 – 0.88 (m, 4H), 0.80 – 0.67 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.0, 138.9, 133.0, 126.3, 121.8, 120.6, 120.0, 49.2, 45.3, 38.8, 34.8, 34.5, 33.6, 26.15, 26.13, 26.0, 25.8, 24.7, 21.4.

HRMS (ESI-MS) Calcd. For C₁₉H₂₅NONa [M+Na]⁺ 306.18284, found: 306.18234.

IR (neat, cm⁻¹): v: 3413, 2921, 1706, 1626, 1482, 1347, 1238, 1164, 1065, 956, 871, 749.

3-(Cyclohexylmethyl)-1-ethyl-3-methylinolin-2-one (1-3ma)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 36 mg product was obtained by 66% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.17 (td, J = 7.2, 1.2 Hz, 1H), 7.09 (dd, J = 7.8, 1.2 Hz, 1H), 6.97 (td, J = 7.2, 1.2 Hz, 1H), 6.78 (d, J = 7.8 Hz, 1H), 3.81 (dq, J = 14.4, 7.3 Hz, 1H), 3.83-3.78 (dq, J = 14.4, 7.2 Hz, 1H), 1.85 (dd, J = 14.4, 6.6 Hz, 1H), 1.64 (dd, J = 14.4, 6.6 Hz, 1H), 1.47 – 1.31 (m, 4H), 1.23 (s, 3H), 1.17 (t, J = 7.2 Hz, 3H), 1.12 – 1.07 (m, 1H), 0.95 – 0.82 (m, 4H), 0.79 – 0.61 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.6, 142.2, 134.8, 127.4, 122.9, 122.1, 108.1, 47.7, 45.5, 34.8, 34.4, 34.38, 33.7, 26.1, 26.08, 26.0, 12.5.

Data is consistent with the literature. 197

1-Benzyl-3-(cyclohexylmethyl)-3-methylindolin-2-one (1-3na)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 41 mg product was obtained by 61% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.23 – 7.13 (m, 5H), 7.10 – 7.04 (m, 2H), 6.93 (t, J = 7.2 Hz, 1H), 6.66 (d, J = 7.8 Hz, 1H), 4.97 (d, J = 15.6 Hz, 1H), 4.72 (d, J = 15.6 Hz, 1H), 1.91 (dd, J = 13.8, 6.6 Hz, 1H), 1.68 (dd, J = 13.8, 6.0 Hz, 1H), 1.45 – 1.34 (m, 4H), 1.29 (s, 3H), 1.07 (d, J = 12.6 Hz, 1H), 0.96 – 0.74 (m, 5H), 0.67 – 0.59 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.0, 142.3, 136.2, 134.6, 128.7, 127.5, 127.4, 122.8, 122.3, 109.0, 47.9, 45.5, 43.7, 34.8, 34.4, 34.0, 26.6, 26.12, 26.10, 26.06.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-3-methyl-1-phenylindolin-2-one (1-30a)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 38 mg product was obtained by 60% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.49 - 7.41 (m, 2H), 7.35 - 7.28 (m, 3H), 7.16 - 7.09 (m, 2H), 7.02 (t, J = 7.2 Hz, 1H), 6.76 (d, J = 7.8 Hz, 1H), 1.96 (dd, J = 13.8, 7.2 Hz, 1H), 1.73 (dd, J = 13.8, 5.4 Hz, 1H), 1.50 - 1.39 (m, 4H), 1.37 (s, 3H), 1.23 - 1.19 (m, 1H), 1.05 - 0.89 (m, 4H), 0.84 - 0.71 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.5, 143.0, 134.8, 134.2, 129.6, 127.8, 127.4, 126.5, 123.0, 122.8, 109.3, 47.9, 45.9, 35.0, 34.4, 33.6, 26.5, 26.2, 26.1, 26.08.

Data is consistent with the literature. 197

3-(Cyclohexylmethyl)-1-methyl-3-phenylindolin-2-one (1-3pa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 22 mg product was obtained by 34% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.32 – 7.24 (m, 3H), 7.21 – 7.15 (m, 4H), 7.15 – 7.11 (m, 1H), 7.05 (t, J = 7.8 Hz, 1H), 6.83 (d, J = 7.8 Hz, 1H), 3.13 (s, 3H), 2.33 (dd, J = 13.8, 6.6 Hz, 1H), 2.07 (dd, J = 13.8, 5.4 Hz, 1H), 1.49 – 1.31 (m, 4H), 0.98 – 0.73 (m, 7H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 179.0, 143.9, 141.6, 132.1, 128.4, 128.1, 127.1, 126.7, 125.3, 122.4, 108.3, 56.1, 45.3, 34.9, 34.6, 33.6, 29.7, 26.5, 26.2, 26.1

Data is consistent with the literature. 197

Methyl 2-(3-(cyclohexylmethyl)-5-methoxy-1-methyl-2-oxoindolin-3-yl)acetate (1-3qa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 48 mg product was obtained by 70% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 6.75 – 6.63 (m, 3H), 3.72 (s, 3H), 3.36 (s, 3H), 3.15 (s, 3H), 2.87 (d, J = 16.2 Hz, 1H), 2.70 (d, J = 16.2 Hz, 1H), 1.76 (dd, J = 13.8, 6.0 Hz, 1H), 1.63 (dd, J = 13.8, 5.4 Hz, 1H), 1.48 – 1.33 (m, 4H), 1.09 (d, J = 11.4 Hz, 1H), 0.96 – 0.85 (m, 4H), 0.82 – 0.74 (m, 1H), 0.69 – 0.63 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 179.2, 170.1, 155.7, 138.0, 132.8, 112.0, 110.7, 108.1, 55.8, 51.5, 49.7, 45.2, 42.6, 34.6, 34.0, 33.9, 26.4, 26.1, 26.04, 26.02.

Data is consistent with the literature. ²⁰⁹

4-(Cyclohexylmethyl)-2,4-dimethylisoquinoline-1,3(2H,4H)-dione (1-3ra)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 38 mg product was obtained by 66% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 8.19 (d, J = 7.8 Hz, 1H), 7.56 (t, J = 7.8 Hz, 1H), 7.40 – 7.29 (m, 2H), 3.32 (s, 3H), 2.26 (dd, J = 14.4, 7.8 Hz, 1H), 1.83 (dd, J = 14.4, 4.8 Hz, 1H), 1.49 (s, 3H), 1.46 – 1.35 (m, 3H), 1.18 (d, J = 8.4 Hz, 1H), 1.08 (d, J = 12.6 Hz, 1H), 0.94 – 0.63 (m, 6H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 176.9, 164.5, 143.9, 133.7, 128.9, 127.2, 125.7, 124.6, 49.6,

46.7, 34.9, 34.3, 33.0, 31.7, 27.2, 26.03, 26.0, 26.96.

Data is consistent with the literature.²⁰⁹

3-(Cyclobutylmethyl)-1,3-dimethylindolin-2-one (1-3ab)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 29 mg product was obtained by 63% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 (t, J = 7.8 Hz, 1H), 7.09 (d, J = 7.8 Hz, 1H), 6.97 (t, J = 7.8 Hz, 1H), 6.75 (d, J = 7.8 Hz, 1H), 3.12 (s, 3H), 1.97 (dd, J = 12.6, 6.6 Hz, 1H), 1.87 – 1.76 (m, 2H), 1.66 – 1.59 (m, 1H), 1.57 – 1.47 (m, 3H), 1.43 – 1.37 (m, 1H), 1.36 – 1.28 (m, 1H), 1.25 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.8, 143.3, 134.2, 127.6, 122.8, 122.2, 107.7, 47.9, 45.6, 32.9, 29.5, 28.9, 26.1, 23.9, 18.9.

Data is consistent with the literature.²⁰⁸

3-(Cyclopentylmethyl)-1,3-dimethylindolin-2-one (1-3ac)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 36 mg product was obtained by 73% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 (t, J = 7.8 Hz, 1H), 7.09 (d, J = 6.0 Hz, 1H), 6.98 (t, J = 7.8 Hz, 1H), 6.76 (d, J = 7.8 Hz, 1H), 3.15 (s, 3H), 1.99 (dd, J = 13.8, 7.2 Hz, 1H), 1.82 (dd, J = 13.8, 6.0 Hz, 1H), 1.43 –1.31 (m, 3H), 1.27 (s, 3H), 1.25 – 1.11 (m, 4H), 0.97 – 0.90 (m, 1H), 0.80 – 0.70 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.2, 143.3, 134.4, 127.6, 122.9, 122.3, 107.9, 48.5, 44.5, 37.2, 33.8, 32.8, 26.2, 25.3, 24.9, 24.86.

Data is consistent with the literature. ²⁰⁸

3-((2,3-Dihydro-1*H*-inden-2-yl)methyl)-1,3-dimethylindolin-2-one (1-3ad)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 43 mg product was obtained by 74% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.20 (t, J = 7.8 Hz, 1H), 7.13 (d, J = 7.8 Hz, 1H), 7.03 – 6.88 (m, 5H), 6.78 (d, J = 7.8 Hz, 1H), 3.17 (s, 3H), 2.59 (dd, J = 15.6, 7.6 Hz, 1H), 2.47 (dd, J = 15.6, 9.6 Hz, 1H), 2.34 (dd, J = 15.6, 7.8 Hz, 1H), 2.29 – 2.19 (m, 2H), 2.03 (dd, J = 13.8, 6.0 Hz, 1H), 1.98 – 1.92 (m, 1H), 1.32 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.8, 143.3, 143.2, 143.1, 134.0, 127.9, 126.1, 126.0, 124.1, 124.0, 122.9, 122.5, 108.0, 48.5, 43.9, 40.2, 39.5, 37.8, 26.3, 25.1.

Data is consistent with the literature.²⁰⁸

3-(((1r,3s,5r,7r)-Adamantan-2-yl)methyl)-1,3-dimethylindolin-2-one (1-3ae)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 25 mg product was obtained by 40% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.17 (t, J = 7.8 Hz, 1H), 7.09 (d, J = 7.2 Hz, 1H), 6.96 (t, J = 7.8 Hz, 1H), 6.75 (d, J = 7.8 Hz, 1H), 3.13 (s, 3H), 2.05 (dd, J = 13.8, 6.0 Hz, 1H), 1.82 – 1.70 (m, 3H), 1.66 – 1.49 (m, 6H), 1.42 – 1.29 (m, 5H), 1.28 (s, 3H), 1.21 – 1.17 (m, 1H), 1.08 – 1.05 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.9, 143.3, 134.3, 127.6, 122.6, 122.3, 107.8, 48.4, 41.7, 41.0, 39.0, 39.0, 38.1, 33.2, 32.6, 31.8, 31.7, 27.7, 27.6, 26.1, 25.1.

Data is consistent with the literature.²⁰⁸

1,3-Dimethyl-3-((tetrahydrofuran-2-yl)methyl)indolin-2-one (1-3af)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 32 mg (dr = 1:1.3) product was obtained by 65% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) mixture of diastereomers δ 7.24 – 7.09 (m, 4H), 7.00 – 6.95 (m, 2H), 6.78 – 6.76 (m, 2H), 3.64 – 3.59 (m, 2H), 3.46 – 3.41 (m, 3H), 3.37 – 3.32 (m, 1H), 3.14 (s, 6H), 2.18 – 2.13 (m, 2H), 1.97 (dd, J = 13.8, 7.8 Hz, 1H), 1.78 – 1.49 (m, 7H), 1.36 – 1.21 (m, 8H). ¹³C NMR (151 MHz, Chloroform-*d*) mixture of diastereomers δ 180.9, 180.6, 143.7, 142.9, 133.7, 133.6, 127.8, 127.7, 123.1, 122.8, 122.5, 121.9, 108.0, 107.9, 76.1, 75.6, 67.1, 67.09, 47.0, 46.8, 43.7, 43.2, 31.7, 31.4, 26.3, 26.2, 25.8, 25.3, 25.0, 24.8.

HRMS (ESI-MS) Calcd. For C₁₅H₁₉NO₂Na [M+Na]⁺ 268.13080, found: 268.13044.

IR (neat, cm⁻¹): v: 3493, 2931, 1708, 1611, 1468, 1347, 1249, 1122, 1055, 930, 750, 699.

tert-Butyl 4-((1,3-dimethyl-2-oxoindolin-3-yl)methyl)piperidine-1-carboxylate (1-3ag)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 53 mg product was obtained by 74% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.21 (t, J = 6.8 Hz, 1H), 7.09 (d, J = 7.2 Hz, 1H), 7.00 (t, J = 7.2 Hz, 1H), 6.79 (d, J = 7.8 Hz, 1H), 3.79 (s, 2H), 3.16 (s, 3H), 2.46 – 2.27 (m, 2H), 1.91 (dd, J = 14.4, 6.0 Hz, 1H), 1.69 (dd, J = 14.4, 5.4 Hz, 1H), 1.33 (s, 9H), 1.26 (s, 3H), 1.08 – 0.91 (m, 3H), 0.90 – 0.70 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 154.7, 143.0, 134.0, 127.8, 122.7, 122.6, 108.1, 79.1, 47.7, 44.4, 33.2, 33.0, 32.5, 28.4, 26.3, 26.2.

Data is consistent with the literature.²⁰⁸

tert-Butyl 4-((1,3-dimethyl-2-oxoindolin-3-yl)methyl)-4-methylpiperidine-1-carboxylate (1-3ah)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 57 mg product was obtained by 76% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.22 – 7.18 (m, 1H), 7.13 (d, J = 7.2 Hz, 1H), 6.97 (t, J = 7.8 Hz, 1H), 6.79 (d, J = 7.8 Hz, 1H), 3.51 – 3.43 (m, 2H), 3.16 (s, 3H), 2.88 – 2.84 (m, 1H), 2.78 – 2.73 (m, 1H), 2.08 (d, J = 14.4 Hz, 1H), 1.83 (d, J = 14.4 Hz, 1H), 1.34 (s, 9H), 1.26 – 1.21 (m, 4H), 1.06 – 0.91 (m, 2H), 0.77 – 0.75 (m, 1H), 0.50 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.9, 154.8, 142.6, 134.0, 127.8, 123.7, 122.2, 108.2, 79.1, 50.1, 46.9, 38.2, 37.9, 32.8, 30.9, 28.5, 28.4, 26.3, 22.5.

HRMS (ESI-MS) Calcd. For C₂₂H₃₂N₂O₃Na [M+Na]⁺ 395.23051, found: 395.23091.

IR (neat, cm⁻¹): v: 3395, 2923, 1683, 1609, 1418, 1367, 1248, 1159, 1090, 1023, 961, 862, 760, 699.

3-Isobutyl-1,3-dimethylindolin-2-one (1-3ai)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 30 mg product was obtained by 69% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 (t, J = 7.8 Hz, 1H), 7.08 (d, J = 7.2 Hz, 1H), 6.98 (t, J = 7.2 Hz, 1H), 6.77 (d, J = 7.8 Hz, 1H), 3.14 (s, 3H), 1.86 (dd, J = 13.8, 7.8 Hz, 1H), 1.68 (dd, J = 13.8, 5.4 Hz, 1H), 1.25 (s, 3H), 1.22 – 1.12 (m, 1H), 0.58 (d, J = 6.6 Hz, 3H), 0.53 (d, J = 6.6 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 181.1, 143.2, 134.2, 127.6, 122.8, 122.3, 107.9, 48.1, 46.8, 26.2, 26.1, 25.5, 24.1, 22.8.

Data is consistent with the literature.²⁰⁸

3-(2-Ethylhexyl)-1,3-dimethylindolin-2-one (1-3aj)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 35 mg (dr = 1:1.1) product was obtained by 64% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) mixture of diastereomers δ 7.24 (t, J = 7.8 Hz, 2H), 7.15 (dd, J = 7.2, 3.0 Hz, 2H), 7.04 (t, J = 7.8 Hz, 2H), 6.82 (d, J = 7.8 Hz, 2H), 3.20 (d, J = 3.0 Hz, 6H), 1.95 – 1.89 (m, 2H), 1.78 – 1.72 (m, 2H), 1.33 (s, 6H), 1.22 – 0.85 (m, 18H), 0.78 (t, J = 7.2 Hz, 3H), 0.75 (t, J = 7.2 Hz, 3H), 0.69 (t, J = 7.2 Hz, 3H), 0.64 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) mixture of diastereomers δ 181.0, 180.98, 143.3, 143.27, 134.3, 134.25, 127.5, 122.9, 122.8, 122.22, 122.2, 107.82, 107.8, 48.09, 48.0, 42.02, 42.0, 35.72, 35.7, 33.2, 32.8, 28.3, 28.2, 26.5, 26.2, 26.1, 25.6, 25.5, 22.8, 22.7, 14.0, 14.99, 10.3, 10.29.

HRMS (ESI-MS) Calcd. For C₁₈H₂₇NONa [M+Na]⁺ 296.19849, found: 296.19804.

IR (neat, cm⁻¹): v: 3418, 2925, 1712, 1611, 1465, 1375, 1247, 1123, 1025, 929, 749, 699.

1,3-Dimethyl-3-neopentylindolin-2-one (1-3ak)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 36 mg product was obtained by 77% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 (d, J = 7.8 Hz, 1H), 7.13 (d, J = 7.2 Hz, 1H), 6.96 (t, J = 7.2 Hz, 1H), 6.78 (d, J = 7.8 Hz, 1H), 3.15 (s, 3H), 2.09 (d, J = 14.4 Hz, 1H), 1.79 (d, J = 14.4 Hz, 1H), 1.22 (s, 3H), 0.54 (s, 9H).

¹³C NMR (151 MHz, Chloroform-d) δ 181.0, 142.9, 134.2, 127.5, 123.9, 122.0, 108.0, 50.8, 47.4,

31.8, 30.8, 28.3, 26.2.

Data is consistent with the literature.²⁰⁸

tert-Butyl (1-(1,3-dimethyl-2-oxoindolin-3-yl)-4-(methylthio)butan-2- yl)carbamate (1-3al)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 53 mg (dr = 1:1.1) product was obtained by 70% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) mixture of diastereomers δ 7.12 – 7.17 (m, 3H), 7.09 (d, J = 7.2 Hz, 1H), 7.03 – 6.99 (m, 2H), 6.79 – 6.74 (m, 2H), 4.13 (d, J = 9.6 Hz, 1H), 3.92 (d, J = 9.0 Hz, 1H), 3.45 (dt, J = 10.2, 4.8 Hz, 1H), 3.20 (dt, J = 10.2, 4.8 Hz, 1H), 3.20 (s, 3H), 3.14 (s, 3H), 2.36 – 2.20 (m, 4H), 2.10 – 1.89 (m, 10H), 1.56 – 1.38 (m, 4H), 1.28 – 1.18 (m, 24H).

¹³C NMR (151 MHz, Chloroform-*d*) mixture of diastereomers δ 181.4, 180.1, 154.7, 154.5, 143.1, 142.9, 133.9, 132.5, 128.1, 127.7, 122.75, 122.70, 122.57, 122.53, 108.4, 108.3, 78.80, 78.76, 47.9, 47.5, 47.2, 46.9, 42.5, 42.0, 36.8, 36.2, 30.3, 30.2, 28.4, 28.3, 26.35, 26.27, 25.6, 25.1, 15.47, 15.46. HRMS (ESI-MS) Calcd. For $C_{20}H_{30}N_2O_3SNa$ [M+Na]⁺ 401.18693 found: 401.18666.

IR (neat, cm⁻¹): v: 3339, 2970, 1697, 1612, 1496, 1364, 1244, 1167, 1124, 1046, 856, 750, 699.

1,3-Dimethyl-3-(pent-4-en-1-yl)indolin-2-one (1-3am)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 29 mg product was obtained by 63% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.22 – 7.16 (m, 1H), 7.09 (d, J = 7.2 Hz, 1H), 6.99 (t, J = 7.8 Hz, 1H), 6.76 (d, J = 7.8 Hz, 1H), 5.58 (m, 1H), 4.87 – 4.75 (m, 2H), 3.13 (s, 3H), 1.92 – 1.77 (m, 3H), 1.65 (t, J = 13.2 Hz, 1H), 1.28 (s, 3H), 1.06 – 0.98 (m, 1H), 0.89 – 0.81 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 143.3, 138.2, 134.2, 127.7, 122.5, 122.46, 114.7, 107.9, 48.4, 38.0, 33.7, 26.1, 23.8.

HRMS (ESI-MS) Calcd. For C₁₅H₂₀NO [M+H]⁺230.15394, found: 230.15397.

IR (neat, cm⁻¹): \tilde{v} : 3416, 2927, 2324, 2087, 1912, 1710, 1611, 1467, 1346, 1248, 1124, 1019, 911, 749, 699.

3-(Hex-5-yn-1-yl)-1,3-dimethylindolin-2-one (1-3an)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 29 mg product was obtained by 60% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.25 – 7.16 (m, 1H), 7.10 (d, J = 7.2 Hz, 1H), 6.99 (t, J = 7.8 Hz, 1H), 6.77 (d, J = 7.8 Hz, 1H), 3.14 (s, 3H), 1.99 (t, J = 7.2 Hz, 2H), 1.87 – 1.76 (m, 2H), 1.67 (td, J = 12.6, 4.2 Hz, 1H), 1.40 – 1.25 (m, 5H), 1.06 – 0.98 (m, 1H), 0.96 – 0.85 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 143.3, 134.1, 127.7, 122.5, 122.5, 107.9, 84.2, 68.3, 48.3, 37.9, 28.6, 26.1, 23.8, 23.7, 18.1.

HRMS (ESI-MS) Calcd. For C₁₆H₁₉NONa [M+Na]⁺ 264.13589, found: 264.13565.

IR (neat, cm⁻¹): v: 4294, 2930, 1706, 1611, 1466, 1346, 1250, 1123, 750, 697.

3-(5-Chloropentyl)-1,3-dimethylindolin-2-one (1-3ao)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 69% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.20 (t, J = 7.8 Hz, 1H), 7.09 (d, J = 7.2 Hz, 1H), 7.00 (t, J = 7.8 Hz, 1H), 6.77 (d, J = 7.8 Hz, 1H), 3.34 (t, J = 6.6 Hz, 2H), 3.14 (s, 3H), 1.89 – 1.79 (m, 1H), 1.68 – 1.63 (m, 1H), 1.59 – 1.54 (m, 2H), 1.30 – 1.17 (m, 5H), 0.97 – 0.89 (m, 1H), 0.85 – 0.71 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 143.3, 134.1, 127.7, 122.5, 122.4, 107.9, 48.4, 44.9, 38.2, 32.2, 26.9, 26.1, 23.9, 23.8.

HRMS (ESI-MS) Calcd. For C₁₅H₂₀ClNONa [M+Na]⁺ 288.11256, found: 288.11238.

IR (neat, cm⁻¹): v: 2932, 1707, 1611, 1466, 1346, 1250, 1124, 1018, 931, 749, 700.

3-((R)-4-((3R,5R,8R,9S,10S,13R,14S,17R)-3-Hydroxy-10,13-dimethylhexadecahydro-1*H*-cyclopenta[a]phenanthren-17-yl)pentyl)-1,3- dimethylindolin-2-one (1-3ap)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 64 mg product was obtained by 63% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.22 – 7.17 (m, 1H), 7.10 (d, J = 7.2 Hz, 1H), 7.00 (t, J = 7.2 Hz, 1H), 6.77 (d, J = 7.8 Hz, 1H), 3.57 – 3.52 (m, 1H), 3.15 (s, 3H), 1.84 – 0.71 (m, 37H), 0.66 (d, J = 6.6 Hz, 3H), 0.51 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.9, 143.3, 134.4, 127.6, 122.5, 122.4, 107.86, 71.9, 56.5, 56.4, 48.5, 42.7, 42.1, 40.4, 40.2, 38.8, 36.5, 36.0, 35.8, 35.5, 35.4, 34.6, 30.6, 28.3, 27.2, 26.4, 26.1, 24.2, 23.8, 23.4, 21.2, 20.8, 18.5, 12.0.

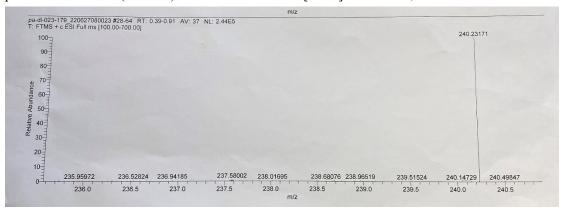
HRMS (ESI-MS) Calcd. For C₃₄H₅₁NO₂Na [M+Na]⁺ 528.38120, found: 528.37952.

IR (neat, cm⁻¹): v: 3414, 2928, 2245, 1701, 1612, 1464, 1375, 1251, 1123, 1037, 912, 852.

Mechanism studies

(1) The radical trapping experiment

A clean oven-dried tube equipped with a PTFE-coated stir bar was charged with arylamide **1-1a** (53 mg, 0.3 mmol), cyclohexyl redox-active ester **1-2a** (55 mg, 0.2 mmol), Ph₃P (10.5 mg, 0.04 mmol), NaI (6 mg, 0.04 mmol) and TEMPO (46.8 mg,0.3 mmol), then the system was degassed and filled with nitrogen gas three times before the solvent anhydrous MeCN (2 mL) was added. The reaction was carried out irradiated using 40 W 456 nm blue LEDs at room temperature under fan cooling. After 36 h, the solvent was removed in vacuo, the desired product **1-3aa** was not detected by NMR analysis of the crude reaction mixture. HRMS of the crude reaction mixture shows the existence of product **1-4**. HRMS (ESI-MS) Calcd. For $C_{15}H_{30}NO$ [M+H]⁺ 240.23219, found: 240.23171.



(2) The radical clock experiment

To a reaction tube with a stir bar, arylamide **1-1a** (53 mg, 0.3 mmol), redox-active ester **1-5** (49 mg, 0.2 mmol), PPh₃ (10.5 mg, 0.04 mmol), NaI (6 mg, 0.04 mmol) was added. After degassed and filled with nitrogen gas three times, the anhydrous solvent MeCN was added. The reaction was performed under blue LEDs irradiation (40 W, 456 nm) at room temperature under fan cooling. After 36 h, the crude reaction mixture was concentrated in vacuo and purified by silica gel column chromatography (Pentane/EA = 10:1) to obtain corresponding product. Colorless liquid: 30 mg, 65% yield. The compound data was in agreement with the compound **1-3am**.

4.2.2 Visible-light-induced photocatalytic deoxygenative benzylation of quinoxalin-2-

(1H)-ones with carboxylic acid anhydrides

General procedure for synthesis of 3-benzylquinoxalin-2(1H)-ones

To a glass vial equipped with a stirring bar, quinoxalin-2(1H)-one **2-1** (0.15 mmol, 1 equiv.), carboxylic acid anhydride **2-2** (0.3 mmol, 2 equiv.), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (0.0045 mmol, 3 mol%), Ph₃P (0.375 mmol, 2.5 equiv.) and DABCO (0.075 mmol, 50 mol%) were added. Then the vial with a screw cap and a silicone seal was evacuated and backfilled with N₂ (three times). After anhydrous acetonitrile (MeCN, 1.5 mL) was added, the reaction was performed under blue LEDs irradiation (40 W, 456 nm) at room temperature. After 12 h, saturated aqueous NaHCO₃ solution (15 mL) was added to the reaction. The mixture was stirred for another 1 h and then extracted with EA (3 × 5 mL). After being dried over anhydrous Na₂SO₄ and concentrated in vacuo, the crude product was purified by column chromatography using pentane/EA as eluent to give the 3-benzylquinoxalin-2(1H)-one **2-3**.

Larger (1 mmol) scale: To a 50 mL vial equipped with a stirring bar, quinoxalin-2(1H)-one **2-1** (1 mmol, 1 equiv.), carboxylic acid anhydride **2-2** (2 mmol, 2 equiv.), $Ir[dF(CF_3)ppy]_2(dtbbpy)PF_6$ (0.03 mmol, 3 mol%), Ph_3P (2.5 mmol, 2.5 equiv.) and DABCO (0.5 mmol, 50 mol%) were added. Then, the vial with sealed aluminous headspace cap was evacuated and backfilled with N_2 (three times). After anhydrous acetonitrile (MeCN, 10 mL) was added, the reaction was performed under blue LEDs irradiation (40 W, 456 nm) at room temperature for 24 h.

Product characterization

3-Benzyl-1-methylquinoxalin-2(1H)-one (2-3aa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 29 mg product was obtained by 77% isolated yield as yellow solid. Larger (1mmol) scale experiment: 182 mg, yellow solid (73%). Melting point: 87.3 - 88.5 °C.

¹H NMR (400 MHz, Chloroform-*d*) δ 7.76 (d, J = 8.0 Hz, 1H), 7.48 – 7.31 (m, 3H), 7.28 – 7.07 (m, 5H), 4.18 (s, 2H), 3.56 (s, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 159.3, 154.7, 137.1, 133.4, 132.8, 129.9, 129.9, 129.6, 128.4, 126.6, 123.6, 113.6, 40.8, 29.1.

3-Benzyl-6-fluoro-1-methylquinoxalin-2(1*H*)-one (2-3ba)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 29 mg product was obtained by 72% isolated yield as yellow solid. Melting point: 95.0 - 97.9 °C.

¹H NMR (400 MHz, Chloroform-*d*) δ 7.53 (dd, J = 8.8, 2.8 Hz, 1H), 7.44 (d, J = 7.2 Hz, 2H), 7.31 – 7.16 (m, 5H), 4.24 (s, 2H), 3.64 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 160.9, 158.7 (d, J = 241.6 Hz), 154.4, 136.7, 133.3 (d, J = 11.3 Hz), 130.0 (d, J = 2.3 Hz), 129.6, 128.4, 126.7, 117.5 (d, J = 24.0 Hz), 115.4 (d, J = 22.5 Hz), 114.6 (d, J = 8.9 Hz), 40.7, 29.4.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ -119.06 to -119.16 (m).

Data is in line with the literature.²³⁶

3-Benzyl-6-chloro-1-methylquinoxalin-2(1*H*)-one (2-3ca)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 33 mg product was obtained by 77% isolated yield as yellow solid. Melting point: 105.6 - 107.7 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.76 (d, J = 2.4 Hz, 1H), 7.42 – 7.32 (m, 3H), 7.22 (t, J = 7.8 Hz, 2H), 7.14 (t, J = 7.2 Hz, 1H), 7.11 (d, J = 8.4 Hz, 1H), 4.17 (s, 2H), 3.56 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 160.7, 154.4, 136.7, 133.3, 132.0, 129.8, 129.6, 129.3, 128.9, 128.5, 126.7, 114.7, 40.7, 29.3.

Data is in line with the literature. 234,297

3-Benzyl-6-bromo-1-methylquinoxalin-2(1H)-one (2-3da)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 75% isolated yield as yellow solid. Melting point: 116.4 - 118.1 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.92 (s, 1H), 7.51 (d, J = 9.0 Hz, 1H), 7.37 (d, J = 7.8 Hz, 2H), 7.22 (t, J = 7.2 Hz, 2H), 7.14 (t, J = 7.2 Hz, 1H), 7.04 (d, J = 9.0 Hz, 1H), 4.17 (s, 2H), 3.55 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 160.7, 154.4, 136.7, 133.6, 132.6, 132.5, 132.4, 129.6, 128.5, 126.7, 116.1, 115.0, 40.7, 29.3.

HRMS (ESI-MS) Calcd. For C₁₆H₁₃ON₂BrNa [M+Na]⁺ 351.0104, found: 351.0094.

IR (neat, cm⁻¹): v: 3289, 3059, 2928, 2645, 2326, 2110, 1884, 1754, 1645, 1586, 1455, 1289, 1200, 1097, 948, 866, 806, 761, 731, 700.

3-Benzyl-1-methyl-6-(trifluoromethyl)quinoxalin-2(1H)-one (2-3ea)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 38 mg product was obtained by 80% isolated yield as yellow solid. Melting point: 102.2 – 102.6 °C.

 1 H NMR (600 MHz, Chloroform-d) δ 8.06 (s, 1H), 7.65 (d, J = 8.4 Hz, 1H), 7.38 (d, J = 7.8 Hz, 2H), 7.28 (d, J = 9.0 Hz, 1H), 7.23 (t, J = 7.8 Hz, 2H), 7.15 (t, J = 7.2 Hz, 1H), 4.20 (s, 2H), 3.60 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 161.1, 154.6, 136.4, 135.7, 132.1, 129.6, 128.5, 127.4 (q, J = 3.9 Hz), 126.8, 126.2 (q, J = 3.6 Hz), 125.9 (q, J = 33.6 Hz), 123.7 (q, J = 272.1 Hz), 114.2, 40.7, 29.4.

¹⁹F NMR (565 MHz, Chloroform-d) δ -61.96.

HRMS (ESI-MS) Calcd. For C₁₇H₁₃F₃N₂ONa [M+Na]⁺ 341.0872, found: 341.0866.

IR (neat, cm⁻¹): v: 3066, 2930, 2649, 2320, 2027, 1946, 1826, 1735, 1657, 1492, 1360, 1236, 1118, 1029, 913, 821, 700, 658.

3-Benzyl-6-(tert-butyl)-1-methylquinoxalin-2(1H)-one (2-3fa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 36 mg product was obtained by 78% isolated yield as yellow solid. Melting point: 126.6 - 127.2 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.78 (d, J = 2.4 Hz, 1H), 7.49 (dd, J = 8.4, 2.4 Hz, 1H), 7.39 (d, J = 6.6 Hz, 2H), 7.21 (t, J = 7.8 Hz, 2H), 7.13 (d, J = 8.4 Hz, 2H), 4.19 (s, 2H), 3.57 (s, 3H), 1.30 (s, 9H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.2, 154.8, 147.0, 137.2, 132.5, 131.1, 129.5, 128.4, 127.6, 126.6, 126.4, 113.2, 40.9, 34.5, 31.3, 29.1.

HRMS (ESI-MS) Calcd. For C₂₀H₂₂N₂ONa [M+Na]⁺ 329.1624, found: 329.1616.

IR (neat, cm⁻¹): v: 2958, 2870, 2322, 2083, 1895, 1659, 1494, 1292, 1086, 966, 893, 735, 698.

3-Benzyl-6-methoxy-1-methylquinoxalin-2(1H)-one (2-3ga)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 26 mg product was obtained by 62% isolated yield as yellow solid. Melting point: 127.2 – 129.3 °C.

¹H NMR (400 MHz, Chloroform-d) δ 7.39 (d, J = 7.3 Hz, 2H), 7.27 – 7.17 (m, 3H), 7.16 – 7.02 (m, 3H), 4.19 (s, 2H), 3.80 (s, 3H), 3.57 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.9, 156.0, 154.4, 137.1, 133.5, 129.6, 128.4, 127.6, 126.6, 119.1, 114.5, 111.4, 55.8, 40.8, 29.3.

HRMS (EI-MS) Calcd. For C₁₇H₁₆N₂O₂ 280.1212, found: 280.1200.

IR (neat, cm $^{-1}$): \tilde{v} : 3067, 2927, 2845, 2653, 2287, 2185, 2096, 1940, 1817, 1731, 1650, 1686, 1496, 1453, 1341, 1282, 1219, 1151, 1028, 963, 889, 744, 697.

Data is in line with the literature.²³⁵

$$CI = \bigcup_{i=1}^{N} \bigcup_{j=1}^{N} \bigcup_{j=1}^{N} \bigcup_{i=1}^{N} \bigcup_{j=1}^{N} \bigcup_{j=1}^{N}$$

3-Benzyl-7-chloro-1-methylquinoxalin-2(1H)-one (2-3ha)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 33 mg product was obtained by 77% isolated yield as yellow solid. Melting point: 161.1 - 164.1 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.68 (d, J = 8.4 Hz, 1H), 7.37 (d, J = 6.6 Hz, 2H), 7.24 – 7.18 (m, 3H), 7.18 – 7.11 (m, 2H), 4.16 (s, 2H), 3.54 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.4, 154.4, 136.8, 135.8, 134.2, 131.3, 131.0, 129.6, 128.4, 126.7, 123.9, 113.6, 40.7, 29.2.

Data is in line with the literature.²⁹⁸

3-Benzyl-7-bromo-1-methylquinoxalin-2(1H)-one (2-3ia)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 38 mg product was obtained by 77% isolated yield as yellow solid. Melting point: 177.6 - 179.2 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.60 (d, J = 8.4 Hz, 1H), 7.39 – 7.30 (m, 4H), 7.21 (t, J = 7.8 Hz, 2H), 7.13 (t, J = 7.4 Hz, 1H), 4.15 (s, 2H), 3.53 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.7, 154.4, 136.7, 134.4, 131.6, 131.2, 129.6, 128.5, 126.8, 126.7, 123.9, 116.6, 40.7, 29.2.

HRMS (ESI-MS) Calcd. For C₁₆H₁₃ON₂BrNa [M+Na]⁺ 351.0104, found: 351.0098.

IR (neat, cm⁻¹): v: 3294, 3077, 2926, 2327, 2107, 1991, 1914, 1652, 1589, 1553, 1451, 1297, 1213, 1100, 1074, 1007, 935, 830, 733, 696, 667.

3-Benzyl-7-(tert-butyl)-1-methylquinoxalin-2(1H)-one (2-3ja)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 37 mg product was obtained by 81% isolated yield as yellow solid. Melting point: 78.4 - 78.9 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.71 (d, J = 8.4 Hz, 1H), 7.37 (d, J = 7.2 Hz, 2H), 7.31 (dd, J = 8.4, 1.8 Hz, 1H), 7.19 (t, J = 7.8 Hz, 2H), 7.14 (d, J = 1.8 Hz, 1H), 7.11 (t, J = 7.8 Hz, 1H), 4.17 (s, 2H), 3.60 (s, 3H), 1.31 (s, 9H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 158.4, 155.0, 153.8, 137.3, 133.0, 130.9, 129.5, 129.4, 128.4, 126.5, 121.4, 110.0, 40.8, 35.4, 31.3, 29.0.

HRMS (ESI-MS) Calcd. For C₂₀H₂₂N₂ONa [M+Na]⁺ 329.1624, found: 329.1615.

IR (neat, cm $^{-1}$): \tilde{v} : 3030, 2960, 2870, 2290, 2075, 1906, 1652, 1455, 1308, 1257, 1094, 1009, 949, 829, 738, 698, 657.

$$\bigcap_{C|} \bigcap_{N \to 0}$$

3-Benzyl-8-chloro-1-methylquinoxalin-2(1*H*)-one (2-3ka)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 15 mg product was obtained by 35% isolated yield as yellow solid. Melting point: 98.6 - 99.7 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.74 (dd, J = 7.8, 1.6 Hz, 1H), 7.51 (dd, J = 7.8, 1.6 Hz, 1H),

7.44 (d, J = 7.8 Hz, 2H), 7.29 (t, J = 7.8 Hz, 2H), 7.21 (t, J = 7.8 Hz, 2H), 4.24 (s, 2H), 3.99 (s, 3H). ¹³C NMR (151 MHz, Chloroform-d) δ 159.5, 155.8, 136.6, 135.2, 133.1, 131.6, 129.6, 129.5, 128.4, 126.7, 123.9, 119.5, 40.6, 35.6.

HRMS (ESI-MS) Calcd. For C₁₆H₁₃ClN₂ONa [M+Na]⁺ 307.0609, found: 307.0610.

IR (neat, cm⁻¹): v: 3296, 2927, 2584, 2324, 2098, 1936, 1788, 1723, 1650, 1598, 1551, 1454, 1337, 1263, 1186, 1098, 999, 898, 730.

3-Benzyl-5-chloro-1-methylquinoxalin-2(1H)-one (2-3la)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 21 mg product was obtained by 49% isolated yield as yellow solid. Melting point: 139.8 – 141.5 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.43 (d, J = 7.8 Hz, 2H), 7.33 – 7.31 (m, 2H), 7.22 (t, J = 7.8 Hz, 2H), 7.13 (t, J = 7.2 Hz, 1H), 7.09 – 7.07 (m, 1H), 4.24 (s, 2H), 3.57 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.8, 154.3, 136.7, 134.8, 134.8, 129.8, 129.7, 129.5, 128.4, 126.7, 124.5, 112.4, 40.8, 29.5.

HRMS (ESI-MS) Calcd. For C₁₆H₁₃ClN₂ONa [M+Na]⁺ 307.0609, found: 307.0602.

IR (neat, cm⁻¹): v: 3298, 3081, 2926, 2855, 2323, 2066, 1912, 1744, 1654, 1586, 1461, 1292, 1109, 1074, 922, 839, 697.

3-Benzyl-1,5-dimethylquinoxalin-2(1*H*)-one (2-3ma)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 30 mg product was obtained by 76% isolated yield as yellow solid. Melting point: 112.4 – 113.1 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.39 (d, J = 7.2 Hz, 2H), 7.29 (t, J = 7.8 Hz, 1H), 7.21 (t, J = 7.8 Hz, 2H), 7.12 (t, J = 7.2 Hz, 1H), 7.09 (d, J = 7.8 Hz, 1H), 7.00 (d, J = 8.4 Hz, 1H), 4.19 (s, 2H), 3.56 (s, 3H), 2.58 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 157.3, 154.7, 138.7, 137.4, 133.4, 131.3, 129.6, 129.5, 128.3, 126.4, 124.8, 111.4, 40.6, 29.2, 17.5.

HRMS (ESI-MS) Calcd. For C₁₇H₁₆N₂ONa [M+Na]⁺ 287. 1155, found: 287.1150.

IR (neat, cm⁻¹): v: 3274, 3028, 2925, 2652, 2323, 2027, 1948, 1748, 1640, 1480, 1374, 1210, 1151, 1037, 939, 835, 781, 698.

3-Benzyl-6,7-difluoro-1-methylquinoxalin-2(1H)-one (2-3na)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 33 mg product was obtained by 77% isolated yield as yellow solid. Melting point: 96.7 - 97.9 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.58 (dd, J = 10.2, 8.4 Hz, 1H), 7.36 (d, J = 7.2 Hz, 2H), 7.22

(t, J = 7.8 Hz, 2H), 7.14 (t, J = 7.2 Hz, 1H), 6.98 (dd, J = 11.4, 7.2 Hz, 1H), 4.15 (s, 2H), 3.53 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.9 (d, J = 3.5 Hz), 154.3, 151.2 (dd, J = 253.1 14.2 Hz), 146.7 (dd, J = 247.0, 13.9 Hz), 136.6, 130.5 (dd, J = 8.6, 2.0 Hz), 129.6, 129.0 (dd, J = 9.4, 3.2 Hz), 128.5, 126.8, 117.6 (dd, J = 17.8, 2.1 Hz), 102.2 (d, J = 23.0 Hz), 40.6, 29.6.

 19 F NMR (376 MHz, Chloroform-*d*) δ -131.17 to -131.38 (m), -142.09 to -142.33 (m).

Data is in line with the literature.²³⁸

3-Benzyl-6,7-dichloro-1-methylquinoxalin-2(1H)-one (2-30a)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 33 mg product was obtained by 69% isolated yield as yellow solid. Melting point: 181.9 - 184.0 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.86 (s, 1H), 7.36 (d, J = 7.8 Hz, 2H), 7.23 (t, J = 7.8 Hz, 2H), 7.19 (s, 1H), 7.15 (t, J = 7.2 Hz, 1H), 4.16 (s, 2H), 3.54 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 160.8, 154.1, 136.4, 133.9, 132.7, 131.8, 130.8, 129.6, 128.5, 127.4, 126.8, 115.1, 40.6, 29.4.

Data is in line with the literature.²³¹

3-Benzyl-6,7-dibromo-1-methylquinoxalin-2(1*H*)-one (2-3pa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 11 mg product was obtained by 18% isolated yield as yellow solid. Melting point: 205.5 - 207.3 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 8.07 (s, 1H), 7.52 (s, 1H), 7.42 (d, J = 7.8 Hz, 2H), 7.29 (t, J = 7.8 Hz, 2H), 7.22 (t, J = 7.8 Hz, 1H), 4.22 (s, 2H), 3.60 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 161.0, 154.1, 136.3, 133.9, 133.2, 132.5, 129.6, 128.5, 126.8, 126.1, 118.7, 118.2, 40.7, 29.3.

HRMS (EI-MS) Calcd. For C₁₆H₁₂Br₂N₂O 405.9311, found: 405.9311.

IR (neat, cm⁻¹): v: 3312, 3088, 2923, 2854, 2325, 2091, 1900, 1740, 1661, 1548, 1454, 1388, 1289, 1198, 1004, 906, 884, 747, 700.

3-Benzyl-1,6,7-trimethylquinoxalin-2(1H)-one (2-3qa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 34 mg product was obtained by 81% isolated yield as yellow solid. Melting point: 171.0 - 171.8 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.60 (s, 1H), 7.45 (d, J = 7.2 Hz, 2H), 7.28 (t, J = 7.8 Hz, 2H), 7.19 (tt, J = 7.2, 1.2 Hz, 1H), 7.02 (s, 1H), 4.24 (s, 2H), 3.63 (s, 3H), 2.39 (s, 3H), 2.34 (s, 3H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 158.0, 154.8, 139.6, 137.4, 132.4, 131.3, 131.2, 130.0, 129.5, 128.3, 126.5, 114.1, 40.7, 29.0, 20.5, 19.1.

Data is in line with the literature.²³⁸

3-Benzyl-1-methylbenzo[g]quinoxalin-2(1H)-one (2-3ra)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 18 mg product was obtained by 40% isolated yield as yellow solid. Melting point: 196.4 - 198.0 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 8.35 (s, 1H), 7.96 (d, J = 8.4 Hz, 1H), 7.88 (d, J = 8.4 Hz, 1H), 7.58 – 7.53 (m, 2H), 7.52 – 7.45 (m, 3H), 7.31 (t, J = 7.8 Hz, 2H), 7.22 (tt, J = 7.2, 1.2 Hz, 1H), 4.31 (s, 2H), 3.72 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.8, 154.6, 136.9, 133.5, 132.1, 131.9, 129.7, 129.6, 129.0, 128.4, 128.4, 127.7, 127.1, 126.6, 125.2, 109.9, 40.8, 29.1.

HRMS (ESI-MS) Calcd. For C₂₀H₁₆N₂ONa [M+Na]⁺ 323.1155, found: 323.1150.

IR (neat, cm⁻¹): v: 3283, 3057, 2925, 2855, 2651, 2321, 2078, 1816, 1594, 1493, 1359, 1279, 1162, 1036, 996, 865, 746, 668.

$$\left(\begin{array}{c} \\ \\ \\ \\ \\ \end{array} \right) \left(\begin{array}{$$

2-Benzyl-4-methylpyrido[2,3-b]pyrazin-3(4H)-one (2-3sa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 31 mg product was obtained by 82% isolated yield as yellow solid. Melting point: 112.5 - 114.0 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 8.46 (dd, J = 4.8, 1.8 Hz, 1H), 8.05 (dd, J = 7.8, 1.8 Hz, 1H), 7.38 (d, J = 7.2 Hz, 2H), 7.25 – 7.18 (m, 3H), 7.14 (t, J = 7.2 Hz, 1H), 4.20 (s, 2H), 3.71 (s, 3H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 160.7, 156.0, 149.0, 144.2, 137.2, 136.6, 129.6, 12 128.5, 128.1, 126.8, 119.5, 40.6, 27.8.

Data is in line with the literature.²³³

3-Benzyl-1-ethylquinoxalin-2(1H)-one (2-3ta)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 32 mg product was obtained by 81% isolated yield as yellow solid. Melting point: 67.4 - 68.2 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.78 (dd, J = 7.8, 1.6 Hz, 1H), 7.45 – 7.37 (m, 3H), 7.26 – 7.18 (m, 4H), 7.13 (t, J = 7.2 Hz, 1H), 4.20 (q, J = 7.2 Hz, 2H), 4.19 (s, 2H), 1.26 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.4, 154.2, 137.2, 133.1, 132.3, 130.2, 129.8, 129.6, 128.4, 126.5, 123.4, 113.4, 40.6, 37.4, 12.4.

3-Benzyl-1-butylquinoxalin-2(1H)-one (2-3ua)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 36 mg product was obtained by 82% isolated yield as yellow solid. Melting point: 77.1 - 78.3 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.77 (dd, J = 7.8, 1.6 Hz, 1H), 7.45 – 7.35 (m, 3H), 7.25 – 7.16 (m, 4H), 7.12 (tt, J = 7.2, 1.2 Hz, 1H), 4.19 (s, 2H), 4.11 (t, J = 7.8 Hz, 2H), 1.66 – 1.59 (m, 2H), 1.42 – 1.33 (m, 2H), 0.90 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.3, 154.5, 137.2, 133.1, 132.5, 130.2, 129.8, 129.5, 128.4, 126.5, 123.3, 113.6, 42.2, 40.7, 29.3, 20.3, 13.8.

Data is in line with the literature.²³⁸

1-Allyl-3-benzylquinoxalin-2(1H)-one (2-3va)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 34 mg product was obtained by 82% isolated yield as yellow solid. Melting point: 79.0 - 81.9 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.78 (dd, J = 7.8, 1.8 Hz, 1H), 7.42 – 7.36 (m, 3H), 7.26 – 7.19 (m, 3H), 7.16 (dd, J = 8.4, 1.2 Hz, 1H), 7.13 (tt, J = 7.2, 1.2 Hz, 1H), 5.86 – 5.78 (m, 1H), 5.16 (broad d, J = 10.2 Hz, 1H), 5.06 (broad d, J = 10.2 Hz, 1H), 4.78 (dt, J = 5.4, 1.8 Hz, 2H), 4.20 (s, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.4, 154.3, 137.1, 133.0, 132.6, 130.7, 130.1, 129.8, 129.5, 128.4, 126.6, 123.6, 118.1, 114.1, 44.6, 40.7.

Data is in line with the literature.²³⁸

3-Benzyl-1-(prop-2-yn-1-yl)quinoxalin-2(1H)-one (2-3wa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 34 mg product was obtained by 83% isolated yield as yellow solid. Melting point: 153.0 - 155.7 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.78 (dd, J = 7.8, 1.8 Hz, 1H), 7.47 (\approx t, J = 7.8 Hz, 1H), 7.39 (d, J = 7.8 Hz, 2H), 7.34 (d, J = 8.4 Hz, 1H), 7.27 (\approx t, J = 7.2 Hz, 1H), 7.22 (t, J = 7.8 Hz, 2H), 7.14 (t, J = 7.4 Hz, 1H), 4.93 (d, J = 2.4 Hz, 2H), 4.19 (s, 2H), 2.19 (t, J = 2.4 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.2, 153.7, 136.8, 133.0, 131.8, 130.1, 130.0, 129.6, 128.4, 126.7, 124.0, 114.1, 76.8, 73.3, 40.7, 31.5.

Methyl 2-(3-benzyl-2-oxoquinoxalin-1(2H)-yl)acetate (2-3xa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 32 mg product was obtained by 69% isolated yield as yellow solid. Melting point: 104.5 - 105.7 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.79 (d, J = 7.8 Hz, 1H), 7.44 – 7.34 (m, 3H), 7.26 (t, J = 7.8 Hz, 1H), 7.21 (t, J = 7.2 Hz, 2H), 7.13 (t, J = 7.2 Hz, 1H), 6.95 (d, J = 8.4 Hz, 1H), 4.92 (s, 2H), 4.20 (s, 2H), 3.67 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 167.6, 159.1, 154.3, 136.8, 132.9, 132.4, 130.4, 130.1, 129.5, 128.4, 126.6, 123.9, 113.0, 52.8, 43.5, 40.6.

Data is in line with the literature.²³⁸

3-Benzyl-1-(2-oxo-2-phenylethyl)quinoxalin-2(1H)-one (2-3ya)

The crude mixture was purified by silica gel column chromatography with 14 pentane/EA (8:1). 46 mg product was obtained by 87% isolated yield as yellow solid. Melting point: 140.9 - 141.2 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 8.06 – 8.00 (m, 2H), 7.88 (dd, J = 7.8, 1.8 Hz, 1H), 7.65 (tt, J = 7.8, 1.2 Hz, 1H), 7.52 (t, J = 7.8 Hz, 2H), 7.46 (d, J = 7.8 Hz, 2H), 7.40 (ddd, J = 8.4, 7.2, 1.6 Hz, 1H), 7.32 – 7.28 (m, 3H), 7.21 (tt, J = 7.8, 1.2 Hz, 1H), 6.91 (dd, J = 8.4, 1.2 Hz, 1H), 5.68 (s, 2H), 4.29 (s, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 191.1, 159.0, 154.5, 136.9, 134.5, 134.3, 132.9, 132.7, 130.2, 129.9, 129.5, 129.0, 128.4, 128.1, 126.6, 123.7, 113.5, 48.5, 40.7.

Data is in line with the literature.²³¹

1,3-Dibenzylquinoxalin-2(1*H*)-one (2-3za)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 43 mg product was obtained by 88% isolated yield as yellow solid. Melting point: 136.5 - 138.2 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.77 (dd, J = 7.8, 1.5 Hz, 1H), 7.41 (d, J = 7.8 Hz, 2H), 7.30 (≈t, J = 7.8 Hz, 1H), 7.25 – 7.17 (m, 5H), 7.17 – 7.08 (m, 5H), 5.37 (s, 2H), 4.25 (s, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.5, 154.9, 137.1, 135.2, 133.1, 132.7, 130.1, 129.9, 129.6, 128.9, 128.5, 127.7, 126.8, 126.6, 123.6, 114.4, 46.0, 40.8.

2-(3-(3-Benzyl-2-oxoquinoxalin-1(2H)-yl)propoxy)-3-methoxybenzaldehyde (2-3aaa)

The crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 24 mg product was obtained by 37% isolated yield as viscous yellow oil.

¹H NMR (600 MHz, Chloroform-*d*) δ 10.35 (s, 1H), 7.93 (d, J = 8.4 Hz, 1H), 7.73 (d, J = 8.4 Hz, 1H), 7.54 (t, J = 7.8 Hz, 1H), 7.47 (t, J = 7.8 Hz, 1H), 7.35 (dd, J = 7.2, 2.4 Hz, 1H), 7.21 (d, J = 7.8 Hz, 2H), 7.11 – 7.97 (m, 5H), 4.61 (t, J = 6.0 Hz, 2H), 4.23 (s, 2H), 4.07 (t, J = 6.6 Hz, 2H), 3.69 (s, 3H), 2.21 (p, J = 6.6 Hz, 2H).

 13 C NMR (151 MHz, Chloroform-*d*) δ 190.1, 155.9, 153.0, 151.7, 149.3, 140.1, 138.6, 137.6, 130.0, 129.3, 129.0, 128.5, 128.3, 126.8, 126.5, 126.4, 124.2, 119.3, 118.0, 71.6, 63.2, 55.9, 40.4, 29.6. Data is in line with the literature.

2,4,6-Tribenzyl-1,2,4-triazine-3,5(2*H*,4*H*)-dione (2-3aba)

The crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 50 mg product was obtained by 87% isolated yield as yellow solid. Melting point: 99.8 – 100.7 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.36 (d, J = 7.2 Hz, 2H), 7.28 (d, J = 6.6 Hz, 2H), 7.27 – 7.11 (m, 11H), 4.99 (s, 2H), 4.95 (s, 2H), 3.82 (s, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 155.8, 149.0, 144.5, 136.2, 135.8, 135.6, 129.5, 129.3, 128.8, 128.7, 128.6, 128.5, 128.2, 128.1, 126.9, 55.2, 44.3, 36.6.

HRMS (ESI-MS) Calcd. For C₂₄H₂₁N₃O₂Na [M+Na]⁺ 406.1526, found: 406.1522.

IR (neat, cm⁻¹): v: 3031, 2853, 2323, 2048, 1954, 1891, 1709, 1654, 1597, 1493, 1349, 1263, 1143, 1073, 915, 822, 696.

3-(4-Fluorobenzyl)-1-methylquinoxalin-2(1*H*)-one (2-3ab)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 31 mg product was obtained by 77% isolated yield as yellow solid. Melting point: 112.8 - 113.3 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.77 (dd, J = 7.8, 1.8 Hz, 1H), 7.45 (ddd, J = 8.4, 7.2, 1.6 Hz, 1H), 7.38 – 7.32 (m, 2H), 7.26 (\approx t, J = 8.4 Hz, 1H), 7.19 (d, J = 8.4 Hz, 1H), 6.93 – 6.84 (m, 2H), 4.15 (s, 2H), 3.59 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 161.8 (d, J = 244.5 Hz), 159.1, 154.7, 133.3, 132.7, 132.6 (d, J = 3.3 Hz), 131.0 (d, J = 7.9 Hz), 130.0, 130.0, 123.7, 115.2 (d, J = 21.3 Hz), 113.6, 39.9, 29.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -116.59 to -116.67 (m).

Data is in line with the literature.²³⁸

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3-(3-Fluorobenzyl)-1-methylquinoxalin-2(1H)-one (2-3ac)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 28 mg product was obtained by 70% isolated yield as yellow solid. Melting point: 77.7 - 78.8 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.85 (dd, J = 7.8, 1.8 Hz, 1H), 7.53 (ddd, J = 8.4, 7.2, 1.8 Hz, 1H), 7.34 (ddd, J = 8.4, 7.2, 1.2 Hz, 1H), 7.29 – 7.22 (m, 3H), 7.18 – 7.14 (broad d, J = 10.2 Hz, 1H), 6.91 – 6.87 (m, 1H), 4.25 (s, 2H), 3.67 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 162.8 (d, J = 245.7 Hz), 158.6, 154.7, 139.5 (d, J = 7.6 Hz), 133.3, 132.7, 130.1, 130.0, 129.7 (d, J = 8.2 Hz), 125.2 (d, J = 2.2 Hz), 123.7, 116.3 (d, J = 21.3 Hz), 113.6, 113.5 (d, J = 21.0 Hz), 40.4 (d, J = 1.8 Hz), 29.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -113.52 (td, J = 9.1, 5.1 Hz).

Data is in line with the literature.²³³

3-(2-Fluorobenzyl)-1-methylquinoxalin-2(1*H*)-one (2-3ad)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 32 mg product was obtained by 80% isolated yield as yellow solid. Melting point: 133.7 - 134.8 °C. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.72 (d, J = 7.9 Hz, 1H), 7.44 (t, J = 7.4 Hz, 1H), 7.29 (t, J = 7.3 Hz, 1H), 7.25 – 7.19 (m, 2H), 7.17 – 7.11 (m, 1H), 7.02 – 6.93 (m, 2H), 4.24 (s, 2H), 3.61 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 161.4 (d, J = 246.6 Hz), 158.1, 154.7, 133.3, 132.7, 131.6 (d, J = 4.2 Hz), 130.1, 130.0, 128.3 (d, J = 8.0 Hz), 124.2 (d, J = 15.7 Hz), 123.8 (d, J = 3.5 Hz), 123.6, 115.3 (d, J = 21.9 Hz), 113.5, 33.8 (d, J = 2.7 Hz), 29.1.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ -116.53 to -116.71 (m).

Data is in line with the literature.²³³

1-Methyl-3-(4-methylbenzyl)quinoxalin-2(1H)-one (2-3ae)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 25 mg product was obtained by 63% isolated yield as yellow solid. Melting point: 112.6 - 113.2 °C. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.88 (dd, J = 7.8, 1.6 Hz, 1H), 7.53 (ddd, J = 8.4, 7.2, 1.6 Hz, 1H), 7.42 – 7.32 (m, 3H), 7.29 – 7.26 (m, 1H), 7.12 (d, J = 7.8 Hz, 2H), 4.25 (s, 2H), 3.67 (s, 3H), 2.32 (s, 3H).

 $^{13}\mathrm{C}$ NMR (151 MHz, Chloroform-*d*) δ 159.5, 154.7, 136.2, 133.9, 133.4, 132.7, 129.9, 129.8, 129.4, 129.1, 123.6, 113.5, 40.4, 29.1, 21.1.

Data is in line with the literature.²³⁸

1-Methyl-3-(3-methylbenzyl)quinoxalin-2(1H)-one (2-3af)

The crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 30 mg product was obtained by 76% isolated yield as yellow solid. Melting point: 82.7 - 83.6 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.86 (dd, J = 7.8, 1.8 Hz, 1H), 7.51 (ddd, J = 8.4, 7.2, 1.8 Hz, 1H), 7.33 (ddd, J = 8.4, 7.2, 1.2 Hz, 1H), 7.30 – 7.23 (m, 3H), 7.18 (t, J = 7.8 Hz, 1H), 7.02 (d, J = 7.5 Hz, 1H), 4.23 (s, 2H), 3.65 (s, 3H), 2.32 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.4, 154.7, 138.0, 136.9, 133.4, 132.8, 130.2, 129.9, 129.8, 128.3, 127.4, 126.5, 123.5, 113.5, 40.7, 29.1, 21.4.

Data is in line with the literature.²³¹

1-Methyl-3-(2-methylbenzyl)quinoxalin-2(1*H*)-one (2-3ag)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 22 mg product was obtained by 55% isolated yield as yellow solid. Melting point: 92.9 – 93.7 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.73 (dd, J = 7.8, 1.6 Hz, 1H), 7.44 (ddd, J = 8.4, 7.4, 1.6 Hz, 1H), 7.28 – 7.22 (m, 2H), 7.19 (d, J = 8.4 Hz, 1H), 7.12 – 7.00 (m, 3H), 4.21 (s, 2H), 3.60 (s, 3H), 2.38 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.2, 154.8, 137.4, 135.5, 133.2, 132.7, 130.3, 130.1, 130.0, 129.8, 126.7, 125.8, 123.5, 113.5, 37.9, 29.1, 20.0.

Data is in line with the literature.²³¹

1-Methyl-3-(2-(trifluoromethyl)benzyl)quinoxalin-2(1H)-one (2-3ah)

The crude mixture was purified by silica gel column chromatography with pentane/EA (8:1). 40 mg product was obtained by 84% isolated yield as yellow solid. Melting point: 149.8 - 151.7 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.73 (broad d, J = 8.5 Hz, 1H), 7.69 (d, J = 7.8 Hz, 1H), 7.53 (≈t, J = 7.8 Hz, 1H), 7.46 (t, J = 7.8 Hz, 1H), 7.35 (t, J = 7.8 Hz, 1H), 7.32 – 7.28 (m, 3H), 4.49 (s, 2H), 3.71 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 158.4, 154.6, 135.8 (q, J = 1.5 Hz), 133.1, 132.6, 131.7, 131.5, 130.1, 130.0, 129.2 (q, J = 29.9 Hz), 126.6, 126.1 (q, J = 5.6 Hz), 124.5 (q, J = 274.2 Hz), 123.6, 113.6, 37.0 (q, J = 2.6 Hz), 29.1.

¹⁹F NMR (564 MHz, Chloroform-d) δ -59.95.

HRMS (ESI-MS) Calcd. For C₁₇H₁₃F₃N₂ONa [M+Na]⁺ 341.0872, found: 341.0870.

IR (neat, cm⁻¹): v: 3036, 2923, 2855, 2262, 2063, 1942, 1736, 1637, 1465, 1371, 1315, 1176, 1099,

989, 952, 754, 674.

3-(4-Methoxybenzyl)-1-methylquinoxalin-2(1H)-one (2-3ai)

The crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 16 mg product was obtained by 38% isolated yield as yellow solid. Melting point: 129.8 – 131.7 °C.

¹H NMR (600 MHz, Chloroform-d) δ 7.85 (dd, J = 7.8, 1.6 Hz, 1H), 7.50 (ddd, J = 8.4, 7.2, 1.6 Hz, 1H), 7.41 – 7.37 (m, 2H), 7.32 (ddd, J = 8.4, 7.2, 1.2 Hz, 1H), 7.25 (d, J = 8.0 Hz, 1H), 6.85 – 6.81 (m, 2H), 4.20 (s, 2H), 3.76 (s, 3H), 3.65 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.5, 158.3, 154.7, 133.3, 132.8, 130.5, 129.9, 129.8, 129.0, 123.5, 113.8, 113.5, 55.2, 39.9, 29.1.

Data is in line with the literature.²³⁸

1-Methyl-3-(naphthalen-2-ylmethyl)quinoxalin-2(1H)-one (2-3aj)

The crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 10 mg product was obtained by 22% isolated yield as yellow solid. Melting point: 131.0 - 132.7 °C.

 1 H NMR (600 MHz, Chloroform-d) δ 7.90 (s, 1H), 7.87 (dd, J = 7.8, 1.6 Hz, 1H), 7.80 – 7.75 (m, 3H), 7.62 (dd, J = 8.4, 1.8 Hz, 1H), 7.52 (ddd, J = 8.4, 7.2, 1.5 Hz, 1H), 7.44 – 7.37 (m, 2H), 7.34 (ddd, J = 8.4, 7.2, 1.2 Hz, 1H), 7.26 (dd, J = 8.4, 1.2 Hz, 1H), 4.43 (s, 2H), 3.65 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 159.2, 154.7, 134.6, 133.5, 133.4, 132.8, 132.3, 130.0, 129.9, 128.0, 128.0, 127.9, 127.7, 127.6, 125.8, 125.4, 123.6, 113.6, 40.9, 29.1.

Data is in line with the literature.²³⁸

Synthetic applications

A mixture of benzylation product 2-3xa (30.9 mg, 0.1 mmol) and hydrazine hydrate NH₂-NH₂·H₂O (0.02 mL, 0.4 mmol) in MeOH (2.5 mL) was heated at reflux for 3 h, then the reaction solution was concentrated in vacuo to give residue 2-4xa (25 mg), which was used without further purification in the next step.

To a solution of residue **2-4xa**, 4-hydroxyacetophenone (21.8 mg, 0.16 mmol) in EtOH (1.5 mL) was added acetic acid (0.02 mL). The resulting mixture was stirred at reflux and monitored by thin layer chromatography (TLC). When the reaction was completed and cooled to room temperature, the final product **2-5xa** was obtained by direct filtration as white solid: 32 mg, 75% yield from **2-3xa**.

Compound **2-5xa** (8:2 mixture of isomers). Melting point: > 220 °C.

Major isomer (80%): 1 H NMR (600 MHz, DMSO- d_6): δ 10.92 (s, 1H), 9.76 (s, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.73 (d, J = 8.3 Hz, 2H), 7.55 (t, J = 7.8 Hz, 1H), 7.38-7.33 (m, 4H), 7.30 (t, J = 7.5 Hz, 2H), 7.22 (t, J = 7.4 Hz, 1H), 6.80 (d, J = 8.4 Hz, 2H), 5.46 (s, 2H), 4.20 (s, 2H), 2.26 (s, 3H).

Minor isomer (20%): δ 10.76 (s, 1H), 9.76 (s, 1H), 7.63 (d, J = 8.3 Hz, 2H), 7.58 (t, J = 7.8 Hz, 1H), 7.46 (d, J = 8.4 Hz, 1H), 7.42-7.38 (m, 4H), 7.30 (t, J = 7.5 Hz, 2H), 7.22 (t, J = 7.4 Hz, 1H), 6.79 (d, J = 8.0 Hz, 2H), 5.15 (s, 2H), 4.18 (s, 2H), 2.29 (s, 3H).

 13 C NMR (151 MHz, DMSO- d_6 , major and minor together) δ 168.4, 163.3, 159.3, 159.2, 159.1, 159.0, 154.5, 154.5, 153.4, 149.5, 137.8, 137.7, 133.5, 132.4, 130.5, 129.6, 129.6, 129.3, 128.8, 128.4, 128.3, 126.9, 124.0, 123.9, 115.6, 115.5, 115.4, 44.6, 44.4, 14.6, 14.0 (some peaks overlapped with the solvent).

Data is in line with the literature.²³⁸

Under standard conditions, substrate 2-1x could react with 4-chlorobenzoic anhydride 2-2l to afford 2-3xl.

2-3xl, white solid, 30 mg, 58% yield. Melting point: 181.6 – 182.0 °C.

 1 H NMR (400 MHz, Chloroform-*d*) δ 7.78 (dd, J = 8.0, 1.6 Hz, 1H), 7.41 (td, J = 8.0, 1.6 Hz, 1H), 7.28 (t, J = 8.8 Hz, 3H), 7.21 – 7.15 (m, 2H), 6.96 (d, J = 8.3 Hz, 1H), 4.91 (s, 2H), 4.15 (s, 2H), 3.68 (s, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 167.5, 158.6, 154.3, 135.3, 132.8, 132.5, 132.4, 130.9, 130.4, 130.3, 128.6, 124.0, 113.0, 52.9, 43.4, 40.0.

Compound **2-3xl** was dissolved in THF (2 mL) and H_2O (2 mL), then LiOH (9.8 mg, 0.18 mmol) was added to reaction and stirred for 1 h. Then HCl (1 M) was added slowly until pH \approx 3 and the solution was filtered to give desired product **2-6xl**.

Compound **2-6xl**, yellow solid, 23 mg, 79% yield. Melting point: > 220 °C.

¹H NMR (400 MHz, DMSO- d_6) δ 7.79 (d, J = 8.0 Hz, 1H), 7.58 (t, J = 7.6 Hz, 1H), 7.47 (d, J = 8.4 Hz, 1H), 7.41 – 7.29 (m, 5H), 5.00 (s, 2H), 4.17 (s, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 169.2, 158.7, 154.2, 136.6, 133.0, 132.3, 131.6, 131.6, 130.8, 129.7, 128.7, 124.2, 115.1, 44.1, 39.3.

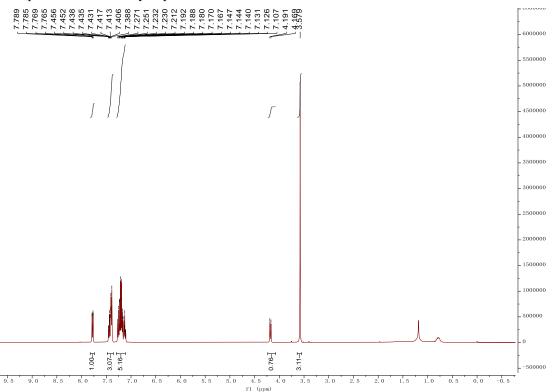
Data is in line with the literature.²⁴⁹

Control experiments

(1) Deuterium labeling experiments

Prepared according to standard conditions using MeCN-*d*₃ replacing anhydrous MeCN, ¹H NMR analysis showed no deuterated product.

To a vial equipped with a stirring bar, quinoxalin-2(1H)-one **2-1a** (24 mg, 0.15 mmol), carboxylic acid anhydride **2-2a** (67.9 mg, 0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (5.1 mg, 0.0045 mmol), Ph₃P (98.4 mg, 0.375 mmol) and DABCO (8.4 mg, 0.075 mmol) were added. The vial was capped. After evacuation and backfilling with N₂ three times, anhydrous MeCN (1.5 mL) and D₂O (5.4 uL, 0.3 mmol) were added via a syringe. The resulting solution was irradiated by 40W blue LEDs for 12 h at room temperature. Then saturated NaHCO₃ solution (15 mL) was added to the reaction system and stirred for 1 h. After extracting with EA (3 × 5 mL), the combined organic layer was dried by anhydrous Na₂SO₄, concentrated in vacuo, and purified by column chromatography (pentane/EA = 8:1) to afford 25 mg of the desired **2-3aa-d**. ¹H NMR analysis presented 62% deuterium incorporation at the benzylic position:



(2) Radical trapping experiment

To a reaction vial equipped with a stirring bar were added quinoxalin-2(1*H*)-one **2-1a** (24 mg, 0.15 mmol), carboxylic acid anhydride **2-2a** (67.9 mg, 0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (5.1 mg, 0.0045 mmol), Ph₃P (98.4 mg, 0.375 mmol), DABCO (8.4 mg, 0.075 mmol) and TEMPO (70.3 mg, 0.45 mmol). The capped vial was evacuated and backfilled with N₂ three times before anhydrous MeCN (1.5 mL) was added, then the reaction mixture was exposed under 25 W blue LEDs

irradiation at room temperature. After 12 h, only a trace amount of **2-3aa** was detected by ¹H NMR analysis of the crude reaction mixture. Trapped product **2-7a** was purified by silica gel column chromatography with pentane/EA (20:1). 11 mg product was obtained by 14% isolated yield as yellow solid.

 1 H NMR (600 MHz, Chloroform-*d*) δ 8.03 − 7.98 (m, 2H), 7.50 (tt, J = 7.4, 1.4 Hz, 1H), 7.39 (≈t, J = 7.8 Hz, 2H), 1.75 − 1.67 (m, 2H), 1.67 − 1.60 (m, 1H), 1.55 − 1.49 (m, 2H), 1.40 − 1.36 (m, 1H), 1.21 (s, 6H), 1.05 (s, 6H).

 13 C NMR (151 MHz, Chloroform-*d*) δ 166.4, 132.8, 129.8, 129.6, 128.5, 60.4, 39.1, 32.0, 20.9, 17.0. Data is in line with the literature.²⁹⁹

(3) Other control experiments

The compounds 2-8 and 2-9 were synthesized according to the literature. ^{248, 300}

2-8: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (\approx d, J = 8.0 Hz, 2H), 7.88 (\approx d, J = 8.2 Hz, 1H), 7.64 (\approx t, J = 7.9 Hz, 1H), 7.59 (\approx t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.5 Hz, 2H), 7.40 – 7.33 (m, 2H), 3.70 (s, 3H). Melting point: 150.8 – 152.2 °C.

¹³C NMR (151 MHz, Chloroform-*d*) δ 191.8, 154.7, 153.3, 134.9, 134.3, 133.9, 132.2, 132.1, 130.9, 130.0, 128.7, 124.2, 114.1, 29.1.

2-9: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 8.0, 1.6 Hz, 1H), 7.54 – 7.43 (m, 3H), 7.32 (\approx t, J = 7.7 Hz, 1H), 7.27 – 7.21 (m, 3H), 7.20 – 7.15 (m, 1H), 5.97 (s, 1H), 4.98 (broad s, 1H), 3.55 (s, 3H). Melting point: 91.7 – 92.8 °C.

¹³C NMR (101 MHz, Chloroform-*d*) δ 158.4, 153.6, 140.5, 133.4, 131.7, 130.7, 129.9, 128.4, 127.9, 127.3, 124.0, 113.8, 73.0, 29.0.

A mixture of **2-8** (39.7 mg, 0.15 mmol), carboxylic acid anhydride **2-2a** (67.9 mg, 0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (5.1 mg, 0.0045 mmol), Ph₃P (98.4 mg, 0.375 mmol), and DABCO (8.4 mg, 0.075 mmol) were added to a vial with a stirring bar. Then the vial with a screw cap and a silicone seal was evacuated and backfilled N₂ (three times). After anhydrous acetonitrile (MeCN, 1.5 mL) was added, the reaction mixture was exposed under blue LEDs irradiation (40 W, 456 nm) at room temperature. After 12 h, no desired product **2-3aa** was detected by TLC.

To a 10 mL vial equipped with a stirring bar, **2-9** (40 mg, 0.15 mmol), carboxylic acid anhydride **2-2a** (67.9 mg, 0.3 mmol), Ir[dF(CF₃)ppy]₂(dtbbpy)PF₆ (5.1 mg, 0.0045 mmol), Ph₃P (98.4 mg, 0.375 mmol), and DABCO (8.4 mg, 0.075 mmol) were added. Then the capped vial was evacuated and backfilled N₂ (three times). After anhydrous acetonitrile (MeCN, 1.5 mL) was added, the reaction mixture was exposed under blue LEDs irradiation (40 W, 456 nm) at room temperature for 12 h. Then saturated aqueous NaHCO₃ solution (15 mL) was added to the reaction. The mixture was stirred for 1 h and then extracted with EA (3 × 5 mL). After being dried over anhydrous Na₂SO₄ and

concentrated in vacuo, the crude product was purified by column chromatography using pentane/EA as eluent to give the 3-benzylquinoxalin-2(1*H*)-one **2-3aa** with 86% yield.

Stern-Volmer fluorescence studies

The Agilent Cary Eclipse Fluorescence Spectrometer is used to conduct Stern-Volmer fluorescence quenching analysis. The following parameters were employed: excitation wavelength = 400 nm, emission wavelength = 410 nm, excitation slit width = 10 nm, emission slit width = 10 nm, scan rate = 600 nm/min, averaging time = 0.1 s, data interval = 1 nm. The samples were measured at Hellma fluorescence QS quartz cuvette (chamber volume = 1.4 mL, light path = $10 \times 4 \text{ mm}$) with a closed screw cap and silicone seal.

All the solutions were prepared with dry degassed MeCN. The concentration of photocatalyst $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ is 2×10^{-5} M in MeCN, then by adding certain amount of a solution of quencher to $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ solution, the samples with different concentrations were obtained, and their fluorescence spectra were collected immediately with forementioned parameters. I_0 is the fluorescence intensity without quencher, I is the fluorescence intensity with quencher.

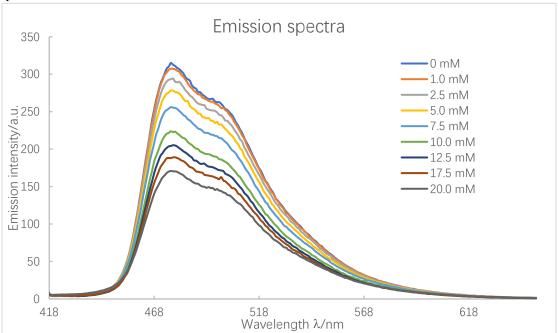


Figure 2-S1 Emission spectra of $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ with increasing concentrations of Ph_3P (The concentration of $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ is 1.8×10^{-5} M in MeCN)

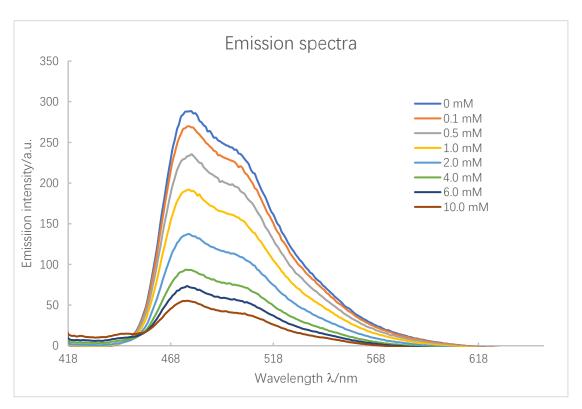


Figure 2-S2 Emission spectra of [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ with increasing concentrations of quinoxalinone 2-1a (The concentration of [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ is 1.8 × 10⁻⁵ M in MeCN)

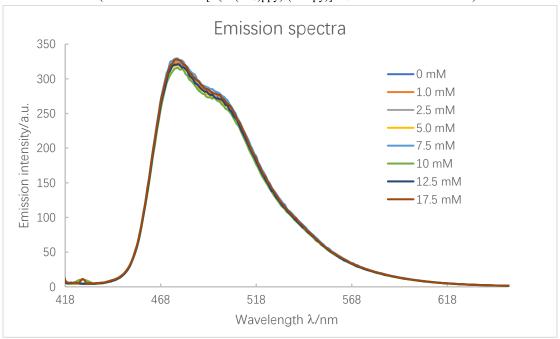


Figure 2-S3 Emission spectra of $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ with increasing concentrations of anhydride 2-2a (The concentration of $[Ir(dF(CF_3)ppy)_2(dtbbpy)]PF_6$ is 1.8×10^{-5} M in MeCN)

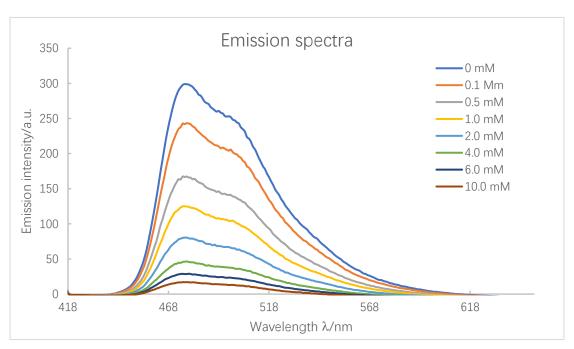


Figure 2-S4 Emission spectra of [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ with increasing concentrations of DABCO (The concentration of [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ is 1.8 × 10⁻⁵ M in MeCN)

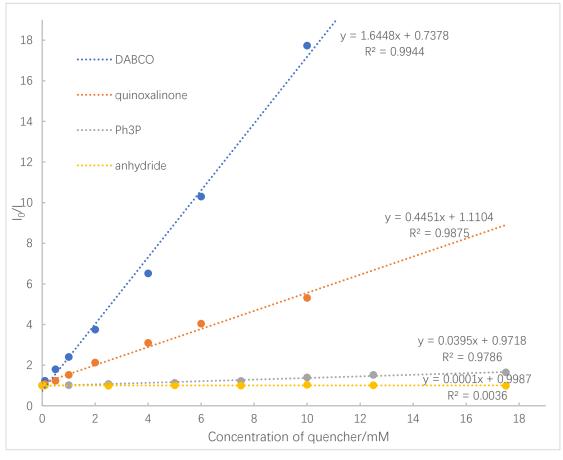


Figure 2-S5 Stern-Volmer plot of [Ir(dF(CF₃)ppy)₂(dtbbpy)]PF₆ with increasing concentrations of DABCO, quinoxalinone 2-1a, Ph₃P, and anhydride 2-2a

4.2.3 Visible-light-mediated radical α -C(sp³)-H gem-difluoroallylation of amides with trifluoromethyl alkenes via halogen atom transfer and 1,5-hydrogen atom transfer General procedure for α -C(sp³)-H gem-difluoroallylation of amides

Procedure A: To a glass vial equipped with a stirring bar, amide **3-1** (0.15 mmol, 1 equiv.) and photocatalyst Ir(ppy)₃ (0.003 mmol, 2 mol%) were added, then the capped vial was evacuated and backfilled N₂ for three times. Next, trifluoromethyl arene **3-2** (0.3 mmol, 2 equiv.) was added, as well as triazinane (0.3 mmol, 2 equiv.) and anhydrous MeCN (1.5 mL). The reaction system was then irradiated by 456 nm blue LEDs (40W) at room temperature for 12 h. Thereafter, the crude product was purified by column chromatography using pentane/EA or pentane/EA/CH₂Cl₂ as eluent to give target product **3-3**.

Procedure B: A glass vial with a stirring bar was charged with amide **3-1f** (0.15 mmol, 1 equiv.), trifluoromethyl arene **3-2** (0.45 mmol, 3 equiv.), Ir(ppy)₃ (0.003 mmol, 2 mol%) and 2.0 mL anhydrous MeCN. Then, the vial was sealed with an open-top screw cap fitted with a PTFE septum, and degassed by bubbling N₂ through the reaction solution for 3 min. After adding triazinane (0.3 mmol, 2 equiv.), the reaction system was exposed to 456 nm blue LEDs (40W) irradiation at room temperature for 12 h. The desired *gem*-difluoroalkene **3-3** could be obtained by column chromatography using pentane/EA as eluent.

Scale-up (1 mmol) procedure C: Amide **3-1** (1 mmol, 1 equiv.) and photocatalyst Ir(ppy)₃ (0.02 mmol, 2 mol%) were added to a glass vial equipped with a stirring bar, then the capped vial was evacuated and backfilled with N₂ for three times. Next, trifluoromethyl arene **3-2** (2 mmol, 2 equiv.) was added, as well as triazinane (2 mmol, 2 equiv.) and anhydrous MeCN (10 mL). The reaction system was then irradiated by 456 nm blue LEDs (40W) at room temperature for 24 h. Thereafter, the crude product was purified by column chromatography using pentane/EA as eluent to give target product **3-3**.



Product Characterization

5,5-Difluoro-2-(4-methoxyphenethyl)-N-methyl-N,4-diphenylpent-4-enamide (3-3aa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (6:1). 38 mg product was obtained, 58% isolated yield, as colorless liquid.

 1 H NMR (600 MHz, Chloroform-*d*) δ 7.20 – 7.09 (m, 6H), 6.89 (d, J = 7.2 Hz, 2H), 6.74 (d, J = 7.8 Hz, 2H), 6.68 (d, J = 7.8 Hz, 2H), 6.63 (d, J = 7.8 Hz, 2H), 3.70 (s, 3H), 3.13 (s, 3H), 2.61 – 2.56 (m, 1H), 2.49 – 2.45 (m, 1H), 2.44 – 2.33 (m, 2H), 2.27 – 2.23 (m, 1H), 1.85 – 1.80 (m, 1H), 1.58 – 1.47 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 174.6, 157.8, 154.0 (dd, J = 291.3 Hz, 288.3 Hz), 143.4, 133.4, 132.8 (t, J = 4.1 Hz), 129.6, 129.1, 128.4, 128.1 (t, J = 3.2 Hz), 127.5, 127.3, 127.1, 113.7, 90.2 (dd, J = 21.4 Hz, 13.7 Hz), 55.3, 39.2 (t, J = 2.4 Hz), 37.5, 32.8, 32.1, 29.8.

 19 F NMR (565 MHz, Chloroform-*d*) δ -89.12 (d, J = 39.2 Hz, 1F), -89.94 (d, J = 38.4 Hz, 1F).

HRMS (ESI-MS) Calcd. For C₂₇H₂₇F₂NO₂Na [M+Na]⁺ 458.1902, found: 458.1902.

IR (neat, cm⁻¹): \tilde{v} : 3480, 2929, 2856, 2320, 2090, 1885, 1725, 1651, 1595, 1447, 1343, 1298, 1176, 1118, 1033, 951, 914, 827, 764, 699, 664.

N-Benzyl-5,5-difluoro-2-(4-methoxyphenethyl)-N,4-diphenylpent-4-enamide (3-3ba)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 44 mg product was obtained by 57% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 – 7.14 (m, 3H), 7.13 – 7.03 (m, 6H), 6.99 (t, J = 7.8 Hz, 2H), 6.83 (d, J = 7.6 Hz, 2H), 6.69 (d, J = 8.4 Hz, 2H), 6.60 (d, J = 8.4 Hz, 2H), 6.40 (d, J = 7.8 Hz, 2H), 4.75 (d, J = 14.2 Hz, 1H), 4.72 (d, J = 14.1 Hz, 1H), 3.68 (s, 3H), 2.65 – 2.61 (m, 1H), 2.50 – 2.46 (m, 1H), 2.42 – 2.37 (m, 1H), 2.30 – 2.24 (m, 2H), 1.88 – 1.81 (m, 1H), 1.59 – 1.53 (m, 1H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 174.3, 157.8, 154.0 (dd, J = 291.7, 288.7 Hz), 141.6, 137.6, 133.3, 132.6 (t, J = 3.9 Hz), 129.3, 129.1, 128.9, 128.5, 128.4, 128.3, 128.0 (t, J = 3.3 Hz), 127.6, 127.4, 127.1, 113.8, 90.2 (dd, J = 21.4, 13.6 Hz), 55.3, 53.1, 39.3, 32.7, 32.0, 29.6.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.78 (d, J = 39.6 Hz, 1F), -89.88 (d, J = 40.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₃₃H₃₁F₂NO₂Na [M+Na]⁺ 534.2215, found: 534.2210.

IR (neat, cm⁻¹): v: 3060, 2928, 2325, 2081, 1886, 1725, 1651, 1595, 1446, 1298, 1240, 1177, 1074, 914, 823, 758, 698.

$$F_2C$$
 Ph

5,5-Difluoro-N-methyl-2-phenethyl-N,4-diphenylpent-4-enamide (3-3ca)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 33 mg product was obtained by 54% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.17 – 7.04 (m, 9H), 6.88 (d, J = 7.8 Hz, 2H), 6.85 – 6.82 (m, 2H), 6.70 – 6.65 (m, 2H), 3.13 (s, 3H), 2.59 (ddt, J = 14.4, 6.6, 3.0 Hz, 1H), 2.52 – 2.44 (m, 2H), 2.41 – 2.27 (m, 2H), 1.91 – 1.83 (m, 1H), 1.60 – 1.53 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 174.5, 154.0 (dd, J = 291.3, 288.3 Hz), 143.4, 141.3, 132.7 (t, J = 3.8 Hz), 129.6, 128.4, 128.3, 128.2, 128.1 (t, J = 3.3 Hz), 127.5, 127.3, 127.1, 125.8, 90.2 (dd, J = 21.4, 13.9 Hz), 39.2 (t, J = 2.4 Hz), 37.5, 33.0, 32.5, 29.9.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.13 (d, J = 39.0 Hz, 1F), -89.92 (d, J = 39.0 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₂₅F₂NONa [M+Na]⁺ 428.1796, found: 428.1791.

IR (neat, cm⁻¹): \tilde{v} : 3479, 3060, 2925, 2857, 2331, 2116, 1994, 1886, 1726, 1652, 1594, 1494, 1388, 1343, 1298, 1235, 1118, 1029, 950, 913, 842, 758, 698.

2-Benzyl-5,5-difluoro-N-methyl-N,4-diphenylpent-4-enamide (3-3da)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 36 mg product was obtained by 61% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.18 – 7.07 (m, 7H), 7.02 – 6.95 (m, 4H), 6.74 – 6.69 (m, 2H), 6.24 (broad s, 2H), 3.01 (s, 3H), 2.87 – 2.77 (m, 1H), 2.68 – 2.61 (m, 1H), 2.56 – 2.44 (m, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 174.0, 154.1 (dd, J = 291.9, 288.7 Hz), 143.1, 139.5, 132.8 (t, J = 3.8 Hz), 129.3, 129.1, 128.4, 128.3, 128.0 (t, J = 3.5 Hz), 127.4, 127.2, 127.1, 126.3, 90.3 (dd, J = 21.3, 13.7 Hz), 42.7 (t, J = 2.6 Hz), 38.2, 37.2, 29.9.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.82 (d, J = 37.9 Hz, 1F), -89.73 (d, J = 38.4 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₅H₂₃F₂NONa [M+Na]⁺ 414.1640, found: 414.1636.

IR (neat, cm⁻¹): \tilde{v} : 3475, 3060, 2924, 2857, 2329, 2109, 1949, 1807, 1726, 1652, 1594, 1494, 1447, 1389, 1340, 1297, 1236, 1118, 1022, 940, 915, 847, 760, 698.

$$F_2C$$
 Ph

5,5-Difluoro-N-methyl-N,4-diphenyl-2-propylpent-4-enamide (3-3ea)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (20:1). 32 mg product was obtained by 62% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 – 7.11 (m, 6H), 7.01 (d, J = 7.8, 2H), 6.83 – 6.79 (m, 2H), 3.13 (s, 3H), 2.61 – 2.55 (m, 1H), 2.47 – 2.43 (m, 1H), 2.35 – 2.29 (m, 1H), 1.56 – 1.47 (m, 1H), 1.21 – 1.13 (m, 2H), 1.01 – 0.94 (m, 1H), 0.62 (t, J = 7.2 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.1, 154.0 (dd, J = 291.4, 288.4 Hz), 143.6, 133.1 (t, J = 4.1 Hz), 129.5, 128.4, 128.1 (t, J = 3.5 Hz), 127.5, 127.4, 127.1, 90.5 (dd, J = 21.3, 13.4 Hz), 39.8 (t, J = 2.6 Hz), 37.4, 34.2, 30.3, 20.4, 13.9.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.20 (d, J = 38.4 Hz, 1F), -90.08 (d, J = 39.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₁H₂₃F₂NONa [M+Na]⁺ 366.1640, found: 366.1635.

IR (neat, cm⁻¹): v: 3484, 3060, 2958, 2869, 2332, 2108, 1885, 1726, 1653, 1594, 1495, 1388, 1340, 1301, 1234, 1118, 998, 765, 698.

2-(Cyclohexylmethyl)-5,5-difluoro-N-methyl-N,4-diphenylpent-4-enamide (3-3fa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 40 mg product was obtained by 67% isolated yield as colorless liquid.

Following procedure C (1 mmol scale), the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 214 mg product was obtained, 54% isolated yield.

 1 H NMR (600 MHz, Chloroform-d) δ 7.18 – 7.11 (m, 6H), 7.00 (d, J = 7.8 Hz, 2H), 6.81 – 6.74 (m, 2H), 3.13 (s, 3H), 2.58 – 2.54 (m, 1H), 2.48 – 2.43 (m, 1H), 2.41 – 2.35(m, 1H), 1.51 – 1.42 (m, 4H), 1.33 – 1.28 (m, 1H), 1.06 – 0.90 (m, 6H), 0.59 – 0.37 (m, 2H).

¹³C NMR (151 MHz, Chloroform-d) δ 175.4, 154.0 (dd, J = 290.8, 288.3 Hz), 143.5, 133.0 (t, J = 3.8 Hz), 129.4, 128.3, 128.2 (t, J = 3.3 Hz), 127.5, 127.3, 127.2, 90.6 (dd, J = 21.7, 14.0 Hz), 39.6, 37.5, 37.3 (t, J = 2.6 Hz), 35.3, 33.4, 33.1, 30.4, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.64 (d, J = 39.6 Hz, 1F), -90.40 (d, J = 40.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₅H₂₉F₂NONa [M+Na]⁺ 420.2109, found: 420.2104.

IR (neat, cm⁻¹): \tilde{v} : 3480, 3060, 2922, 2852, 2663, 2334, 2132, 1950, 1884, 1728, 1654, 1595, 1446, 1341, 1236, 1118, 1009, 941, 839, 764, 698, 665.

2-(4-Cyanobenzyl)-5,5-difluoro-N-methyl-N,4-diphenylpent-4-enamide (3-3ga)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (4:1). 38 mg product was obtained by 60% isolated yield as white solid. Melting point: 89.2 - 90.5 °C.

 1 H NMR (600 MHz, Chloroform-*d*) δ 7.40 (d, J = 8.2 Hz, 2H), 7.19 – 7.12 (m, 4H), 7.06 (t, J = 7.8 Hz, 2H), 6.97 – 6.92 (m, 2H), 6.81 (d, J = 7.8 Hz, 2H), 6.32 (broad s, 2H), 3.02 (s, 3H), 2.91 – 2.84 (m, 1H), 2.67 – 2.61 (m, 1H), 2.56 – 2.47 (m, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 173.2, 154.2 (dd, J = 292.3, 288.9 Hz), 145.4, 142.9, 132.4 (t, J = 3.6 Hz), 132.0, 129.8, 129.5, 128.5, 128.0 (t, J = 3.5 Hz), 127.8, 127.4, 127.0, 118.9, 110.2,

89.9 (dd, J = 21.1, 14.2 Hz), 42.4 (t, J = 2.7 Hz), 37.8, 37.3, 30.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.66 (d, J = 39.3 Hz, 1F), -89.28 (d, J = 38.4 Hz, 1F). HRMS (EI-MS) Calcd. For $C_{26}H_{22}F_2N_2O$ 416.1695, found: 416.1693.

IR (neat, cm⁻¹): \tilde{v} : 3495, 3060, 2929, 2862, 2228, 1953, 1728, 1650, 1596, 1496, 1390, 1299, 1170, 1117, 1021, 941, 843, 699, 665.

(Z)-2-(3,3-Difluoro-2-phenylallyl)-N-methyl-N-phenyloctadec-9-enamide (3-3ha)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 54 mg product was obtained by 68% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 – 7.12 (m, 6H), 7.01 (d, J = 7.8, 2H), 6.83 – 6.77 (m, 2H), 5.31 – 5.22 (m, 2H), 3.13 (s, 3H), 2.62 – 2.54 (m, 1H), 2.48 – 2.42 (m, 1H), 2.34 – 2.27 (m, 1H), 1.95 – 1.87 (m, 4H), 1.54 – 1.48 (m, 1H), 1.26 – 1.09 (m, 18H), 1.01 – 0.93 (m, 3H), 0.81 (t, J = 6.6 Hz, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.0, 154.0 (dd, J = 291.6, 288.6 Hz), 143.6, 133.1 (t, J = 4.1 Hz), 130.0, 129.8, 129.5, 128.3, 128.1 (t, J = 3.5 Hz), 127.5, 127.4, 127.1, 90.4 (dd, J = 21.6, 13.7 Hz), 39.9 (t, J = 2.9 Hz), 37.4, 31.9, 30.2, 29.8, 29.7, 29.5, 29.4, 29.34, 29.32, 29.1, 27.24, 27.19, 27.17, 22.7, 14.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.19 (d, J = 39.0 Hz, 1F), -90.10 (d, J = 39.0 Hz, 1F). HRMS (ESI-MS) Calcd. For C₃₄H₄₇F₂NONa [M+Na]⁺ 546.3518, found: 546.3510.

IR (neat, cm⁻¹): v: 3458, 3004, 2924, 2854, 2322, 2092, 1991, 1657, 1595, 1451, 1387, 1300, 1236, 1119, 1023, 956, 844, 764, 698.

$$F_2C$$
 Ph

5,5-Difluoro-2-(4-hydroxybenzyl)-N-methyl-N,4-diphenylpent-4-enamide (3-3ia)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (2:1). 25 mg product was obtained by 41% isolated yield as white solid. Single crystals were obtained via slow evaporation of CH_2Cl_2 . Melting point: 165.9 - 167.2 °C.

¹H NMR (400 MHz, Chloroform-*d*) δ 7.20 – 6.91 (m, 8H), 6.69 – 6.55 (m, AA'BB' 4 spin system, AA' and BB' parts, 4H), 6.31 (broad s, 2H), 3.02 (s, 3H), 2.76 (dd, J = 12.8, 8.0 Hz, 1H), 2.67 – 2.60 (m, 1H), 2.57 – 2.38 (m, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 174.5, 154.7, 154.1 (dd, J = 292.8, 292.7 Hz), 143.0, 132.8 (t, J = 4.0 Hz), 131.0, 130.1, 129.3, 128.4, 128.0 (t, J = 3.3 Hz), 127.6, 127.2, 127.1, 115.2, 90.2 (dd, J = 21.4, 13.7 Hz), 42.9 (t, J = 2.5 Hz), 37.4, 37.3, 29.9.

¹⁹F NMR (565 MHz, Chloroform-d) δ -88.80 (d, J = 38.4 Hz, 1F), -89.72 (d, J = 38.4 Hz, 1F). HRMS (EI-MS) Calcd. For C₂₅H₂₃F₂NO₂ 407.1691, found: 407.1687.

IR (neat, cm⁻¹): \tilde{v} : 3160, 2924, 2856, 2690, 2317, 2107, 1994, 1887, 1734, 1588, 1510, 1449, 1376, 1233, 1120, 1023, 940, 831, 758, 696.

2-(3,3-Difluoro-2-phenylallyl)-N-methyl-N-phenylhex-5-ynamide (3-3ja)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 34 mg product was obtained by 64% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.19 – 7.10 (m, 6H), 6.99 – 6.85 (m, 4H), 3.13(s, 3H), 2.60 – 2.50 (m, 2H), 2.48 – 2.44 (m, 1H), 2.09 – 2.04 (m, 1H), 1.99 – 1.89 (m, 1H), 1.86 – 1.77 (m, 1H), 1.63 – 1.60 (m, 1H), 1.46 – 1.41 (m, 1H).

¹³C NMR (151 MHz, Chloroform-d) δ 174.1, 154.1 (dd, J = 291.3, 288.3 Hz), 143.2, 132.7 (t, J = 3.8 Hz), 129.6, 128.3, 128.1 (t, J = 3.2 Hz), 127.6, 127.4, 127.2, 90.1 (dd, J = 21.3, 14.3 Hz), 83.4, 68.8, 38.9 (t, J = 2.4 Hz), 37.6, 30.1, 30.0, 16.3.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.24 (d, J = 39.0 Hz, 1F), -89.74 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₂H₂₁F₂NONa [M+Na]⁺ 376.1483, found: 376.1479.

IR (neat, cm⁻¹): v: 3478, 3301, 3060, 2930, 2859, 2324, 2114, 1991, 1895, 1726, 1650, 1495, 1444, 1390, 1343, 1235, 1119, 1030, 955, 766, 698.

2-(Cyclohexylmethyl)-5,5-difluoro-N-methyl-4-phenyl-N-(p-tolyl)pent-4-enamide (3-3ka)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 38 mg product was obtained by 61% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.18 – 7.12 (m, 3H), 7.00 – 6.95 (m, 2H), 6.93 (d, J = 7.8 Hz, 2H), 6.67 (d, J = 7.4 Hz, 2H), 3.10 (s, 3H), 2.56 – 2.35 (m, 3H), 2.25 (s, 3H), 1.54 – 1.41 (m, 4H), 1.34 (d, J = 13.2 Hz, 1H), 1.09 – 0.92 (m, 6H), 0.62 – 0.56 (m, 1H), 0.45 – 0.39 (m, 1H).

¹³C NMR (151 MHz, Chloroform-d) δ 175.5, 154.0 (dd, J = 290.7, 288.3 Hz), 140.9, 137.4, 133.0 (t, J = 3.6 Hz), 130.0, 128.3 (t, J = 3.0 Hz), 128.2, 127.1, 127.0, 90.6 (dd, J = 21.3, 14.0 Hz), 39.4, 37.5, 37.3 (t, J = 2.6 Hz), 35.4, 33.6, 33.1, 30.5, 26.5, 26.2, 26.1, 21.0.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.83 (d, J = 39.0 Hz, 1F), -90.45 (d, J = 40.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NONa [M+Na]⁺ 434.2266, found: 434.2261.

IR (neat, cm⁻¹): v: 3480, 3031, 2922, 2851, 2664, 2324, 2075, 2007, 1902, 1728, 1654, 1512, 1445, 1386, 1235, 1117, 1012, 941, 825, 759, 697.

2-(Cyclohexylmethyl)-5,5-difluoro-*N*-(4-methoxyphenyl)-*N*-methyl-4-phenylpent4-enamide (3-3la)

Following procedure A, the crude mixture was purified by silica gel column chromatography with

pentane/EA (10:1). 26 mg product was obtained by 40% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.21 – 7.13 (m, 3H), 6.99 (d, J = 7.8 Hz, 2H), 6.74 – 6.60 (m, 4H), 3.71 (s, 3H), 3.09 (s, 3H), 2.55 – 2.50 (m, 1H), 2.46 – 2.42 (m, 1H), 2.40 – 2.36 (m, 1H), 1.52 – 1.46 (m, 4H), 1.37 – 1.30 (m, 1H), 1.08 – 0.91 (m, 6H), 0.62 – 0.56 (m, 1H), 0.49 – 0.40 (m, 1H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 175.7, 158.7, 154.0 (dd, J = 290.8, 288.6 Hz), 136.3, 133.1 (t, J = 3.8 Hz), 128.29, 128.25, 127.1, 114.5, 90.6 (dd, J = 21.4, 14.0 Hz), 55.5, 39.5, 37.6, 37.2 (t, J = 2.6 Hz), 35.4, 33.6, 33.1, 30.5, 26.5, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.70 (d, J = 40.1 Hz, 1F), -90.39 (d, J = 40.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NO₂Na [M+Na]⁺ 450.2215, found: 450.2202.

IR (neat, cm⁻¹): \tilde{v} : 3465, 2922, 2850, 2324, 2083, 1886, 1728, 1651, 1509, 1387, 1343, 1296, 1241, 1169, 1116, 1034, 941, 837, 760, 698.

2-(Cyclohexylmethyl)-5,5-difluoro-N-(4-fluorophenyl)-N-methyl-4-phenylpent-4-enamide (3-3ma)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 24 mg product was obtained by 38% isolated yield as colorless liquid.

¹H NMR (400 MHz, Chloroform-*d*) δ 7.25 – 7.16 (m, 3H), 7.06 – 6.98 (m, 2H), 6.79 (t, *J* = 8.4 Hz, 2H), 6.70 (t, *J* = 6.8 Hz, 2H), 3.09 (s, 3H), 2.57 – 2.50 (m, 1H), 2.48 – 2.42 (m, 1H), 2.38 – 2.28 (m, 1H), 1.55 – 1.43 (m, 4H), 1.33 – 1.26 (m, 1H), 1.12 – 0.94 (m, 6H), 0.63 – 0.41 (m, 2H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 175.4, 161.5 (dd, J = 248.6 Hz), 154.0 (dd, J = 291.6, 289.2 Hz), 139.5 (d, J = 3.2 Hz), 133.0 (t, J = 3.7 Hz), 128.9 (d, J = 8.7 Hz), 128.4, 128.2 (t, J = 3.2 Hz), 127.3, 116.3 (d, J = 22.7 Hz), 90.5 (dd, J = 32.2, 14.2 Hz), 39.6, 37.6, 37.3 (t, J = 2.5 Hz), 35.3, 33.4, 33.2, 30.4 (d, J = 1.8 Hz), 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.62 (d, J = 39.6 Hz, 1F), -90.21 (d, J = 39.0 Hz, 1F), -113.74 (tt, J = 8.8, 5.0 Hz, 1F).

HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₃NONa [M+Na]⁺ 438.2015, found: 438.2012.

IR (neat, cm⁻¹): v: 3483, 3059, 2923, 2851, 2329, 1893, 1729, 1655, 1506, 1446, 1387, 1302, 1231, 1118, 1011, 941, 843, 760, 689.

N-(4-Chlorophenyl)-2-(cyclohexylmethyl)-5,5-difluoro-*N*-methyl-4-phenylpent-4-enamide (3-3na)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 26 mg product was obtained by 40% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.23 – 7.17 (m, 3H), 7.08 (d, J = 8.4 Hz, 2H), 7.01 – 6.96 (m, 2H), 6.67 (d, J = 7.8 Hz, 2H), 3.09 (s, 3H), 2.53 – 2.49 (m, 1H), 2.47 – 2.43 (m, 1H), 2.35 – 2.31 (m, 1H), 1.54 – 1.44 (m, 4H), 1.35 – 1.29 (m, 1H), 1.11 – 0.94 (m, 6H), 0.58 (qd, J = 12.3, 2.9 Hz,

1H), 0.48 (qd, J = 12.2, 3.7 Hz, 1H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 175.2, 154.0 (dd, J = 291.5, 289.5 Hz), 142.0, 133.3, 132.9 (t, J = 4.0 Hz), 129.6, 128.6, 128.4, 128.2 (t, J = 3.2 Hz), 127.4, 90.4 (dd, J = 21.3, 14.3 Hz), 39.5, 37.5, 37.4 (t, J = 2.5 Hz), 35.4, 33.5, 33.2, 30.4 (d, J = 1.8 Hz), 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.74 (d, J = 39.6 Hz, 1F), -90.18 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₂NOClNa [M+Na]⁺ 454.1720, found: 454.1716.

IR (neat, cm⁻¹): v: 3482, 3059, 2922, 2851, 2664, 2325, 2060, 1901, 1729, 1656, 1490, 1339, 1303, 1278, 1164, 1095, 1012, 942, 837, 759, 720, 698.

$$F_3C$$
 F_2C
 Ph

2-(Cyclohexylmethyl)-5,5-difluoro-*N*-methyl-4-phenyl-*N*-(4-(trifluoromethyl)phenyl)pent-4-enamide (3-30a)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 23 mg product was obtained by 32% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.35 (d, J = 7.8 Hz, 2H), 7.21 – 7.16 (m, 3H), 7.02 – 6.96 (m, 2H), 6.84 (d, J = 7.8 Hz, 2H), 3.13 (s, 3H), 2.57 – 2.43 (m, 2H), 2.34 – 2.27 (m, 1H), 1.53 – 1.43 (m, 4H), 1.29 (d, J = 12.6 Hz, 1H), 1.13 – 0.95 (m, 6H), 0.60 – 0.44 (m, 2H).

¹³C NMR (151 MHz, Chloroform-d) δ 175.1, 154.0 (dd, J = 290.5, 288.7 Hz), 146.6, 132.9 (dd, J = 3.2, 2.1 Hz), 129.6 (q, J = 32.9 Hz), 128.5, 128.2 (t, J = 3.3 Hz), 127.6, 127.5, 126.6 (q, J = 3.9 Hz), 123.6 (q, J = 272.4 Hz), 90.3 (dd, J = 21.7, 15.1 Hz), 39.7, 37.4 (t, J = 2.7 Hz), 37.3, 35.4, 33.4, 33.2, 30.5, 26.4, 26.14, 26.10.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -62.63 (s, 3F), -89.82 (d, J = 40.1 Hz, 1F), -90.11 (d, J = 39.6 Hz, 1F).

HRMS (ESI-MS) Calcd. For C₂₆H₂₈F₅NONa [M+Na]⁺ 488.1983, found: 488.1972.

IR (neat, cm⁻¹): v: 3061, 2924, 2852, 2658, 2323, 2094, 2011, 1949, 1731, 1661, 1612, 1447, 1389, 1324, 1239, 1123, 1068, 1013, 942, 850, 759, 728, 700.

$$F_2C$$
 Ph

5,5-Difluoro-*N*-methyl-*N*,4-diphenylpent-4-enamide (3-3pa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 10 mg product was obtained by 22% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.26 (t, J = 7.3 Hz, 2H), 7.24 – 7.17 (m, 3H), 7.15 (t, J = 7.2 Hz, 1H), 7.09 (d, J = 7.7 Hz, 2H), 6.95 (d, J = 7.5 Hz, 2H), 3.15 (s, 3H), 2.62 (t, J = 7.9 Hz, 2H), 2.05 (t, J = 7.9 Hz, 2H).

 13 C NMR (151 MHz, Chloroform-d) δ 171.8, 153.5 (dd, J = 290.8, 287.8 Hz), 143.8, 132.9 (t, J = 3.9 Hz), 129.7, 128.4, 128.1 (t, J = 3.6 Hz), 127.8, 127.3, 127.2, 91.4 (dd, J = 21.3, 13.9 Hz), 37.3, 32.3 (t, J = 2.9 Hz), 23.7.

¹⁹F NMR (376 MHz, Chloroform-d) δ -90.25 (d, J = 41.0 Hz, 1F), -90.60 (d, J = 41.0 Hz, 1F).

HRMS (ESI-MS) Calcd. For C₁₈H₁₇F₂NONa [M+Na]⁺ 324.1170, found: 324.1170.

IR (neat, cm $^{-1}$): \tilde{v} : 3482, 3059, 2927, 2323, 1952, 1729, 1655, 1495, 1384, 1233, 1118, 1022, 917, 764, 698.

5,5-Difluoro-*N*-methyl-2-((1-methyl-1*H*-indol-3-yl)methyl)-*N*,4-diphenylpent-4-enamide (3-3ra)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (4:1). 26 mg product was obtained by 39% isolated yield as white solid. Melting point: 127.3 -128.4 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.15 – 7.02 (m, 6H), 6.90 (d, J = 8.0 Hz, 2H), 6.85 (t, J = 7.7 Hz, 2H), 6.71 (t, J = 7.4 Hz, 1H), 6.61 (s, 1H), 6.34 (d, J = 7.9 Hz, 1H), 6.18 (broad s, 2H), 3.60 (s, 3H), 3.00 (s, 3H), 2.92 (dd, J = 13.9, 8.7 Hz, 1H), 2.78 (quint, J = 7.2 Hz, 1H), 2.69 – 2.61 (m, 2H), 2.50 (broad dd, J = 14.7, 7.7 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 174.8, 154.0 (dd, J = 291.7, 288.3 Hz), 143.2, 137.0, 132.7 (t, J = 4.1 Hz), 129.2, 128.4, 128.1 (t, J = 3.5 Hz), 127.7, 127.4, 127.18, 127.16, 127.1, 121.4, 119.0, 118.5, 111.8, 108.9, 90.4 (dd, J = 21.0, 13.0 Hz), 41.0, 37.3, 32.5, 29.5, 27.7.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.90 (d, J = 39.0 Hz, 1F), -89.95 (d, J = 39.0 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₈H₂₆F₂N₂ONa [M+Na]⁺ 467.1905, found: 467.1903.

IR (neat, cm⁻¹): v: 3379, 3059, 2923, 2853, 2320, 2115, 1983, 1890, 1726, 1652, 1473, 1444, 1377, 1325, 1231, 1119, 1012, 933, 868, 738, 699.

$$F_2C$$

2-(3,3-Difluoro-2-phenylallyl)-N-methyl-N-(pyridin-2-yl)hexanamide (3-3sa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (3:1). 22 mg product was obtained by 41% isolated yield as yellow liquid.

¹H NMR (600 MHz, Chloroform-d) δ 8.28 (dd, J = 5.4, 1.8 Hz, 1H), 7.45 (t, J = 7.9 Hz, 1H), 7.23 – 7.04 (m, 5H), 7.02 (dd, J = 7.5, 4.9 Hz, 1H), 6.71 (broad s, 1H), 3.17 (s, 3H), 2.70 (d, J = 11.1 Hz, 1H), 2.50 (dd, J = 13.4, 7.2 Hz, 1H), 2.39 (broad s, 1H), 1.65 – 1.56 (m, 1H), 1.28 (broad s, 1H), 1.15 – 0.92 (m, 4H), 0.72 (t, J = 7.2 Hz, 3H).

 13 C NMR (151 MHz, Chloroform-*d*) δ 175.5, 156.1, 154.0 (dd, J = 290.8, 288.3 Hz), 149.1, 138.0, 133.2 (t, J = 3.9 Hz), 128.3, 128.2 (t, J = 3.5 Hz), 127.2, 121.9, 120.9, 90.5 (dd, J = 21.7, 14.0 Hz), 40.8, 35.5, 31.7, 30.5, 29.3, 22.6, 13.8.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ -89.69 (d, J = 39.6 Hz, 1F), -90.33 (d, J = 39.9Hz, 1F). HRMS (EI-MS): Calcd. For C₂₁H₂₃F₂N₂O [M-H]⁺ 357.1773, found 357.1772.

 $IR \ (neat,\,cm^{-1}): \ \tilde{v}: \ 3304, \ 3060, \ 2929, \ 2862, \ 2322, \ 2113, \ 1986, \ 1727, \ 1654, \ 1583, \ 1465, \ 1339, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12300, \ 12$

1127, 1011, 937, 847, 792, 754.

$$F_2C$$

4-(3-Chlorophenyl)-2-(cyclohexylmethyl)-5,5-difluoro-*N*-methyl-*N*-phenylpent-4-enamide (3-3fb)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 36 mg product was obtained by 55% isolated yield as white solid. Melting point: 60.7 - 61.8 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 – 7.12 (m, 4H), 7.10 (t, J = 7.7 Hz, 1H), 7.04 (s, 1H), 6.89 (d, J = 7.8 Hz, 1H), 6.80 (d, J = 6.6 Hz, 2H), 3.13 (s, 3H), 2.57 – 2.51 (m, 1H), 2.46 – 2.40 (m, 1H), 2.38 – 2.30 (m, 1H), 1.53 – 1.44 (m, 4H), 1.29 (d, J = 11.4 Hz, 1H), 1.10 – 0.93 (m, 6H), 0.56 (qd, J = 12.5, 3.1 Hz, 1H), 0.47 (qd, J = 12.0, 3.6 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.2, 154.1 (dd, J = 291.7, 289.3 Hz), 143.4, 135.0 (t, J = 3.8 Hz), 134.2, 129.6, 129.5, 128.3 (t, J = 3.2 Hz), 127.7, 127.4, 127.2, 126.4 (t, J = 3.2 Hz), 89.9 (dd, J = 22.5, 13.9 Hz), 39.7, 37.5, 37.3 (t, J = 2.9 Hz), 35.3, 33.4, 33.2, 30.3, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.38 (d, J = 36.2 Hz, 1F), -88.90 (d, J = 37.3Hz, 1F).

HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₂NOClNa [M+Na]⁺ 454.1720, found: 454.1726.

IR (neat, cm⁻¹): v: 3062, 2922, 2853, 2314, 2102, 1966, 1733, 1640, 1594, 1448, 1342, 1239, 1122, 1025, 896, 775, 700.

2-(Cyclohexylmethyl)-5,5-difluoro-*N*-methyl-*N*-phenyl-4-(*m*-tolyl)pent-4-enamide (3-3fc)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 37 mg product was obtained by 60% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.15 – 7.09 (m, 3H), 7.07 (t, J = 7.8 Hz, 1H), 6.97 (d, J = 7.8 Hz, 1H), 6.84 (s, 1H), 6.81 (d, J = 7.8 Hz, 1H), 6.75 (d, J = 7.2 Hz, 2H), 3.12 (s, 3H), 2.59 – 2.53 (m, 1H), 2.46 – 2.35 (m, 2H), 2.22 (s, 3H), 1.53 – 1.41 (m, 4H), 1.27 (d, J = 13.0 Hz, 1H), 1.12 – 0.95 (m, 6H), 0.58 – 0.41 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.5, 153.9 (dd, J = 289.8, 288.6 Hz), 143.5, 137.8, 133.1 (t, J = 3.6 Hz), 129.3, 128.9 (t, J = 3.2 Hz), 128.3, 128.0, 127.5, 127.2, 125.3 (t, J = 3.2Hz), 90.6 (dd, J = 21.1, 14.3 Hz), 39.7, 37.5, 37.3 (t, J = 2.7 Hz), 35.3, 33.3, 33.2, 30.5, 26.5, 26.21, 26.16, 21.4.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.95 (d, J = 40.7 Hz, 1F), -90.37 (d, J = 40.1 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NONa [M+Na]⁺ 434.2266, found: 434.2262. IR (neat, cm⁻¹): \tilde{v} : 3481, 2922, 2851, 2322, 2114, 1883, 1728, 1654, 1595, 1494, 1388, 1242, 1118, 1027, 959, 838, 782, 700.

2-(Cyclohexylmethyl)-5,5-difluoro-4-(3-methoxyphenyl)-*N*-methyl-*N*-phenylpent4-enamide (3-3fd)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 41 mg product was obtained by 64% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.16 – 7.05 (m, 4H), 6.80 – 6.70 (m, 3H), 6.63 – 6.57 (m, 2H), 3.70 (s 3H), 3.13 (s, 3H), 2.59 – 2.52 (m, 1H), 2.47 – 2.36 (m, 2H), 1.53 – 1.43 (m, 4H), 1.30 (d, J = 13.2 Hz, 1H), 1.11 – 0.95 (m, 6H), 0.59 – 0.42 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.5, 159.5, 154.0 (dd, J = 291.3, 288.7 Hz), 143.5, 134.5 (t, J = 3.5 Hz), 129.4, 129.3, 127.5, 127.2, 120.7 (t, J = 3.0 Hz), 114.0 (t, J = 3.3 Hz), 112.9, 90.6 (dd, J = 21.3, 14.0 Hz), 55.2, 39.8, 37.5, 37.4 (t, J = 2.4 Hz), 35.3, 33.4, 33.2, 30.6, 26.4, 26.2, 26.1. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.44 (d, J = 39.6 Hz, 1F), -89.63 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NO₂Na [M+Na]⁺ 450.2215, found: 450.2219.

IR (neat, cm $^{-1}$): \tilde{v} : 3062, 2922, 2850, 2323, 2082, 1950, 1729, 1654, 1594, 1493, 1388, 1284, 1117, 1019, 960, 836, 774, 699.

$$F_2C$$
 SMe

2-(Cyclohexylmethyl)-5,5-difluoro-*N*-methyl-4-(3-(methylthio)phenyl)-*N*-phenylpent-4-enamide (3-3fe)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 35 mg product was obtained by 52% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.17 – 7.05 (m, 5H), 6.95 (s, 1H), 6.80 – 6.72 (m, 3H), 3.13 (s, 3H), 2.59 – 2.52 (m, 1H), 2.46 – 2.34 (m, 2H), 2.38 (s, 3H), 1.53 – 1.44 (m, 4H), 1.28 (d, *J* = 13.3 Hz, 1H), 1.11 – 0.95 (m, 6H), 0.59 – 0.43 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.4, 154.0 (dd, J = 290.7, 289.0 Hz), 143.4, 138.7, 133.9 (t, J = 3.9 Hz), 129.4, 128.7, 127.6, 127.2, 126.4 (t, J = 3.3 Hz), 125.6, 125.0 (t, J = 3.2 Hz), 90.4 (dd, J = 21.7, 13.9 Hz), 39.9, 37.5, 37.3 (t, J = 3.0 Hz), 35.3, 33.32, 33.29, 30.5, 26.4, 26.2, 26.1,

15.8.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.14 (d, J = 38.4 Hz, 1F), -89.50 (d, J = 39.0 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NOSNa [M+Na]⁺ 466.1987, found: 466.1991.

IR (neat, cm⁻¹): v: 3061, 2922, 2850, 2325, 2085, 1952, 1728, 1653, 1593, 1494, 1444, 1340, 1236, 1119, 1020, 955, 886, 775, 699.

2-(Cyclohexylmethyl)-5,5-difluoro-4-(2-fluorophenyl)-*N*-methyl-*N*-phenylpent-4-enamide (3-3ff)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (20:1). 19 mg product was obtained by 30% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 − 7.14 (m, 1H), 7.14 − 7.09 (m, 3H), 6.98 − 6.89 (m, 3H), 6.86 − 6.81 (m, 2H), 3.13 (s, 3H), 2.49 (dt, J = 6.6, 2.4 Hz, 2H), 2.30 − 2.24 (m, 1H), 1.52 − 1.43 (m, 4H), 1.33 (d, J = 13.0 Hz, 1H), 1.09 − 0.93 (m, 6H), 0.59 (qd, J = 12.5, 3.0 Hz, 1H), 0.43 (≈qd, J = 12.9, 4.1 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.3, 160.0 (d, J = 248.4 Hz), 153.8 (t, J = 289.5 Hz), 143.5, 130.8 (≈q, J = 2.4 Hz), 129.42, 129.36, 127.5, 127.1, 124.0 (d, J = 3.6 Hz), 120.7 (ddd, J = 15.3, 4.8, 2.1 Hz), 115.8 (d, J = 22.3 Hz), 85.2 (dd, J = 25.0, 16.8 Hz), 39.3, 37.41, 37.39 (t, J = 2.6 Hz), 35.4, 33.6, 32.8, 30.7, 26.4, 26.3, 26.2.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -87.01 (dd, J = 35.8, 13.1 Hz, 1F), -89.94 (d, J = 35.6 Hz, 1F), -113.30 to -113.39 (m, 1F).

HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₃NONa [M+Na]⁺ 438.2015, found: 438.1999.

IR (neat, cm⁻¹): v: 3484, 3063, 2922, 2851, 2326, 2102, 1737, 1654, 1594, 1493, 1388, 1248, 1118, 1010, 944, 760, 699.

$$F_2C$$

2-(Cyclohexylmethyl)-5,5-difluoro-4-(4-fluorophenyl)-N-methyl-N-phenylpent-4- enamide (3-3fg)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 40 mg product was obtained by 64% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 – 7.16 (m, 3H), 6.98 – 6.92 (m, 2H), 6.88 – 6.79 (m, 4H), 3.13 (s, 3H), 2.53 – 2.47 (m, 1H), 2.46 – 2.41 (m, 1H), 2.37 – 2.32 (m, 1H), 1.54 – 1.45 (m, 4H), 1.32 (d, J = 13.2 Hz, 1H), 1.07 – 0.94 (m, 6H), 0.58 (qd, J = 12.6, 3.0 Hz, 1H), 0.43 (qd, J = 12.5, 3.5 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.3, 161.8 (d, J = 246.6 Hz), 154.0 (t, J = 289.5 Hz), 143.5, 129.9 (dt, J = 8.3, 3.2 Hz), 129.5, 128.9 (≈q, J = 3.6 Hz), 127.7, 127.3, 115.3 (d, J = 21.7 Hz), 89.7 (dd, J = 22.3, 14.0 Hz), 39.4, 37.5, 37.3 (t, J = 2.4 Hz), 35.3, 33.5, 33.1, 30.6, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.64 (d, J = 40.1 Hz, 1F), -90.34 (dd, J = 40.1, 3.4 Hz, 1F), -114.69 to -114.76 (m, 1F).

HRMS (ESI-MS) Calcd. For C₂₅H₂₉F₃NO [M+H]⁺ 416.2196, found: 416.2198.

IR (neat, cm⁻¹): v: 3483, 3061, 2922, 2851, 2327, 1997, 1892, 1728, 1653, 1509, 1389, 1236, 1118, 1010, 890, 837, 771, 700, 656.

4-(4-Chlorophenyl)-2-(cyclohexylmethyl)-5,5-difluoro-*N*-methyl-*N*-phenylpent-4-enamide (3-3fh)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 32 mg product was obtained by 49% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.22 – 7.16 (m, 3H), 7.14 (d, J = 8.2 Hz, 2H), 6.92 (d, J = 8.4 Hz, 2H), 6.80 (d, J = 7.2 Hz, 2H), 3.13 (s, 3H), 2.53 – 2.47 (m, 1H), 2.45 – 2.40 (m, 1H), 2.38 – 2.32 (m, 1H), 1.53 – 1.46 (m, 4H), 1.32 (d, J = 13.2 Hz, 1H), 1.08 – 0.95 (m, 6H), 0.58 (qd, J = 12.5, 3.1 Hz, 1H), 0.45 (qd, J = 12.0, 3.7 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.2, 154.0 (dd, J = 291.3, 289.0 Hz), 143.5, 133.0, 131.6 (t, J = 3.6 Hz), 129.50, 129.49 (t, J = 3.3 Hz), 128.5, 127.7, 127.2, 89.8 (dd, J = 22.0, 13.6 Hz), 39.6, 37.5, 37.3 (t, J = 2.7 Hz), 35.3, 33.5, 33.2, 30.4, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.72 (d, J = 37.9 Hz, 1F), -89.40 (dd, J = 38.4, 2.8 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₂NOClNa [M+Na]⁺ 454.1720, found: 454.1697.

IR (neat, cm⁻¹): v: 3062, 2922, 2851, 2323, 2107, 1900, 1726, 1654, 1594, 1494, 1341, 1240, 1094, 941, 829, 771, 700.

4-(4-Bromophenyl)-2-(cyclohexylmethyl)-5,5-difluoro-N-methyl-N-phenylpent-4- enamide (3-3fi)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 40 mg product was obtained by 55% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7. 29 (d, J = 9.0 Hz, 2H), 7.22 – 7.13 (m, 3H), 6.86 (d, J = 8.4 Hz, 2H), 6.79 (d, J = 7.2 Hz, 2H), 3.12 (s, 3H), 2.52 – 2.47 (m, 1H), 2.44 – 2.40 (m, 1H), 2.38 – 2.31 (m, 1H), 1.53 – 1.46 (m, 4H), 1.31 (d, J = 12.7 Hz, 1H), 1.09 – 0.95 (m, 6H), 0.57 (qd, J =

12.6, 3.0 Hz, 1H), 0.45 (qd, J = 11.9, 3.5 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.1, 153.9 (dd, J = 291.6, 289.2 Hz), 143.4, 132.1 (t, J = 3.6 Hz), 131.5, 129.8 (t, J = 3.5 Hz), 129.5, 127.7, 127.2, 121.1, 89.8 (dd, J = 22.3, 13.9 Hz), 39.6, 37.5, 37.3 (t, J = 2.6 Hz), 35.3, 33.5, 33.2, 30.3, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.60 (d, J = 38.4 Hz, 1F), -89.28 (dd, J = 38.4 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₅H₂₈F₂NOBrNa [M+Na]⁺ 498.1215, found: 498.1207.

IR (neat, cm⁻¹): v: 3484, 3062, 2922, 2851, 2329, 1983, 1725, 1653, 1492, 1389, 1239, 1118, 1006, 940, 827, 700, 659.

2-(Cyclohexylmethyl)-5,5-difluoro-*N*-methyl-*N*-phenyl-4-(*p*-tolyl)pent-4-enamide (3-3fj)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 44 mg product was obtained by 71% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.18 – 7.09 (m, 3H), 6.98 (d, J = 7.8 Hz, 2H), 6.89 (d, J = 7.8 Hz, 2H), 6.77 (d, J = 7.2 Hz, 2H), 3.12 (s, 3H), 2.56 – 2.51 (m, 1H), 2.45 – 2.35 (m, 2H), 2.27 (s, 3H), 1.54 – 1.42 (m, 4H), 1.30 (d, J = 13.2 Hz, 1H), 1.10 – 0.93 (m, 6H), 0.55 (qd, J = 12.2, 3.4 Hz, 1H), 0.44 (qd, J = 12.1, 3.7 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-d) δ 175.5, 153.9 (dd, J = 290.4, 287.8 Hz), 143.5, 136.8, 130.1 (t, J = 3.9 Hz), 129.4, 129.0, 128.0 (t, J = 3.0 Hz), 127.4, 127.3, 90.4 (dd, J = 21.1, 14.0 Hz), 39.7, 37.5, 37.4 (t, J = 2.4 Hz), 35.3, 33.4, 33.1, 30.5, 26.5, 26.2, 26.1, 21.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -90.08 (d, J = 40.7 Hz, 1F), -90.71 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NONa [M+Na]⁺ 434.2266, found: 434.2260.

IR (neat, cm⁻¹): v: 3480, 3032, 2922, 2851, 2326, 2114, 1902, 1726, 1654, 1446, 1340, 1235, 1113, 1008, 941, 819, 770, 700.

$$F_2C$$
 OM6

$\hbox{$2$-(Cyclohexylmethyl)-5,5$-difluoro-4-(4-methoxyphenyl)-N-methyl-N-phenylpent4-enamide (3-3fk)}$

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (10:1). 33 mg product was obtained by 51% isolatedyield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 – 7.13 (m, 3H), 6.91 (d, J = 9.0 Hz, 2H), 6.83 – 6.78 (m, 2H), 6.71 (d, J = 8.7 Hz, 2H), 3.74 (s, 3H), 3.13 (s, 3H), 2.54 – 2.48 (m, 1H), 2.45 – 2.36 (m, 2H), 1.54 – 1.44 (m, 4H), 1.31 (d, J = 12.8 Hz, 1H), 1.09 – 0.95 (m, 6H), 0.56 (qd, J = 12.5, 3.1 Hz, 1H), 0.44 (qd, J = 11.9, 3.7 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.5, 158.6, 153.9 (dd, J = 289.5, 287.2, Hz), 143.6, 129.4, 129.3 (t, J = 3.0 Hz), 127.5, 127.3, 125.2 (t, J = 3.6 Hz), 113.8, 90.0 (dd, J = 21.3, 13.9 Hz), 55.3, 39.6, 37.5, 37.4 (t, J = 2.7 Hz), 35.3, 33.5, 33.1, 30.6, 26.4, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -90.58 (d, J = 42.4 Hz, 1F), -91.23 (d, J = 42.4 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₆H₃₁F₂NO₂Na [M+Na]⁺ 450.2215, found: 450.2205.

IR (neat, cm⁻¹): v: 3479, 3039, 2922, 2850, 2240, 1885, 1727, 1653, 1598, 1448, 1389, 1240, 1113, 1029, 941, 832, 700.

$$F_2C$$
 t -Bu

4-(4-(*tert*-Butyl)phenyl)-2-(cyclohexylmethyl)-5,5-difluoro-*N*-methyl-*N*-phenylpent-4-enamide (3-3fl)

Following procedure B, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 47 mg product was obtained by 69% isolated yield as colorless liquid.

 1 H NMR (600 MHz, Chloroform-d) δ 7.19 (d, J = 8.4 Hz, 2H), 7.14 – 7.09 (m, 3H), 6.93 (d, J = 8.4 Hz, 2H), 6.79 – 6.71 (m, 2H), 3.12 (s, 3H), 2.58 – 2.53 (m, 1H), 2.46 – 2.41 (m, 1H), 2.40 – 2.33 (m, 1H), 1.52 – 1.39 (m, 4H), 1.25 (s, 9H), 1.21 – 1.17 (m, 1H), 1.10 – 0.93 (m, 6H), 0.53 (qd, J = 11.6, 3.2 Hz, 1H), 0.41 (qd, J = 13.3, 3.7 Hz, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 175.5, 154.0 (dd, J = 290.2, 288.0 Hz), 150.1, 143.6, 129.9 (t, J = 3.9 Hz), 129.4, 127.9 (t, J = 3.0 Hz), 127.4, 127.3, 125.2, 90.3 (dd, J = 21.3, 14.3 Hz), 39.6, 37.5, 37.3 (t, J = 2.6 Hz), 35.3, 34.5, 33.4, 33.1, 31.3, 30.3, 26.5, 26.2, 26.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -90.00 (d, J = 40.7 Hz, 1F), -90.72 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₂₉H₃₇F₂NONa [M+Na]⁺ 476.2735, found: 476.2729.

IR (neat, cm⁻¹): v: 3482, 3037, 2923, 2853, 2664, 2066, 1988, 1728, 1656, 1595, 1448, 1340, 1236, 1107, 942, 834, 771, 700.

8,8-Difluoro-5-(methyl(phenyl)carbamoyl)-7-phenyloct-7-en-1-yl 5-(2,5-dimethylphenoxy)-2,2-dimethylpentanoate (3-3ta)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (15:1). 55 mg product was obtained by 60% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-d) δ 7.18 – 7.11 (m, 6H), 6.99 (d, J = 7.8 Hz, 2H), 6.92 (d, J = 7.8 Hz, 1H), 6.83 – 6.77 (m, 2H), 6.58 (d, J = 7.2 Hz, 1H), 6.53 (s, 1H), 3.87 (t, J = 6.6 Hz, 2H), 3.83 (t, J = 5.4 Hz, 2H), 3.12 (s, 3H), 2.61 – 2.55 (m, 1H), 2.45 (ddd, J = 14.5, 8.0, 2.4 Hz, 1H), 2.34 – 2.28 (m, 1H), 2.23 (s, 3H), 2.09 (s, 3H), 1.63 – 1.51 (m, 4H), 1.34 – 1.16 (m, 5H), 1.12 (s, 6H), 1.04

-0.99 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 177.8, 174.7, 157.0, 154.0 (dd, J = 291.6, 288.3 Hz), 143.5, 136.5, 132.9 (t, J = 4.1 Hz), 130.3, 129.6, 128.4, 128.0 (t, J = 3.5 Hz), 127.6, 127.3, 127.2, 123.6, 120.7, 112.0, 90.4 (dd, J = 21.4, 13.7 Hz), 67.9, 64.1, 42.1, 39.9 (t, J = 2.4 Hz), 37.5, 37.1, 31.5, 30.2, 28.6, 25.2, 23.6, 21.4, 15.8.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.04 (d, J = 39.0 Hz, 1F), -89.86 (d, J = 38.7 Hz, 1F). HRMS (ESI-MS) Calcd. For C₃₇H₄₅F₂NO₄Na [M+Na]⁺ 628.3209, found: 628.3196.

IR (neat, cm⁻¹): v: 3500, 2929, 2866, 2174, 1950, 1884, 1724, 1655, 1593, 1498, 1451, 1388, 1238, 1129, 1042, 805, 765, 699.

$$O \longrightarrow P_2 C \longrightarrow$$

2-(3,3-Difluoro-2-phenylallyl)-*N*-methyl-6-((1-methyl-2-oxo-1,2,3,4-tetrahydroquinolin-7-yl)oxy)-*N*-phenylhexanamide (3-3ua)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA/CH₂Cl₂ (1:1:0.1). 36 mg product was obtained by 45% isolated yield as colorless liquid. 1 H NMR (400 MHz, Chloroform-d) δ 7.19 – 7.10 (m, 6H), 7.02 – 6.94 (m, 3H), 6.87 –6.80 (m, 2H), 6.45 (d, J = 2.3 Hz, 1H), 6.42 (dd, J = 8.1, 2.4 Hz, 1H), 3.84 – 3.72 (m, 2H), 3.25 (s, 3H), 3.13 (s, 3H), 2.75 (t, J = 6.6 Hz, 2H), 2.63 – 2.42 (m, 4H), 2.40 – 2.30 (m, 1H), 1.64 – 1.42 (m, 3H), 1.32 – 1.16 (m, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 174.7, 170.6, 158.6, 154.1 (dd, J = 292.7, 289.4 Hz), 143.5, 141.6, 132.9 (t, J = 3.8 Hz), 129.6, 128.4, 128.1, 128.0 (t, J = 3.3 Hz), 127.6, 127.4, 127.2, 118.3, 107.1, 103.0, 90.3 (dd, J = 21.2, 13.5 Hz), 67.8, 40.0 (t, J = 2.4 Hz), 37.5, 32.1, 31.5, 30.3, 29.6, 29.2, 24.6, 23.8.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -88.97 (d, J = 39.0 Hz, 1F), -89.83 (d, J = 38.4 Hz, 1F). HRMS (ESI-MS) Calcd. For $C_{32}H_{34}F_2N_2O_3Na$ [M+Na]⁺ 555.2430, found: 555.2413.

IR (neat, cm⁻¹): v: 3482, 3032, 2937, 2865, 2244, 1911, 1727, 1654, 1613, 1446, 1355, 1230, 1125, 1066, 1000, 914, 844, 765, 729, 700.

(3-3va)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA/CH₂Cl₂ (2:1:0.1). 71 mg mixture of diastereomers (dr = 1.1:1, determined by 19 F NMR analysis of the mixture) was obtained by 58% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 – 7.11 (m, 6H), 7.00 (d, J = 6.4 Hz, 2H), 6.84 – 6.77 (m, 2H), 5.20 (dt, J = 10.8, 3.6 Hz, 1H), 3.86 – 3.78 (m, 2H), 3.18 – 3.13 (m, 1H), 3.12 (s, 3H), 2.77 (dd, J = 14.4, 4.8 Hz, 1H), 2.61 – 2.54 (m, 1H), 2.45 (ddd, J = 14.6, 8.0, 2.4 Hz, 1H), 2.35 – 2.28 (m, 1H), 1.88 (td, J = 14.6, 4.1 Hz, 1H), 1.84 – 1.75 (m, 2H), 1.61 – 1.20 (m, 22H), 1.13 – 0.95 (m, 4H), 1.06 (s, 3H), 0.91 (s, 3H), 0.85 (s, 3H), 0.83 (s, 6H), 0.71 (s, 3H), 0.64 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 177.6, 174.7, 154.0 (dd, J = 291.6, 288.4 Hz), 143.8, 143.7, 143.5, 132.9 (t, J = 4.1 Hz), 129.6, 128.4, 128.0 (t, J = 3.3 Hz), 127.6, 127.3, 127.2, 122.40, 122.35, 90.3 (dd, J = 21.4, 13.7 Hz), 79.0, 63.90, 63.86, 55.2, 47.6, 46.6, 45.9, 41.7, 41.3, 39.9 – 39.8 (m), 39.3, 38.8, 38.5, 37.5, 37.0, 33.9, 33.1, 32.8, 32.5, 31.53, 31.51, 30.7, 30.3, 28.6, 28.5, 28.1, 27.68, 27.66, 27.2, 25.88, 25.87, 23.71, 23.68, 23.6, 23.4, 23.0, 18.3, 17.0, 15.6, 15.3.

¹⁹F NMR (565 MHz, Chloroform-d) diastereomer 1 (minor) δ -88.99 (d, J = 39.0 Hz, 1F), -89.83 (d, J = 39.0 Hz, 1F), diastereomer 2 (major) -89.01 (d, J = 39.0 Hz, 1.1F), -89.86 (d, J = 39.0 Hz, 1.1F).

HRMS (ESI-MS) Calcd. For C₅₂H₇₁F₂NO₄Na [M+Na]⁺ 834.5243, found: 834.5238.

IR (neat, cm⁻¹): v: 3465, 2940, 2867, 2246, 2092, 1946, 1728, 1649, 1595, 1455, 1382, 1298, 1238, 1167, 1042, 1000, 914, 764, 731, 700.

(3-3wa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (4:1). 42 mg product was obtained by 44% isolated yield as white solid. Melting point: 116.8 – 118.1 °C.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.18 – 7.09 (m, 7H), 6.99 (\approx d, J = 7.9 Hz, 2H), 6.85 – 6.79 (m, 2H), 6.60 (dd, J = 8.4, 2.8 Hz, 1H), 6.53 (d, J = 2.4 Hz, 1H), 3.80 – 3.68 (m, 2H), 3.13 (s, 3H), 2.86 – 2.78 (m, 2H), 2.62 – 2.55 (m, 1H), 2.50 – 2.39 (m, 2H), 2.36 – 2.29 (m, 2H), 2.18 (td, J = 10.8, 4.8 Hz, 1H), 2.06 (dt, J = 18.6, 9.0 Hz, 1H), 2.02 – 1.86 (m, 3H), 1.62 – 1.21 (m, 12H), 0.83 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 220.9, 174.8, 157.1, 154.1 (dd, J = 291.7, 288.3 Hz), 143.5, 137.7, 132.9 (t, J = 4.1 Hz), 131.9, 129.6, 128.4, 128.0 (t, J = 3.2 Hz), 127.6, 127.4, 127.2, 126.3, 114.5, 112.2, 90.4 (dd, J = 21.3, 13.4 Hz), 67.5, 50.4, 48.0, 44.0, 40.0 (t, J = 2.6 Hz), 38.4, 37.5, 35.9, 31.63, 31.61, 30.3, 29.7, 29.2, 26.6, 26.0, 23.9, 21.6, 13.9.

¹⁹F NMR (565 MHz, Chloroform-d) δ -89.00 (d, J = 39.0 Hz, 1F), -89.88 (d, J = 38.4 Hz, 1F). HRMS (ESI-MS) Calcd. For C₄₀H₄₆F₂NO₃ [M+H]⁺ 626.3440, found: 626.3443.

IR (neat, cm⁻¹): v: 3455, 3034, 2927, 2865, 2244, 2068, 1734, 1651, 1597, 1495, 1449, 1278, 1238, 1117, 1056, 913, 764, 730, 698.

(3xa)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA/CH₂Cl₂ (10:1:0.1). 33 mg diastereomers **1** was obtained by 33% isolated yield as white solid. Melting point: 135.7 - 136.4 °C. 18 mg diastereomers **2** was obtained by 18% isolated yield as colorless liquid.

Diastereomer 1 ¹H NMR (400 MHz, Chloroform-d) δ 7.19 – 7.08 (m, 6H), 6.97 – 6.82 (m, 4H), 4.64 (tt, J = 11.6, 4.8 Hz, 1H), 3.15 (s, 3H), 2.51 – 2.45 (m, 2H), 2.43 – 2.33 (m, 1H), 1.95 (s, 3H), 1.87 – 1.57 (m, 8H), 1.50 – 1.23 (m, 9H), 1.01 – 0.69 (m, 9H), 0.85 (s, 3H), 0.54 (s, 3H), 0.25 (d, J = 6.4 Hz, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 175.3, 170.6, 154.1 (dd, J = 291.5, 289.4 Hz), 143.5, 132.8 (t, J = 3.3 Hz), 129.5, 128.3, 128.2 (t, J = 3.1 Hz), 127.5, 127.4, 127.1, 90.4 (dd, J = 21.4, 14.3 Hz), 74.4, 56.8, 56.5, 42.7, 41.9, 40.4, 40.1, 37.9, 37.6, 37.4 (t, J = 2.4 Hz), 35.8, 35.1, 34.6, 34.2, 32.3, 31.1, 28.4, 27.0, 26.7, 26.3, 24.2, 23.3, 21.5, 20.8, 18.7, 12.1.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.93 (d, J = 40.7 Hz, 1H), -90.39 (d, J = 41.8 Hz, 1H). HRMS (ESI-MS) Calcd. For C₄₂H₅₅F₂NO₃Na [M+Na]⁺ 682.4042, found: 682. 4019.

IR (neat, cm $^{-1}$): \tilde{v} : 2931, 2865, 2246, 2119, 1989, 1730, 1652, 1595, 1448, 1383, 1240, 1118, 1025, 912, 761, 730, 699.

Diastereomer 2 ¹H NMR (600 MHz, Chloroform-d) δ 7.26 – 7.22 (m, 2H), 7.21 – 7.19 (m, 1H), 7.14 – 7.07 (m, 5H), 6.62 (broad s, 2H), 4.64 (tt, J = 11.4, 4.8 Hz, 1H), 3.08 (s, 3H), 2.75 – 2.68 (m, 1H), 2.43 – 2.34 (m, 2H), 1.95 (s, 3H), 1.82 – 1.58 (m, 8H), 1.49 – 1.26 (m, 9H), 1.05 – 0.80 (m, 9H), 0.84 (s, 3H), 0.51 (s, 3H), 0.24 (d, J = 6.6 Hz, 3H).

 13 C NMR (151 MHz, Chloroform-d) δ 175.3, 170.6, 153.8 (dd, J = 291.3, 288.4 Hz), 143.5, 133.3 (t, J = 3.8 Hz), 129.3, 128.4, 128.2 (t, J = 3.5 Hz), 127.4, 127.3, 127.2, 90.9 (dd, J = 21.3, 13.1 Hz), 74.4, 57.0, 56.5, 42.8, 41.9, 40.4, 40.1, 39.2, 37.8, 37.4, 35.8, 35.0, 34.6, 33.8, 32.3, 28.8, 28.4, 27.0, 26.6, 26.3, 24.2, 23.3, 21.5, 20.8, 17.8, 12.0.

¹⁹F NMR (565 MHz, Chloroform-*d*) δ -89.00 (d, J = 39.0 Hz, 1F), -90.53 (d, J = 39.6 Hz, 1F). HRMS (ESI-MS) Calcd. For C₄₂H₅₅F₂NO₃Na [M+Na]⁺ 682.4042, found: 682.4039.

IR (neat, cm $^{-1}$): \tilde{v} : 2932, 2863, 2206, 2160, 1983, 1730, 1657, 1595, 1495, 1447, 1381, 1237, 1026, 927, 768, 699.

1'-Methylspiro[cyclohexane-1,3'-indolin]-2'-one (3-4y)

Following procedure A, the crude mixture was purified by silica gel column chromatography with pentane/EA (20:1). 27 mg product was obtained by 83% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.38 (d, J = 7.4 Hz, 1H), 7.20 (t, J = 7.6 Hz, 1H), 6.97 (t, J = 7.6 Hz, 1H), 6.77 (d, J = 7.7 Hz, 1H), 3.13 (s, 3H), 1.90 – 1.83 (m, 2H), 1.80 – 1.73 (m, 2H), 1.72 – 1.61 (m, 3H), 1.60 – 1.52 (m, 1H), 1.52 – 1.45 (m, 2H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 180.7, 142.8, 135.4, 127.4, 123.9, 121.9, 107.9, 47.5, 33.0, 26.1, 25.2, 21.2.

The data is consistent with the literature.³⁰¹

Radical trapping experiment

To a reaction vial equipped with a magnetic stirring bar were added amide **3-1a** (0.15 mmol, 61.4 mg), Ir(ppy)₃ (0.003 mmol, 2 mg) and TEMPO (0.45 mmol, 70.3 mg). The capped vial was evacuated and backfilled with N₂ three times before anhydrous MeCN (1.5 mL), trifluoromethyl arene **3-2a** (0.3 mmol, 45 uL) and triazinane (0.3 mmol, 43 uL) was added. Then the reaction was stirred at room temperature under blue LEDs irradiation (40 W). After 12 h, only a trace amount of **3-3aa** was detected by ¹⁹F NMR of the crude reaction mixture. The crude mixture was further purified by silica gel column chromatography with pentane/EA (4:1). 25 mg TEMPO adduct **3-5a** was obtained by 38% isolated yield as colorless liquid.

¹H NMR (600 MHz, Chloroform-*d*) δ 7.35 – 7.22 (m, 5H), 6.96 (d, J = 8.4 Hz, 2H), 6.71 (d, J = 8.4 Hz, 2H), 4.33 (dd, J = 9.0, 4.8 Hz, 1H), 3.70 (s, 3H), 3.23 (s, 3H), 2.44 (td, J = 13.2, 4.8 Hz, 1H), 2.29 (td, J = 13.2, 4.8 Hz, 1H), 2.08 – 1.99 (m, 1H), 1.86 – 1.78 (m, 1H), 1.52 – 1.35 (m, 3H), 1.29 – 1.23 (m, 3H), 1.09 (s, 3H), 1.01 (s, 3H), 0.97 (s, 3H), 0.58 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 172.2, 157.8, 143.7, 133.9, 129.2, 129.1, 128.4, 127.5, 113.7, 78.4, 60.6, 59.3, 55.3, 40.8, 40.2, 37.8, 33.7, 32.8, 32.4, 30.4, 20.3, 20.1, 17.2.

HRMS (ESI-MS) Calcd. For C₂₇H₃₉N₂O₃[M+H]⁺ 439.2955, found: 439.2961.

IR (neat, cm $^{-1}$): \tilde{v} : 3479, 2928, 2324, 2080, 1883, 1739, 1662, 1509, 1458, 1383, 1245, 1126, 1035, 989, 825, 771, 699.

Stern-Volmer fluorescence studies

The Agilent Cary Eclipse Fluorescence Spectrometer is used to conduct Stern-Volmer fluorescence quenching analysis. The following parameters were employed: excitation wavelength = 377 nm, emission wavelength = 400 nm, excitation slit width = 5 nm, emission slit width = 10 nm, scan rate = 600 nm/min, averaging time = 0.1s, data interval = 1 nm. The samples were measured at Hellma fluorescence QS quartz cuvette (chamber volume = 1.4 mL, light path = 10×4 mm) with a closed screw cap and silicone seal. All the solutions were prepared with dry degassed MeCN. The concentration of photocatalyst Ir(ppy)₃ is 2×10 -6 M in MeCN, then by adding certain amount of a solution of quencher to Ir(ppy)₃ solution, the samples with different concentrations were obtained,

and their fluorescence spectra were collected immediately with forementioned parameters. I_0 is the fluorescence intensity without quencher, I is the fluorescence intensity with quencher.

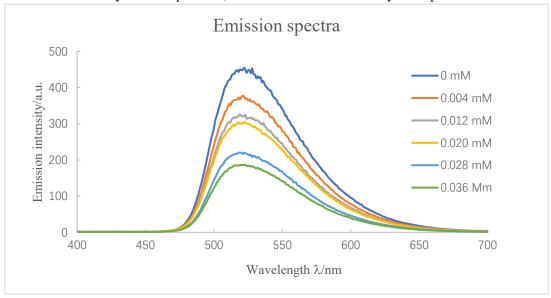


Figure 3-S1: Emission spectra of $Ir(ppy)_3$ with increasing concentrations of triazinane (The concentration of $Ir(ppy)_3$ is 2×10^{-6} M in MeCN)

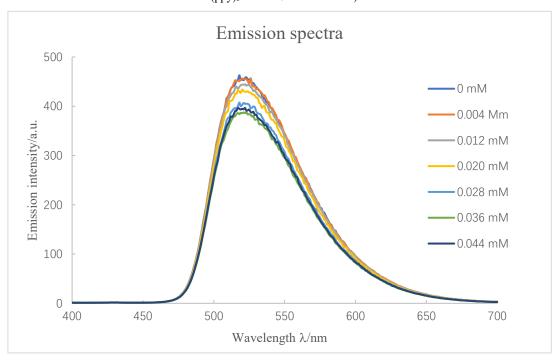


Figure 3-S2: Emission spectra of $Ir(ppy)_3$ with increasing concentrations of amide 3-1f (The concentration of $Ir(ppy)_3$ is 2×10^{-6} M in MeCN)

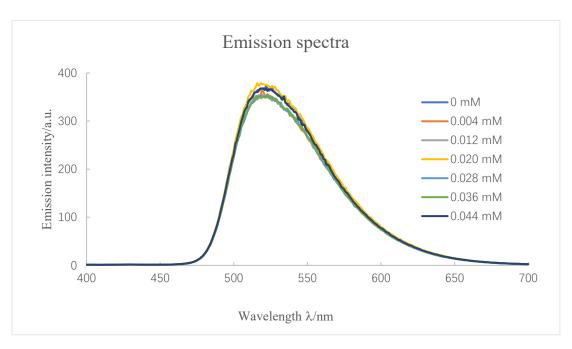


Figure 3-S3: Emission spectra of $Ir(ppy)_3$ with increasing concentrations of trifluoromethyl alkene 3-2a (The concentration of $Ir(ppy)_3$ is 2×10^{-6} M in MeCN)

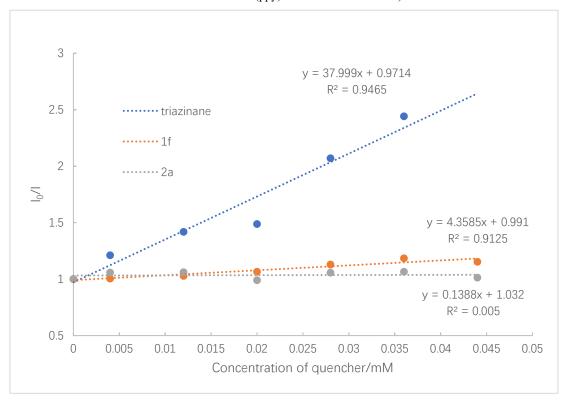


Figure 3-S4: Stern-Volmer plot of Ir(ppy)₃ with increasing concentrations of triazinane, amide 3-1f, and trifluoromethyl arene 3-2a.

5 List of abbreviation

| aq aqueous Ar aryl Binap 1.1'-binaphthyl-2.2'-diphemyl phosphine Boc #ert-butyloxycarbonyl BDE bond dissociation energy Bpin pinacol boronic ester br broad Bu butyl Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgHz)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diinsopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DMSO dimethyl actante EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl Hz Hertz | Ac | acetyl |
|--|---|--|
| Binap | aq | aqueous |
| BDE bond dissociation energy Bpin pinacol boronic ester br broad Bu butyl Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichlorothane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DMA N,N-diimethylacetamide DMA N,N-diimethylacetamide DMSO dimethyl actane EI clectron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorescent HRMS high resolution mass spectroscopy | Ar | aryl |
| BDE bond dissociation energy Bpin pinacol boronic ester br broad Bu butyl Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DDPA N,N-diinethylglomine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DMB 1,2-dimethoxyethyl ether DMA N,N-dimethylsuffoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorence HEMS high resolution mass spectroscopy | Binap | 1.1'-binaphthyl-2.2'-diphemyl phosphine |
| Bpin broad Bu butyl Bn benzyl BNAH l-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE l,2-dichloroethane DDPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME l,2-dimethoxyethane DMA N,N-dimethylsufoxide dr diasteromeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acctate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorescent Dintyl henry HE Hantzsch ester HRMS high resolution mass spectroscopy | Boc | tert-butyloxycarbonyl |
| br broad Bu butyl Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH ₂)Cl ₂ dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastercomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | BDE | bond dissociation energy |
| Bu butyl Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Bpin | pinacol boronic ester |
| Bn benzyl BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine *C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorescent lamp Dictionario distorescent lamp L-2-dimethoxyethyl glyoximato)cobalt(III) DX dictate EI electrospray ionization ET energy transfer Et ethyl Hantzsch ester HRMS high resolution mass spectroscopy | br | broad |
| BNAH 1-benzyl-1,4-dihydronicotinamide Bz benzoyl bpy 2,2'-bipyridine C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Bu | butyl |
| Bz benzoyl bpy 2,2'-bipyridine C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastercomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Bn | benzyl |
| bpy 2,2'-bipyridine C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | BNAH | 1-benzyl-1,4-dihydronicotinamide |
| °C centigrade Calcd calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dinastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Bz | benzoyl |
| Calcul calculated Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH ₂)Cl ₂ dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | bpy | 2,2'-bipyridine |
| Cbz benzyloxycarbonyl CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | °C | centigrade |
| CFL compact fluorescent lamp Cy cyclohexyl Co(dmgH)(dmgH2)Cl2 dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Calcd | calculated |
| Cy cyclohexyl Co(dmgH)(dmgH ₂)Cl ₂ dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Cbz | benzyloxycarbonyl |
| Co(dmgH)(dmgH ₂)Cl ₂ dichlorobis(dimethylglyoximato)cobalt(III) DCE 1,2-dichloroethane 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. estyl electrospray ionization ET energy transfer ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | CFL | compact fluorescent lamp |
| DCE 1,2-dichloroethane DCA 9,10-dicyanoanthracene DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Су | cyclohexyl |
| DCA DIPEA N,N-diisopropylethylamine diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Co(dmgH)(dmgH ₂)Cl ₂ | dichlorobis(dimethylglyoximato)cobalt(III) |
| DIPEA N,N-diisopropylethylamine | DCE | 1,2-dichloroethane |
| diglyme bis(2-methoxyethyl) ether DMF N,N-dimethylformamide DME 1,2-dimethoxyethane DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | DCA | 9,10-dicyanoanthracene |
| DMF DME 1,2-dimethylformamide DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | DIPEA | N,N-diisopropylethylamine |
| DME DMA N,N-dimethylacetamide DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | diglyme | bis(2-methoxyethyl) ether |
| DMA N,N-dimethylacetamide dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | DMF | N,N-dimethylformamide |
| DMSO dimethyl sulfoxide dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester high resolution mass spectroscopy | DME | 1,2-dimethoxyethane |
| dr diastereomeric excess dtbbpy 4,4'-di-tert-butyl-2,2'-dipyridyl EA ethyl acetate EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | DMA | N,N-dimethylacetamide |
| dtbbpy4,4'-di-tert-butyl-2,2'-dipyridylEAethyl acetateEIelectron ionizationequiv.equivalentESIelectrospray ionizationETenergy transferEtethylFmocfluorenylmethyloxycarbonylhhourHEHantzsch esterHRMShigh resolution mass spectroscopy | DMSO | dimethyl sulfoxide |
| dtbbpy4,4'-di-tert-butyl-2,2'-dipyridylEAethyl acetateEIelectron ionizationequiv.equivalentESIelectrospray ionizationETenergy transferEtethylFmocfluorenylmethyloxycarbonylhhourHEHantzsch esterHRMShigh resolution mass spectroscopy | dr | diastereomeric excess |
| EI electron ionization equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | dtbbpy | |
| equiv. equivalent ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | EA | ethyl acetate |
| ESI electrospray ionization ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | EI | electron ionization |
| ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | equiv. | equivalent |
| ET energy transfer Et ethyl Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | | |
| Fmoc fluorenylmethyloxycarbonyl h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | ET | |
| h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Et | ethyl |
| h hour HE Hantzsch ester HRMS high resolution mass spectroscopy | Fmoc | fluorenylmethyloxycarbonyl |
| HRMS high resolution mass spectroscopy | h | |
| | НЕ | Hantzsch ester |
| | HRMS | high resolution mass spectroscopy |
| | Hz | |

| IR | infrared spectroscopy |
|----------------------|--|
| <i>i</i> -Pr | isopropyl |
| Incl. | include |
| J | coupling constant (in NMR spectroscopy) |
| Me | methyl |
| mol | mole |
| MS | mass spectroscopy |
| N.D. | not detected |
| NHC | N-heteroccyclic carbene |
| Ni(cod) ₂ | bis(cyclooctadiene)nickel(0) |
| Ni(COD)(DQ) | bis(1,5-cyclooctadiene)(duroquinone) nickel(0) |
| NMR | nuclear magnetic resonance |
| ox | oxidized |
| PCET | proton-coupled electron transfer |
| Ph | phenyl |
| phen | phenanthroline |
| Ру | pyridyl |
| рру | 2-phenylpyridine |
| Piv | pivaloyl |
| rt | room temperature |
| Ref. | reference |
| SET | single electron transfer |
| t-Bu | tert-butyl |
| TBAI | tetrabutylammonium iodide |
| THF | tetrahydrofuran |
| TMS | trimethylsilyl |
| TFA | trifluoroacetic acid |
| TfOH | trifluoromethanesulfonic acid |
| TFE | trifluoroethanol |
| Tol | toluene |
| Ts | p-toluenesulfonyl |
| δ | chemical shift |

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7 Acknowledgements

Firstly, I would like to express my deepest gratitude to my supervisor, Prof. Dr. Frederic W. Patureau. His unwavering support, insightful guidance, and profound expertise have been invaluable throughout my doctoral journey. From the very beginning, he has not only provided me with the academic freedom to explore my research interests but also continuously challenged and inspired me to think critically and push the boundaries of my work. His patience, encouragement, and belief in my abilities have played a crucial role in shaping both my scientific development and personal growth. I am truly grateful for the opportunity to be his student.

I would also like to extend my thanks to my colleagues who have supported me in countless ways during my PhD. I sincerely appreciate the generous help and collaboration from Jiaxiang Xiang, Yue Zhao, Raolin Huang, Xinben Wang, Yun Yang, Fang Xiao, Long Huang, Vinzenz Thönnißen, Alina Paffen, Alija Spahic, Shiny Nandi, Pooja Vemuri, Alexander Schacht, Leander Bruck and Melissa Hohenadel. Their insightful discussions, technical assistance, and, most importantly, their friendship have made this journey not only more manageable but also much more enjoyable. The time spent working together, discussing science, and sharing laughter in the lab is something I will always cherish.

I am also deeply grateful to the China Scholarship Council (CSC) for providing me with the predoctoral stipend that made it possible for me to pursue my research at RWTH Aachen University. Their financial support has been instrumental in allowing me to focus entirely on my studies and research.

Finally, I would like to thank my family and friends for their endless love, patience, and encouragement. Their unwavering belief in me has been a source of strength through all the challenges and triumphs of this journey. Without their support, this achievement would not have been possible.

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